ANTICANCER-IMMUNE RESPONSE TOWARDS BREAST CANCER CELL LINES MDA-MB-231 AND TERATOGENIC ASSESSMENT OF Pereskia bleo LEAVES

TAIF KAREEM KHALAF DALFI

UNIVERSITI SAINS MALAYSIA

TINDAK BALAS ANTIKANSER-IMUN TERHADAP SEL KANSER PAYUDARA MDA-MB-231 DAN PENILAIAN TERATOGENIK DAUN *Pereskia bleo*

by

TAIF KAREEM KHALF DALFI

Thesis submitted in fulfilment of the requirements for the degree of Doctor of Philosophy

August 2024

ACKNOWLEDGEMENT

First, I am grateful to Allah for giving me the endurance to accomplish my study despite all the difficulties and challenging circumstances. I want to express my deepest gratitude to my supervisor, Dr. Norzila Ismail, for her exceptional guidance and unwavering support throughout my academic journey. Her role as a supervisor has been instrumental in shaping my experience and success during this period of study. Her expertise and insightful feedback have been invaluable, pushing me to grow academically and personally. Beyond the professional realm, her encouragement and friendly demeanour have made the challenges of academia more manageable. I am truly fortunate to have had her mentorship and friendship during this time, and I appreciate the countless ways she has helped me navigate the complexities of my studies. I also would like to express my gratitude to my co-supervisor, Dr. Wan Ezumi Mohd Fuad for her patience, time invested in me, great support and continuous guidance throughout the course of my study. I would like to acknowledge the financial support for this project from Fundamental Research Grant Scheme (FRGS/1/2019/WAB11/USM/03/1) from the Ministry of Higher Education Malaysia. I am deeply indebted to my family for their never-ending assistance and comprehension, as well as for pushing me to believe in myself in times of uncertainty and challenges. I am deeply thankful to all members of the laboratory staff from the Pharmacology Department, Central Research Laboratory (CRL), and Animal Research And Service Center (ARASC) for assisting me, and last but not least, to the School of medical sciences for all the facilities and support for my Ph.D. research.

TABLE OF CONTENTS

ACK	NOWLEI	DGEMENT	ii
TABLE OF CONTENTSiii			
LIST	OF TAB	LES	ix
LIST	OF FIGU	J RES	xii
LIST	OF SYM	BOLS	xxi
LIST	OF ABB	REVIATIONS	xxii
LIST	OF APPI	ENDICES	xxiv
ABST	FRAK		XXV
ABST	FRACT		. xxviii
CHA	PTER 1	INTRODUCTION	1
1.1	Backgro	und of the study	1
1.2	Problem	statement and rationale of the study	3
1.3	Objectiv	e of study	4
	1.3.1	General objective	4
	1.3.2	Main objective of the study	4
CHA	PTER 2	LITERATURE REVIEW	6
2.1	Medicin	al plants	6
	2.1.1	Anti-cancer activities of medicinal plants	8
	2.1.2	Immunomodulatory effects by medicinal plants	13
	2.1.3	Phytochemicals in medicinal plants	13
2.2	Pereskia	a bleo (Cactaceae)	15
	2.2.1	General description of Pereskia bleo	16
	2.2.2	Classification of <i>P. bleo</i>	17
	2.2.3	Medicinal uses of <i>P. bleo</i>	17
	2.2.4	Chemical constituents of <i>P. bleo</i>	18

	2.2.5	Bioactivity of <i>P. bleo</i>	20
2.3	Cancer.		21
	2.3.1	Cancer and prevalence	21
	2.3.2	Breast cancer	25
	2.3.3	Breast cancer therapy modalities	26
		2.3.3(a) Surgery	26
		2.3.3(b) Chemotherapy	27
		2.3.3(c) Radiotherapy	29
		2.3.3(d) Immunotherapy	31
2.4	Role of	common immune cells in cancer	40
	2.4.1	Natural killer cells (NK cells)	40
	2.4.2	The categorisation of NK cell receptors	43
	2.4.3	NK cell killing mechanism	47
	2.4.4	NK cells as immunotherapy for cancer	48
2.5	Apoptos	sis	50
	2.5.1	Morphological and biochemical alteration of apoptotic cells in apoptosis	51
		2.5.1(a) Morphological condition in apoptosis	51
		2.5.1(b) Biochemical process in apoptosis	52
	2.5.2	Apoptosis pathways	53
		2.5.2(a) The extrinsic death receptor pathway	53
		2.5.2(b) The intrinsic or mitochondrial pathway	54
	2.5.3	Tumour suppressing protein p53	57
	2.5.4	The perforin and granzyme B-mediated killing pathways	60
	2.5.5	Targeting Apoptosis and cancer treatment	61
2.6	General	descriptive of teratogenicity	62
	2.6.1	Embryonic periods and teratogenicity in foetal development	63
	2.6.2	Medicinal plants during trimesters of pregnancy	65

	2.6.3	Teratogenic effects of Anti-cancer drugs in foetal development	73
CHAI	PTER 3	MATERIAL AND METHODS	
3.1	Overviev	v of the study	
3.2	Breast ce	ll lines	84
3.3	Plant mat	terials	84
3.4	Preparati	on of the extracts	84
3.5	Preparati	on of stock solution	85
3.6	Cell lines	s and culture	86
3.7	Anti-prol	liferative assay	86
3.8	Morphole	ogy of cell death analysis	88
	3.8.1	Bright-field inverted microscopy	. 88
	3.8.2	Fluorescence microscopy	89
3.9	Apoptosi	s assay	89
3.10	Cell cycl	e assay	90
3.11	Protein expression (Bax, caspase-3, p53, and Bcl-2)		. 91
3.12	Blood incubation		. 93
	3.12.1	Serial dilution	. 93
	3.12.2	Ethical approval	. 94
	3.12.3	Location of blood sampling	. 94
	3.12.4	Items used in the blood collection	. 94
	3.12.5	Blood Sampling for cytokine measurement to determine the optimal concentration of MEPB leaves	94
	3.12.6	Incubation of blood sample	. 95
	3.12.7	Enzyme linked immunosorbent assay (ELISA)	. 96
3.13	NK cell s	source	. 97
3.14	Isolation	of Natural killer (NK) cells	. 98
3.15	Purificati	on of NK cells	99

3.16	Co-culture of human NK cells with MDA-MB-231 cell lines 100		
	3.16.1	Apoptosis assay	101
	3.16.2	ELISA (IFN-γ, perforin, and granzyme B)	102
3.17	Toxicity	and teratogenicity experiment	104
	3.17.1	Experimental Design	104
		3.17.1(a) Preparation of MEPB leaves	104
		3.17.1(b) Dosage preparation	104
		3.17.1(c) Guidelines for experiment	104
		3.17.1(d) Ethical approval for animal use	104
		3.17.1(e) Source of experimental animal	104
		3.17.1(f) Housing environment for Sprague-Dawley rats	105
	3.17.2	Experimental protocol	105
		3.17.2(a) Female toxicity experiment on virgin rat	105
		3.17.2(b) Teratogenicity assessment experiment	106
	3.17.3	Histological assessment	113
3.18	Statistic	al analysis	113
CHA	PTER 4	RESULTS	115
4.1	Inhibitor	ry concentration (IC ₅₀) of <i>P. bleo</i> leaves extracts	115
4.2	The mor	phological features of MDA-MB-231 cells	120
	4.2.1	Bright field inverted microscopy	120
	4.2.2	Fluorescence microscopic assessment of an apoptotic body	122
4.3	Annexin	V/PI assay detected apoptotic MDA-MB-231 cells	124
4.4	MEPB l	eaves induced cell cycle arrest in the G1 phase	129
4.5	Protein e	expression data	132
4.6	Cytokine	e expression outcomes in healthy blood samples	138
4.7	Purificat	tion of NK cells	141
4.8	NK cell counts in healthy women vs breast cancer patient		142

4.9	Apoptos	is assay data in a co-culture experiment143
4.10		perforin, and granzyme B expression data in a co-culture ent
4.11	Toxicity	and teratogenicity results
	4.11.1	Toxicity assessment on virgin rats152
		4.11.1(a) Assessment of oestrous cycle
		4.11.1(b) Assessment of body weight in rats
		4.11.1(c) Measurement of clinical signs or symptoms164
		4.11.1(d) Assessment of absolute body weights
		4.11.1(e) Histopathological observation
	4.11.2	Assessment of Teratogenicity (pregnancy outcomes)174
CHAI	PTER 5	DISCUSSION
5.1	Cytotoxi	icity of <i>P. bleo</i> leaves extracts
5.2	Morphol	logical features of MDA-MB-231 breast cancer cells
	5.2.1	Bright-field inverted microscopy
	5.2.2	Fluorescence microscopy
5.3	Annexin	VFITC/PI assay detected apoptosis progression
5.4	MEPB le	eaves induced MDA-MB-231 cell cycle arrest in G phase
5.5		eaves induce pro-apoptotic proteins and block anti-apoptotic protein A-MB-231 breast cancer cells
5.6		leaves induce cytokines and cytotoxic granules in healthy blood
	5.6.1	IFN-γ expression
	5.6.2	IL-8 expression
	5.6.3	IL-10 expression
	5.6.4	IL-12 expression
	5.6.5	IL-18 expression
	5.6.6	Perforin & Granzyme B expression

5.7	Purification and count of NK cells	210
5.8	Natural killer cell-mediated cytotoxicity2	211
5.9	Toxicity and Teratogenicity	215
CHAP	TER 6 CONCLUSION AND FUTURE RECOMMENDATIONS 2	27
6.1	Conclusion	227
6.2	Recommendations	228
REFERENCES		
APPENDICES		
LIST OF PUBLICATIONS		

LIST OF TABLES

Page

Table 2.1:	List of bioactive compounds of plants that have anti-cancer
	activities7
Table 2.2:	Plants derived anti-cancer drugs9
Table 2.3:	Type of medicinal plants that have anti-cancer activities towards various cancers
T 11 A 4	
Table 2.4:	<i>P. bleo</i> categorization17
Table 2.5:	Traditional usage and methods of preparation of <i>P. bleo.</i>
Table 2.6:	Components of <i>P. bleo</i> leaves19
Table 2.7:	The incidence rate of cancer in both sexes for all ages24
Table 2.8:	CAR-T cell therapy in triple-negative breast cancer in vivo and in
	<i>vitro</i> studies
Table 2.9:	NK cell activating and inhibitory receptors in humans46
Table 2.10:	Prevalence rate (%) of pregnancies that use herbal medicine
Table 2.11:	Listed medicinal plants utilised during pregnancy and classified
	them into two groups: safe or nan-harmful (A) and harmful (B)69
Table 2.12:	Teratogenic potential of anti-cancer drugs in trimesters74
Table 3.1:	Type of breast cell lines used in the experiments
Table 3.2:	Serial concentration of <i>P. bleo</i> leaves extracts
Table 3.3:	The criteria of healthy donors95
Table 3.4:	Selection criteria for healthy and breast cancer patient donors97
Table 3.5:	Experimental design for co-culture instruction101
Table 3.6:	Shows an ELISA protocol was used to assess the level of IFN- γ ,
	perforin, and granzyme B in the co-cultured experiment for healthy
	and breast cancer samples103

Table 4.1:	The classification of the IC50 value of crude extract (Geran <i>et al.</i> ,1972)
Table 4.2:	The IC ₅₀ value (µg/ml) obtained from the ELISA reader showing the cytotoxic activity of three extracts of <i>P. bleo</i> leaves against normal (MCF-10A) and breast cancer cells (MDA-MD-231) after 72 hrs of incubation
Table 4.3:	Apoptotic progression of MDA-MB-231 cells after treatment with MEPB leaves for 24, 48, and 72 hrs
Table 4.4:	MEPB leaves induced cell cycle arrest in MDA-MB-231 cells for 24, 48, and 72 hrs of incubation compared to the untreated cells. After the incubation, DNA content in MDA-MB-231 cells in the G1, S, and G2 phases was determined by flow cytometry131
Table 4.5:	The percentage of Bax, caspase-3, p53, and Bcl-2 protein expression in the experimental groups
Table 4.6:	The ELISA was used to assess cytokine expression (pg/mL) in healthy blood samples after 20 hrs of incubation140
Table 4.7:	Apoptotic progression of MDA-MB-231 breast cancer cells in all the experimental groups
Table 4.8:	Total cytokine expression in co-culture experiment from healthy women $(n = 3)$ and breast cancer patients $(n = 3)$ 150
Table 4.9:	The average length for one and two durations of oestrous cycle in female virgin rats treated with MEPB leaves at doses of 250, 500, and 1000 mg/kg/day compared to the control group (DW)157
Table 4.10:	The average length of the one oestrous cycle phase (hours) in virgin rats for control (DW) and groups treated with MEPB leaves at doses of 250, 500, and 1000 mg/kg/day159
Table 4.11:	The average body weight of female virgin rats treated with MEPB leaves at 250, 500, and 1000 mg/kg/day compared to the control rats (DW)

- Table 4.13:Toxicity evaluation of MEPB leaves at doses of 250, 500, and 1000mg/kg/day in female virgin rats (treatment and control groups)......164
- Table 4.14:Normal absolute organ weights of all dams' internal organs after
29 days (10 days before pregnancy and 19 days during pregnancy)
with various doses of MEPB leaves (250, 500, and 1000
mg/kg/day) compared to the control group (DW)......165
- Table 4.15:The evidence from pregnant rats treated with various doses ofMEPB leaves demonstrated no effect on pregnancy outcomes. 174

- Table 4.18:The measurement parameter data revealed no foetal abnormalities
observed across the dosing spectrum for 19 days of treatment when
compared to the control group (DW).179

LIST OF FIGURES

Page

Figure 2.1:	A: The stem of <i>P. bleo</i> , B. leaves and the orangish-red flowers of
	<i>P. bleo</i> 17
Figure 2.2:	Incidence (A) and mortality rate of cancer (B) worldwide, 202022
Figure 2.3:	NK cells development42
Figure 2.4:	Morphology of apoptotic cells in apoptosis (Smith et al., 2017)53
Figure 2.5:	The extrinsic and intrinsic pathways (Loreto et al., 2014)55
Figure 2.6:	The Bcl-2 proteins govern apoptosis via mitochondrial pathways57
Figure 3.1:	The study overview
Figure 3.2:	The serial dilution of extract concentrations85
Figure 3.3:	A visual representation of the staining cell distribution in each
	quadrant90
Figure 3.4:	The process of serial dilution of MEPB leaves94
Figure 3.5:	Blood sample grouping
Figure 3.6:	Isolated human peripheral blood mononuclear cells (PBMC) from
	healthy women and breast cancer patients. A: layered diluted
	blood. B: After centrifugation, a test-tube shows four unique layers
	of colour, including the plasma layer, the PBMCs, the LSM phase
	or layer, and the erythrocyte phase, respectively
Figure 3.7:	A diagrammatic illustration of the MDA-MB231 stained cell
	distribution in each quadrant102
Figure 3.8:	Pregnancy was confirmed by the presence of sperms (white
	arrows) in the vaginal smear after one day of mating107
Figure 3.9:	Anatomical dissection of the dam109
Figure 3.10:	
	Ovary and oviduct were extracted109

Figure 3.12:	Collected foetus from the horn110
Figure 3.13:	Photograph showing the parameters used in the current experiment. A: CRL of foetuses was recorded. B: The length of the upper and lower limbs was subjected. C: Length and width of the tongue were recorded. D: Length and width of the eyes, external ear, and abdominal circumference were recorded111
Figure 3.14:	Visible (A) and microscopic (B) examination of foetuses (yellow circle)
Figure 4.1:	Graph illustrating viability (%) of MDA-MB231 cells following 72 hrs of treatment with three extracts of <i>P. bleo</i> leaves (Hexane, ethyl acetate, and methanolic extracts). The number of independent trials with replicates used to generate the data was three $(n = 3)$, which was expressed as mean \pm SEM
Figure 4.2:	Graph displaying viability (%) of MDA-MB-231 and MCF-10A cells following 72 hrs of treatment with Tamoxifen. The number of independent trials with replicates used to generate the data was three (n = 3), which was expressed as mean \pm SEM118
Figure 4.3:	Graph representing viability (%) of MCF-10A cells following 72 hrs of treatment with three extracts of <i>P. bleo</i> leaves (Hexane, ethyl acetate, and methanolic extracts). The number of independent trials with replicates used to generate the data was three ($n = 3$), which was expressed as mean \pm SEM
Figure 4.4:	The morphological alterations (arrows) of MDA-MB-231 cells following incubation for 24, 48, and 72 hrs, which were observed with an Olympus CKX53 inverted microscope ($40\times$ magnification). A: Untreated MDA-MB-231 cells, B: MDA-MB- 231 cells treated with 64.57 µg/mL of MEPB leaves, and C: MDA- MB-231 cells treated with 6.17 µg/mL of tamoxifen
Figure 4.5:	Hoechst 33258 staining shows the apoptotic progression of MDA- MB-231 breast cancer cells (arrows) in untreated (A) and treatment

with MEPB leaves for 24 (B), 48 (C), and 72 hrs (D). The images

were captured with an Olympus BX41 fluorescent microscope at 40× magnification and a wavelength of 330–385 nm......123

- Figure 4.10: The percentage of Bax protein expression in MDA-MB-231 cells treated with MEPB leaves for 24, 48, and 72 hrs, respectively, compared to the untreated cells. The data is presented as the mean

 \pm SEM of three triplicate trials (n = 3). ****P < 0.0001, indicate significant differences between the untreated and treated groups. ..133

- Figure 4.16: The flow cytometry illustration of the apoptotic progression of MDA-MB-231 cells in all the healthy donor experimental groups after 24 hrs of incubation. Group 1: MDA-MB-231 cells cultured alone, Group 2: MDA-MB-231 cells cultured and treated with MEPB at 7.5 μg/mL, Group 3: MDA-MB-231 cells co-cultured

- Figure 4.18: The flow cytometry showed apoptotic progression of MDA-MB-231 cells in all breast cancer patient donor experimental groups after 24 hrs of incubation. Group 1: MDA-MB-231 cells cultured alone, Group 2: MDA-MB-231 cells cultured and treated with MEPB at 7.5 µg/mL, Group 3: MDA-MB-231 cells co-cultured with NK cells isolated from breast cancer patients, and Group 4: MDA-MB-231 cells co-cultured with NK cells isolated from breast cancer patients and treated with MEPB leaves at 7.5 µg/ml. Dot plots showed the distribution of stained MDA-MB-231 cells throughout the four quadrants (Q1: Represents the number of viable cells; Q2 and Q3: Represent the early and late apoptotic cells, respectively; and Q4: Represents the number of necrotic cells).
- Figure 4.19: The proportion of apoptotic cell stages in all breast cancer patient donor experimental groups after 24 hrs of incubation. Group1: MDA-MB-231 cells cultured alone, Group 2: MDA-MB-231 cells

- Figure 4.20: Comparison level of IFN-γ expression (A), perforin expression (B), and granzyme B expression (C) (pg/mL) in three independent healthy donors and three breast cancer patient donors......151
- Figure 4.22: Unstained vaginal smears from female virgin rats were analysed under light microscopy (10× magnification) to determine the normal oestrous phase in the treated groups with MEPB leaves at doses of 250, 500, and 1000 mg/kg/day for 10 days. Nucleated epithelial cells (red arrows) and few nucleated cornified epithelial cells (orange arrows) were present during proestrous (A), cornified epithelial cells (green arrows) were present during oestrous (B), leukocytes (blue arrows) and cornified epithelial cells (purple arrows) were present during metoestrous (C), and dioestrous consists solely of leukocytes (yellow arrows) (D)......154

- Figure 4.29: Dam liver histology following staining with haematoxylin and eosin (H&E) at 20× magnification. The H&E showed the normal structure of hepatocytes (yellow arrows) in all the rats in the control group receiving DW (A) and rats treated with MEPB leaves at 250 (B), 500 (C), and 1000 (D) mg/kg/day......171

- Figure 4.32: Microscopic and visible examination indicated that all the foetuses in the rats treated with MEPB leaves at doses of 250 mg/kg/day (Group 2), 500 mg/kg/day (Group 3), and 1000 mg/kg/day (Group

LIST OF SYMBOLS

°C	Degree Celsius
min	Minutes
mL	millilitre
cm	centimeter
%	Percentage
α	Alpha
γ	Gamma
g	Gram
mg	Milligrams
μg	Micrograms
/	Per, and, or
mg/mL	Milligrams/ millilitre
µg/mL	Micrograms/ millilitre
μl	Microliter
mm	Millimeter
nm	Nanometer
pg/mL	Picograms/ millilitre
no	Number
n	Number
Kg	Kilogram
mg/kg	Milligrams/Kilogram
+	Positive
-	Negative
VS	versus
Q	Quarter
kD	Kilo Dalton
hrs	Hours

LIST OF ABBREVIATIONS

ADCC	Antibody-Dependent Cell Cytotoxicity
AIF	Apoptosis-inducing factor
Apaf-1	Apoptotic protease activation factor-1
APCs	Antigen presentation cells
API	Apoptotic protein inhibitors
Bcl-2	B-cell lymphoma 2
BSA	Bovine serum albumin
BW	Body weight
CO ₂	Carbon dioxide
CRL	Crown–rump length
Cyto C	Cytochrome C
DISC	Death inducing signalling complex protein
DMEM	Dulbecco's Modified Eagles Medium
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
DRs	Death receptors
DW	Distill water
ELISA	Eenzyme-linked immunosorbent assay
End G	Endonuclease G
ER	Estrogen receptor
FADD	Fas-associated death domain
FasL	Fas Ligand
FBS	Foetal bovine serum
FDA	Food and Drug Adminstration
G1 phase	Gap phase 1
G2 phase	Gap phase 2
GD	Gestation day
GM-CSF	Granulocyte-macrophage colony-stimulating factor
HtrA2	High-temperture requirement protein A2
IAPs	Inhibitor of apoptosis (IAP) proteins
IC50	Half maximal inhibitory concentration
ICAD	Caspase-activated DNas
IFN-γ	Interferon gamma
ILs	Interlukines
IP	Intraperitoneal
ITIMs	Immunoreceptors tyrosine based inhibitory motifs
KIR	Killer cell immunoglobulin-like receptors
LIRs	Leukocyte inhibitory receptors
Log 10	Logarithm with base 10
LSM	Lymphocytes Separation Media
MDM2	Mouse double minute protein 2
MEPB	Methanol extract of Pereskia bleo
MHC	Major histocompatibility complex
MOMP	Mitochondrial Outer Membrane Permeabilization
NK cells	Natural killer cells
NKT	Natural Killer T

OD	Optical density
OECD	Organisation for Economic Cooperation and Development
P. bleo	Pereskia bleo
P53	Tumour suppressor protein
PBMCs	Peripheral blood mononuclear cells
PBS	Phosphate buffer saline
PD-1	Programmed cell death 1
PD-L1	Programmed cell death ligand 1
PHA	Phythaemagglutinin
PI	Propidium iodide
PS	Phosphatidylserine
RNA	Ribonucleic acid
ROI	Reactive oxygen intermediates
ROS	Reactive oxygen species
S phase	Synthesis Phase
SEM	Stander error mean
Smac	Second mitochondrial-dervied activator of caspases
tBID	Turncated BID
TME	Tumour microenviroment
TRADD	TNF receptor-associated death domain
TRAIL	TNF-related apoptosis-inducing ligand
VS	Versus
WHO	World Health Organization

LIST OF APPENDICES

Appendix A	List of preparation medium
Appendix B	Blood incubation experiment
Appendix C	Reagent preparation for ELISA
Appendix D	Representative typical curve of human IFN- γ ELISA in healthy blood samples
Appendix D	Representative typical curve of human IL-12 ELISA in healthy
Appendix E	blood samples
	Representative typical curve of human IL-18 ELISA in healthy
Appendix F	blood samples
Appendix G	Representative typical curve of human IL-8 ELISA in healthy blood samples
Appendix O	Representative typical curve of human IL-10 ELISA in healthy
Appendix H	blood samples
	Representative typical curve of human perforin ELISA in healthy
Appendix I	blood samples
Appendix J	Representative typical curve of human granzyme B ELISA in healthy blood sample
Appendix 3	Representative typical curve of human IFN- γ ELISA in co-culture
Appendix K	experiment
	Representative typical curve of human perforin ELISA in co-
Appendix L	culture experiment
Appendix M	Representative typical curve of human granzyme B ELISA in co- culture experiment
Appendix M	culture experiment
Appendix N	List of chemical/reagents/kits

TINDAK BALAS ANTIKANSER-IMUN TERHADAP SEL KANSER PAYUDARA MDA-MB-231 DAN PENILAIAN TERATOGENIK DAUN Pereskia bleo

ABSTRAK

Rawatan konvensional bagi kanser payudara, terutamanya terapi radiasi dan kemoterapi, memberi kesan sampingan kepada pesakit. Oleh itu, fokus di peringkat global telah meningkat dalam mencari kaedah rawatan yang tidak toksik dan bersifat sitoselektif, termasuk kajian ke atas herba. Pelbagai jenis herba menunjukkan bioaktiviti yang sangat baik walau bagaimanapun terdapat juga herba yang bersifat toksik dan teratogenik pada dos-dos tertentu. Kajian ini dijalankan untuk menilai sifat anti-kanser daun *P. bleo* dari segi keupayaannya untuk mengaruh apoptosis dan untuk menilai kebolehannya mengaruh tindakbalas imun anti-kanser dengan meningkatkan ketoksikan sel Pembunuh Semulajadi (sel NK) terhadap sel kanser payudara MDA-MB-231, serta menilai ketoksikan dan keteratogenannya dalam model haiwan. Ekstrak heksana, etil asetat dan metanol daun P. bleo telah diuji ketoksikannya ke atas sel normal MCF-10A dan MDA-MB-231 melalui ujian MTT. Esktrak yang paling aktif ialah metanol, maka metanol digunakan untuk eksperimen yang seterusnya. Annexin V/PI dan analisis sitometri aliran digunakan untuk meneroka keupayaan ekstrak metanol daun P. bleo (MEPB) dalam mengaruh apoptosis, penyekatan kitaran sel dan pengekspresan protein apoptotik. Ujian imunosorben berkait enzim (ELISA) digunakan untuk mengukur aras interferon-gamma (IFN- γ), interleukin (IL)-8, IL-10, IL-12, IL-18, perforin, dan granzyme B dalam darah penderma sihat bagi menentukan kepekatan MEPB terbaik dalam mengaktifkan sel NK. Ketulenan dan kiraan sel NK dinilai menggunakan analisis sitometri aliran dan tripan biru. Analisis sitometri aliran dan ELISA digunakan untuk menilai keupayaan MEPB untuk meningkatkan ketoksikan sel NK terhadap sel MDA-MB-231. Ketoksikan dan keteratogenan daun MEPB dinilai dengan meneliti kitaran estrus, berat badan, tingkah laku umum dan tanda klinikal, analisis histopatologi, berat badan mutlak organ viseral janin, dan hasil kehamilan termasuk bilangan corpora lutea dan tapak implantasi, kematian sebelum dan selepas implantasi (%), berat rahim gravida, bilangan janin hidup dan mati, berat badan janin, nisbah jantina, dan pemeriksaan keseluruhan janin. Kajian ini menggunakan 40 ekor tikus betina yang termasuk 10 kumpulan tikus sebagai kawalan (air suling) dan 30 kumpulan tikus yang dirawat MEPB (250, 500, dan 1000 mg/kg/hari). Ujian MTT menunjukkan ketoksikan sederhana MEPB terhadap sel kanser payudara MDA-MB-231 dengan nilai IC₅₀ 64.57µg/ml. Data sitometri aliran menunjukkan bahawa MEPB boleh menyekat sel dalam fasa G0/G1 dan merangsang apoptosis dalam sel MDA-MB-231, dengan meningkatkan ekspresi Bax, p53, dan caspase-3 sambil menurunkan ekspresi Bcl-2. Hasil kajian menunjukkan bahawa daun MEPB mempunyai keupayaan untuk meningkatkan aras IFN-γ, IL-12, IL-18, perforin, dan granzyme B dan menurunkan aras IL-8 dan IL-10 dalam darah penderma sihat. Pesakit kanser payudara didapati mempunyai kurang sel NK berbanding penderma yang sihat, dan kira-kira 87.09% sel NK telah ditulenkan dengan berkesan. MEPB mengaruh sel NK untuk membunuh sel MDA-MB-231 secara apoptosis, melalui peningkatan perforin, granzyme B dan IFN-y, dalam darah pesakit kanser payudara dan penderma sihat. Keputusan menunjukkan bahawa kumpulan tikus yang dirawat dengan pelbagai dos daun MEPB tidak menjejaskan parameter ketoksikan, hasil kehamilan, dan parameter ketoksikan ke atas fetus. Penemuan kami menyimpulkan bahawa daun MEPB mengaruh apoptosis dalam sel MDA-MB-231, mempunyai keupayaan yang signifikan untuk mengatur sitokin, meningkatkan ketoksikan sel NK

terhadap sel kanser, tanpa sebarang bukti ketoksikan dan keteratogenan dalam kumpulan tikus yang dirawat dengan MEPB.

ANTICANCER-IMMUNE RESPONSE TOWARDS BREAST CANCER CELL LINES MDA-MB-231 AND TERATOGENIC ASSESSMENT OF Pereskia bleo LEAVES

ABSTRACT

Conventional treatment for breast cancer, especially radiation and chemotherapy, have significant adverse effects on patients. Thus, an increased global focus on finding nontoxic and cytoselective treatments has emerged, which includes the study on herbs. Various herbs showed excellent bioactivities; however, there are herbs that can be toxic and teratogenic at certain dosage. The current study was conducted to assess the anti-cancer properties of P. bleo leaves in terms of their ability to induce apoptosis and to determine its anti-cancer-immune response by stimulating Natural Killer (NK) cells' cytotoxicity against MDA-MB-231 breast cancer cells, as well as to evaluate its toxicity and teratogenicity in the animal model. Hexane, ethyl acetate and methanolic extracts of *P. bleo* leaves were tested for their cytotoxicity against normal cells MCF-10A and MDA-MB-231 cell lines by MTT assay. Methanolic extract showed the best activities and was used for subsequent experiments. Annexin V/PI assay and flow cytometric analysis were used to measure the induction of apoptosis, cell cycle arrest, and apoptotic protein expression by methanolic extract of *P. bleo* leaves (MEPB). Enzyme-linked immunosorbent assay (ELISA) was utilised to measure the level of interferon-gamma (IFN- γ), interleukins (IL)-8, IL-10, IL-12, IL-18, perforin, and granzyme B in healthy blood donors to determine the best concentration of MEPB leaves for activating NK cells. Flow cytometry and trypan Blue were used to measure NK cell counts and purity for subsequent experiments. Flow cytometric analysis and ELISA were used to determine

the ability of MEPB to enhance NK cell cytotoxicity against MDA-MB-231 cells. The toxicity and teratogenicity of MEPB leaves were evaluated by observing the oestrous cycle, body weight, general behaviour and clinical signs, histopathological analyses, absolute body weights of dam's visceral organs, and pregnancy outcomes, including the numbers of corpora lutea and implantation sites, pre- and post-implantation death (%), gravid uterine weight, number of live and dead foetuses, foetal body weight, sex ratio, and gross examination of the foetuses. The study used 40 female rats and was divided into 10 rat-control groups (distilled water) and 30 rat-MEPB groups (250, 500, and 1000 mg/kg/day). The MTT assay showed moderate cytotoxicity of MEPB leaves towards MDA-MB-231 breast cancer cells with an IC₅₀ value of 64.57µg/mL. The flow cytometry data indicated that MEPB can arrest the cell cycle at the G0/G1 phase and stimulate apoptosis in MDA-MB-231 cells, increasing the Bax, p53, and caspase-3 while decreasing Bcl-2 expression. The results indicated that MEPB leaves could upregulate IFN-y, IL-12, IL-18, perforin, and granzyme B levels and downregulate IL-8 and IL-10 levels in healthy blood. Breast cancer patients were found to have fewer NK cells than healthy donors, and approximately 87.09% of NK cells were effectively isolated. MEPB enhanced NK cells to kill MDA-MB-231 cells via apoptosis by upregulating perforin, granzyme B, and IFN- γ in healthy and breast cancer patient donors. The study also showed that the rat groups treated with various doses of MEPB leaves did not affect toxicity parameters, pregnancy outcomes, and foetotoxicity parameters. Our findings concluded that MEPB leaves induced apoptosis in MDA-MB-231 cells, with a significant capacity to regulate cytokines and increase NK cell cytotoxicity towards cancer cells without any evidence of toxicity and teratogenicity in rat-MEPB groups.

CHAPTER 1

INTRODUCTION

1.1 Background of the study

Cancer is a debilitating disease and one of the biggest threats to life, often leading to a slow and gradual decline (de Martel *et al.*, 2020). Hippocrates, a Greek physician, invented the term "cancer" between 460 and 370 B.C. It originates from the word "karkinos," which means "carcinoma" (Sudhakar, 2009). Globally, the mortality rate of cancer is rising yearly. Statistically, cancer is reported to be the 4th most common cause of death (National Cancer Registry, 2018). Effective treatments for breast cancer commonly include surgery, chemotherapy, targeted therapy, and endocrine therapy, such as tamoxifen (Dhanasekaran, 2020). These therapies cause side effects, such as headaches, fatigue, weakness, hair loss, nausea, vomiting, diarrhoea, mouth sores, dry mouth, damage to the immune system, potential organ failure, and digestion of both normal and cancer cells (Ko *et al.*, 2018).

Owing to the significant side effects of most therapies, researchers have been prompted to look at natural substances, particularly medicinal plants, for their possible anticancer benefits (Panyajai *et al.*, 2022). In the Holy Qur'an, about 19 medicinal plants are listed (Urbi *et al.*, 2014). The phytochemical compounds confer therapeutic qualities to plant-based medicines (Cui *et al.*, 2018). Due to their abundance of bioactive chemicals, the medicinal plants recognised by the World Health Organization (WHO) showed that 5–15% of them contain anticancer medicines (Shabani, 2016).

Malaysia ranks 12th in biodiversity, especially for *Pereskia bleo (P. bleo)* (Paramanick & Sharma, 2017). *P. bleo* is described as a traditional medicine and a member of the Cactaceae family. Previous studies acknowledged the activity of this plant towards various diseases, such as headaches, diabetes, cardiovascular diseases,

neurological disorders, ulcers, gastric pain, cancer, blood pressure, rheumatism oedema, and obesity (Vijayablan *et al.*, 2021). Malaysians consume *P. bleo* leaves as a vegetable in soups and salads (Garcia *et al.*, 2019). *P. bleo* leaves are approved for their antibacterial, antioxidant, anti-inflammatory, and anti-cancer properties (Fattepur *et al.*, 2020). However, according to previous data, the methanolic extract of *P. bleo* leaves had cytotoxic effects and induced apoptosis against breast cancer cells (MCF-7) (Malek *et al.*, 2007).

Several studies have reported that the bioactive compounds of medicinal plants can enhance immune cells, such as NK cells, to fight cancer (Grudzien & Rapak, 2018). Natural killer (NK) cells are employed in cancer immunotherapy (Morvan & Lanier, 2016). NK cells, one of the most common innate immune systems, play a role in the natural defences of the body (Choucair *et al.*, 2019). On the other hand, NK cells are designed to digest target cells, such as infection or cancer, while avoiding harm to normal cells (Voss & Bryceson, 2017). Many studies demonstrated that activated NK cells act as an anti-tumour agent (Gauthier *et al.*, 2019). Both perforin and granzyme B, found on the surface of NK cells, play an essential role in digesting target cells, eventually resulting in the death of cells (Liesche *et al.*, 2018). However, NK cells can release cytokines like interferon-gamma (IFN- γ), which help to destroy the target cells (Chiossone *et al.*, 2018).

Most of the therapeutic drugs used to cure related and unrelated pregnancy issues are harmful to pregnancy and exhibit teratogenic effects on the foetus (de Faria *et al.*, 2004). Due to concerns about the health of the foetus, many pregnant women prefer to treat their symptoms with medicinal plants instead of pharmaceutical medicines (Holst *et al.*, 2009). The foetus is most vulnerable to teratogenicity during the first trimester of pregnancy; therefore, pregnant women must exercise caution

during this time (Calvasina *et al.*, 2007). There is currently a lack of data on the risk or safety of medicinal plants during pregnancy. Previous studies showed that the adverse side effects of anticancer-derived plants on normal cells can lead to teratogenic, mutagenic, structural malformations, damage, growth retardation, as well as carcinogenic and sometimes leading to death or congenital disability (Bentil, 2015). For instance, an experiment was conducted on a pregnant woman who utilised anticancer-derived plants such as cyclophosphamide, which resulted in the newborn foetuses having abnormalities in structure and function (Nama & Shehata, 2012). According to a previous study, Methotrexate, which is used to treat leukaemia, lymphoma, and breast cancer, led to anomalies, miscarriages, and skeletal abnormalities with ambiguous genitalia during the first trimester (Addar, 2004). A study revealed that the drug Bleomycin, used to treat lymphoma, ovarian cancer, and teratoma can cause plagioccephaly syndactyly (fourth and fifth fingers) during the second and third trimesters (Cardonick et al., 2010). Furthermore, the side effects of the Carthamus tinctorius plant turned out to be eyelid defects, brain defects, renal toxicity, and hepatic toxicity (Baradaran et al., 2014). According to the findings of an experiment conducted on rats, large dosages of ginger administered to pregnant rats resulted in increased foetal weight, foetal loss, and bone maturation (Hepner et al., 2002).

1.2 Problem statement and rationale of the study

Radiation and chemotherapeutic medicines are used to treat cancer today, and both have significant side effects on patients. Thus, an increased global focus on finding nontoxic treatments for healthy cells that are toxic to cancer cells has emerged. *P. bleo* has been proven in earlier investigations to have anti-cancer properties. To provide useful basic pharmacological information on this medicinal plant, further research is needed to understand the potential of *P. bleo* extract to produce cytotoxicity and increase immunological activation. This study might help researchers better understand the role of *P. bleo* leaves in apoptosis induction and cytotoxicity of NK cells against cancer cells. However, the findings of this study may potentially aid alternative medicine, as improved *P. bleo* leaves have been shown to have beneficial effects on cancer cytotoxicity. However, information on the interaction with toxicity is still lacking. Previously, the acute oral toxicity of *P. bleo* on mortality or adverse effects of the plant at a maximum dose of 2500 mg/kg was tested by Sim and colleagues, but it is still unclear. Therefore, in this study, the teratogenicity assessment of *P. bleo* was done similar to Sim *et al.* (2010a). Medicinal herbs may help relieve related and unrelated pregnancy issues (Westfall, 2004). On the other hand, they play a vital role in abortion induction and teratogenic effects (Tang *et al.*, 2012). During the first trimester, the foetus is most sensitive to teratogenicity; thus, pregnant mothers must be cautious (Calvasina *et al.*, 2007).

1.3 Objective of study

1.3.1 General objective

To assess the anti-cancer immune response towards MDA-MB-231 breast cancer cell lines, toxicity and teratogenicity of *P. bleo* leaves extract.

1.3.2 Main objective of the study

1. To determine the antiproliferative activities of *P. bleo* leaves extract on breast cancer cell lines (MDA-MB 231) and normal breast cell lines (MCF-10A).

- 2. To assess the mode of cancer cell death induced by the *P. bleo* leaves extract on the selected cancer cells via a cell cycle arrest assay, Annexin V staining, and apoptotic protein expression, including Bax, Caspase-3, p53, and Bcl-2.
- To analyse the killing effect of NK cells on breast cancer cells induced by *P*. *bleo* leaves extract by evaluating the expression of interferon-gamma (IFN-γ), perforin, and granzyme B.
- 4. To evaluate the female toxicity and teratogenic effects of *P. bleo* leaves extract.

CHAPTER 2

LITERATURE REVIEW

2.1 Medicinal plants

Plants have always been a source of medicine. Many modern medications have been derived from plants formerly used in traditional treatments. They play an essential role in preserving health and developing novel therapies in various regions of the world (Dutta *et al.*, 2020).

In basic terms, "herb" refers to medicinal plants. Medicinal plants are a broad category of plants utilised in medicine to treat disease and have health-promoting properties. Worldwide, more than 94 plant species are used medicinally and in traditional healthcare systems. Previous studies have reported around 122 medicinal compounds isolated from these plants (Yuan et al., 2016). Statistics show that 65-85% of the world's population uses herbal medicine as their major source of healthcare (Kifle et al., 2021). The use of herbal medicine is estimated to be between 5.9% and 48.3% in Europe, 17.9% in the United States, and 12% in Canada (Eardley et al., 2012). Over 80% of people in Asia and Africa rely on medicinal plants (Chemburkar et al., 2014). Traditional medicine is commonly practised in many Asian nations despite the availability of allopathic treatment (Gunjan et al., 2012). Medicinal plants are described in Malaysia as crude plant concoctions used in alternative medicine techniques, such as Malay medicine, traditional Malay medicine, Ayurvedic medicine, naturopathy, homoeopathy, and some are also sold as nutritious foods or dietary supplements (Jantan, 2006). According to a previous study, most patients supplement their conventional medical prescriptions with herbal medicine because most individuals believe herbal medication is natural, harmless, and has fewer adverse effects than synthetic medicines or the notion that natural treatments are more dependable and effective than pharmaceuticals (Vidya & Lohit, 2019). Phytochemicals are natural compounds found in various parts of medicinal plants (e.g., roots, leaves, and flowers) that act together with nutrients and fibres to protect against disease (Hussein & El-Anssary, 2019). Therefore, medicinal plants that are rich in phytochemicals have been utilised as anti-cancer, anti-inflammatory, anti-microbial, and antioxidant agents (Ashaari *et al.*, 2020) (Table 2.1).

Source of plant	Name of Phytochemical compound	Treatment for Anti-cancer effects		
Podophyllum peltatum and Podophyllum emodii	Podophyllotoxin	Skin cancers, warts lymphomas, bronchial, and testicular cancer (Tan <i>et al.</i> , 2011)		
Podophyllum hexeandrum	Podophyllotoxin	Testicular, lung cancer, leukemias, ulcers, wounds, constipation, and tuberculosis therapy (Imbert, 1998)		
Cephalotaxus harringtonia	Cephalotaxus alkaloids	Chronic and acute myelogenous leukemia (Feldman <i>et al.</i> , 1996)		
Colchicum autumnale (Colchicaceae)	Colchicine	Crystal arthritis, cirrhosis, and gout (Negi <i>et al.</i> , 2015)		
Bleekeria vitensis and Ochrosia elliptica	Ellipticine	Cure ependymoblastoma, leukemia, myeloma, melanoma, breast, and colon cancer (Isah, 2016)		
Tinospora cordifolia, Berberis vulgaris, Berberis aquifolium, and Rhizoma coptidis	Berberine	Breast, prostate, and colorectal cancer (Barzegar <i>et al.</i> , 2015)		
<i>Combretum caffrum</i> (Combretaeae)	Combretastatins	Leukemia, lung, and colon cancer (Lauritano <i>et al.</i> , 2016)		
T 1 11 .	Combretastatins	Cure medullary thyroid and anaplastic thyroid cancer (Garon <i>et al.</i> , 2016)		
<i>Terminalia bellerica</i> (Combretaceae)	Triterpenoid acids	Both <i>in vitro</i> and <i>in vivo</i> against leukemia, pancreatic, and breast cancer (Cragg & Newman, 2005)		

Table 2.1:List of bioactive compounds of plants that have anti-cancer activities.

Table 2.1: Continued						
Ziziphus mauritiana, Ziziphusrugosa, Ziziphus oenoplia, and Betula Sp. (Betulaceae)	Betulinic acid Melanoma (Prakash <i>et al.</i> , 2013)					
Red pepper	Capsaicin	Anti-cancer, antimutagenic, antimetastatic, anti-angiogenic, and chemopreventive functions in pancreatic, prostatic, liver, skin, leukemia, lung, bladder, colon, and endothelial cells (Clark & Lee, 2016)				
Zingiber officinale	Gingerol	Colon, pancreas, ovarian, and breast cancer (Park <i>et al.</i> , 2006; Oyagbemi <i>et al.</i> , 2010)				
Crocus sativus L.	Saffron (Crocetin)	Lung, liver, skin, pancreatic colorectal, and breast cancer therapy (Hoshyar & Mollaei, 2017)				
Mushroom	Vitamin D	Colon cancer (Gorham <i>et al.</i> , 2007), breast, pancreatic, and ovarian cancer (Buyru <i>et al.</i> , 2003)				

2.1.1 Anti-cancer activities of medicinal plants

The immune system comprises special cells, tissues, and organs that substantially defend the body from external invasion of infectious or harmful microorganisms caused by immune system malfunction (Pandey *et al.*, 2023). Neutrophils, macrophages, basophils, monocytes, dendritic cells, natural killer cells (NK cells), and lymphocytes (T and B) play a vital role in the effector functions of innate and adaptive immunity and serve as the first lines of defence against various pathogens, such as viruses, bacteria, and cancers in the human body (Miyake *et al.*, 2017).

Cancer is triggered by DNA damage in cells or can be caused by changes in DNA (mutations), which results in abnormal growth and division (Alhmoud *et al.*, 2020). Currently, the most common types of cancer therapy are surgery,

chemotherapy, and radiotherapy. These methods negatively impact healthy cells, leading to more studies aimed at discovering new and safe cancer treatments (Knight *et al.*, 2021). The plant kingdom has significant impacts on human health. The major role of herbal medicines is to restore the capacity of the body to defend, control, and cure diseases. They are usually taken in the form of powders, pills, and extracts (Lichota & Gwozdzinski, 2018). Approximately 60% of anti-cancer drugs are derived from plant products (Kooti *et al.*, 2017) (Table 2.2).

Source of plant	Drug	Use of treatment		
Catharanthus roseus (Apocynaceae)	Vinca Alkaloids, vinblastine (VLB), and vincristine (VCR)	Lymphomas, leukemias, breast cancer, testicular cancer, lung cancer, and Kaposi's sarcoma (Singh <i>et al.</i> , 2013)		
(Apocynaccac)	vincristine (VCR)	Against leukemia, acute lymphocytic leukemia in childhood (Zaid <i>et al.</i> , 2012)		
Taxus baccata, Tsuga canadensis, and Corylus avellana	Taxanes (Paclitaxel and taxol), milataxel, ortataxel, and tesetaxel)	Metastatic breast cancer, AIDS- related Kaposi sarcoma, non- small cell lung carcinoma, and bladder cancer (Christensen, 2022) Ovarian, lung, pancreas, and prostate cancer (Xie & Zhou, 2017)		
	Larotaxel	Urethral bladder, pancreatic, lung, and breast cancer (Ojima <i>et al.</i> , 2016)		
Podophyllum peltatum	Etoposide	Hodgkin's and non-Hodgkin's lymphoma, lung, gastric, breast, and testicular cancer (Montecucco <i>et al.</i> , 2015)		
Camptotheca acuminata	Campothecin derivatives, such as topotecan (hycamtin) and irinotecan	To cure colorectal, ovarian, and small-cell lung cancer (Rahier <i>et al.</i> , 2005)		

Table 2.2:Plants derived anti-cancer drugs.

Table 2.2: Continued

.

.

.

Cephalotaxus harringtonia	Cephalotaxus (harringtonine and homoharringtonine cephalotaxus alkaloids) (Feldman <i>et al.</i> , 1996)	Homoharringtonine: To cure chronic and acute myelogenous leukemia (Feldman <i>et al.</i> , 1996) Harringtonine plus with Homoharringtonine: To treat chronic myelogenous leukemias, acute myelogenous leukemia (Cragg <i>et al.</i> , 2006)		
Bleekeria vitensis and Ochrosia elliptica (stem, bark, leaf and root) (Stiborová at al	Ellipticine derivatives (N-2- (diethylaminoethyl)– 9– budrouvallipticipium	To cure ependymoblastoma, leukemia, myeloma, melanoma, breast and colon cancer (Kizek <i>et al.</i> , 2012; Isah, 2016)		
root) (Stiborová <i>et al.</i> , 2014)	hydroxyellipticinium chloride, 2-N-methyl 9- hydroxyellipticine)	Ellipticine (elliptinium): To cure breast cancer (Ohashi <i>et al.</i> , 1995)		
Tinospora cordifolia, Berberis vulgaris, Berberis aquifolium, and Rhizoma coptidis (root and rhizome) (Mantena et al., 2006)	Berberine	To treat breast, prostate and colorectal cancer (Barzegar <i>et al.</i> , 2015)		
Combretum caffrum	Combretastatins	To cure leukemia, lung cancer and colon cancers (Lauritano <i>et</i> <i>al.</i> , 2016)		

Terpenoids, flavonoids, alkaloids, and steroids are secondary metabolites found in medicinal plants with anti-cancer properties (Sumner, 2000). These secondary metabolites inhibit cancer-stimulating enzymes, repair DNA, stimulate anti-cancer enzyme synthesis in cells (caspase-3, caspase-7, caspase-8, caspase-9, caspase-10, and caspase-12), and induce antioxidant effects (Sakarkar & Deshmukh, 2011). Plant metabolites stimulate apoptosis in cancer cells (Sohi *et al.*, 2003). *In vitro* experiments show that several medicinal plants have anti-cancer properties due to their abundant bioactive compounds, which are essential in inhibiting cancer growth and can enhance the ability of the immune system to target cancer cells. For instance, the root extract of the *Dicoma anomala Sond* plant exhibited anti-cancer agents on MCF-7 breast cancer cells (Shafiq *et al.*, 2020), the *Fagaropsis angolensis* plant on prostate cancer cells known as DU-145 and breast cancer cells known as HCC1395 (Misonge *et al.*, 2019), whereas the *Prunus avium* plant on breast cancer cells known as MDA-MB-453 (Layosa *et al*, 2021). Ji and co-workers showed that polysaccharides isolated from the *Cynanchum paniculatum* plant had a powerful anti-cancer effect on both mice hepatic cancer cells (H22), human liver cancer cells (HepG2), mice sarcoma cells (S180), and lung cancer cells (A549), which they attributed to the capacity to activate immune cells, splenic NK cells and peritoneal macrophages (Ji *et al.*, 2022). The list of medicinal plants with anti-cancer properties is in Table 2.3.

Name plant	Extracts	Mechanism of action
Achillea wilhelmsii	Methanol extract of leaves	Induce apoptosis in colon, stomach, breast, and melanoma cells (Uddin <i>et al.</i> , 2011)
Allium sativum	Methanolic extract	Anti-cancer activity against MCF-7 breast cancer and bladder cancer cells (Abdullaev, 2001; Karmakar <i>et al.</i> , 2011)
Vernonia amygdalina	Extract	Reverses the cancer in MCF-7 breast cancer cells and increased the basal apoptotic but decreased the angiogenic activity in mice (Sigstedt <i>et al.</i> , 2008) Anti-cancer activity in breast MCF-7 and MDA- MB-231breast cancer cells and inhibits the proliferation of cells (Gresham <i>et al.</i> , 2008)
Morus alba	Methanolic extracts	Anti-proliferative effects on pulmonary carcinoma (Calu-6), colon carcinoma (HCT-116), and MCF-7 breast cancer cells (Chon <i>et al.</i> , 2009)

Table 2.3:Type of medicinal plants that have anti-cancer activities towardsvarious cancers.

Table 2.3: Continued				
	Albanol root extract	Apoptosis-inducing, cytotoxic activity in HL-60 cells Decrease the level of pro-caspases 3, 8, and 9 by induced topoisomerase II. Bax/Bcl-2 ratio increased. Induced HL-60 apoptotic cell death through stimulation of the death receptor (Kikuchi <i>et al.</i> , 2010)		
	Methanolic leaf extract	Inhibition of HepG2 cells (Naowaratwattana et al., 2010)		
	Aqueous and ethanol extracts	Anti-cancer activity against HepG2 and SMMC-7721liver cancer cells, BGC-823 gastric cancer cells, LoVo and SW-116 colon cancer cells, and CaEs-17 esophagus cancer cells (Li <i>et al.</i> , 2012)		
Paris polyphylla	Extract	Anti-cancer effects in ECA109 oesophageal cancer cells by: Increasing the connexin26 mRNA and protein expression Increased Bad genes expression Decreased the expression of Bcl-2 genes Inhibiting the growth of ECA109 cells by apoptosis (Li <i>et al.</i> , 2012)		
<i>Fennel</i> (Aerial part)	Alcoholic extract	Enhance lymphocytes to reduce expression of IFN- γ and IL-4 (Pacifico <i>et al.</i> , 2018)		
<i>Laurus</i> nobilis Linn. (Lauraceae)	Ethanol extract	Enhance an acute lung injury mouse model to reduce IL-1 β , IL-6, and TNF- α expression (Lee <i>et al.</i> , 2019)		
Peppermint (<i>Mentha</i> piperita)	Maceration (Water)	Reduce IL-1, IL-6, and TNF- α in Male Wistar rats (Osman <i>et al.</i> , 2020)		

An alkali-soluble polysaccharide extracted from the *Angelica sinensis* plant can boost the functions of immune cells, including NK cells, macrophages, and lymphocytes. It could also raise the levels of immune cytokines, including IFN- γ , interleukin (IL)-2, and tumour necrosis factor- α (TNF- α), which ultimately enhance apoptosis in mice hepatic cancer cells (H22) by blocking the G0/G1 phase (Yu *et al.*, 2021). Almutairi and co-workers documented that the extracted seed of *Annona muricata* had anti-cancer properties and could induce apoptosis by expressing proapoptotic proteins, such as p53 and Bax protein (Almutairi *et al.*, 2023).

2.1.2 Immunomodulatory effects by medicinal plants

Immunostimulators and immunosuppressants are common immunomodulators (Gruppen *et al.*, 2018). Several medicinal plants have biological properties, including immunomodulatory effects (Lin et al., 2023). Previous research indicates that the most effective method to preventing and treating diseases is through immunostimulation with natural ingredients. Based on prior research, medicinal plants, such as Cyrtomium macrophyllum (Ren et al., 2014), Phyllanthus urinaria (Ilangkovan et al., 2013), and Asparagus racemosus (Gautam et al., 2004), have been reported to have immunomodulatory properties by suppressing or stimulating immune cells (Krensky et al., 2011). In a human model experiment, the researchers reported that Astragalus membranaceus root can decrease IL-6 levels (Denzler et al., 2010). Duansak and colleagues demonstrated in Wistar Furth rat models that the *Aloe vera* plant can lower TNF- α and IL-6 (Duansak *et al.*, 2003). A prior study showed that the rhizome extracts of the Acorus calamus inhibit nitric oxide, IL-2, and TNF-α production (Mehrotra et al., 2003). Hussain and co-workers documented that ethanolic and aqueous extracts of the Picrorhiza kurroa plant can stimulate humoral responses by antibody production, the release of mediators of hypersensitivity reactions, and tissue responses to these mediators in the target organs (Hussain et al., 2013).

2.1.3 Phytochemicals in medicinal plants

Plants produce a wide range of phytochemicals, which are classified into two categories based on their role in plant metabolism: primary metabolites, which comprise proteins, amino acids, sugars, and chlorophyll (Hussein & El-Anssary,

2019), and secondary metabolites, including alkaloids, flavonoids, terpenoids, phenolics, and steroids, sesquiterpenes, diterpenes, triterpene saponins, and triterpene aglycones. These chemical compounds are naturally synthesised in the various parts of plants, such as, leaves, stems, roots, seeds, and flowers (Jan *et al.*, 2021). Flavonoids, tannins, alkaloids, and phenolic chemicals have the most significant biological properties and exert substantial physical effects on the human body. For example, phenolic compounds play a role in increasing bile secretion and reducing blood cholesterol (Ghasemzadeh *et al.*, 2010), and flavonoids have potential as a treatment for various malignancies (Kleemann *et al.*, 2011). Alkaloids, including vindesine, vinorelbine, vinblastine, and vincristine, have been proven in previous studies to have potent anti-cancer properties. For instance, vincristine and vinblastine against breast cancer, lymphoblastic leukaemia, and skin cancer (Mishra & Verma, 2017), vinorelbine against liver and colon cancer (Liu *et al.*, 2020), whereas vindesine against lung cancer (Arora *et al.*, 2010).

Previous studies demonstrated and confirmed that secondary metabolites play an essential role in human therapy against various ailments, including chronic or infectious diseases and cancer (Sharifi-Rad *et al.*, 2016). Previous studies documented that bioactive compounds have anti-bacterial, antioxidant, anti-fungal, anti-diabetic, anti-cancer, anti-viral, and anti-inflammatory properties (Umaru *et al.*, 2018). A variety of phytochemicals, including tannins, alkaloids, phenols, naphthoquinones, flavonoids, saponins, steroids, carbs, mucilage, gum, and resin, were extracted from the methanolic extract of *Carissa macrocarpa* leaves. According to the findings, these secondary metabolites boost the anti-bacterial effects of *Carissa macrocarpa* on *Escherichia coli* and *Staphylococcus aureus* (Ramasar *et al.*, 2022). Four secondary chemicals were extracted from *Senna petersiana* leaves, including GammaLinolenate, columnidin, L-lysine citrate, and hercynine. These compounds exhibited their anti-bacterial properties towards *Staphylococcus aureus*, *Escherichia coli*, *Pseudomonas aeruginosa*, and *Mycobacterium smegmatis* (Matotoka *et al.*, 2023). According to the phytochemical profile, the stem bark of *Beilschmiedia roxburghiana* is abundant with flavonoids, polyphenols, alkaloids, and tannins. These phytochemicals enhance antioxidant and anti-bacterial agents of various bacteria, such as *Salmonella typhi*, *Staphylococcus aureus*, *Shigella sonnei*, *Acinetobacter baumannii*, and *Klebsiella pneumonia* (Khanal *et al.*, 2022).

Apart from that, *Abutilon indicum*, comprising leaf, root, flower, and seed has been used as an antioxidant due to its rich bioactive compounds, such as mucilage, tannins, organic acids, traces of asparagine, magnesium phosphates, calcium carbonate, alkaline sulphates, sterols, alkaloids, glycosides, titerpenoids, amino acids, oils, terpenoids, steroids, terpenes, and flavonoids, which might contribute to its wide range of uses in the human body, including treatment of fever, toothache, blood dysentery, allergies, piles, leprosy, cystitis, bronchitis, and gonorrhoea. Additionally, it is used as an aphrodisiac and a diuretic (Panda, 1999).

2.2 Pereskia bleo (Cactaceae)

Cacti are well-known desert plants distinguished by their distinctive stem and leaf development forms. There are roughly 2000 species in the Cactaceae family, which has 100 genera (Zareisedehizadeh *et al.*, 2014). Maihuenioideae, Pereskioideae, Opuntioideae, and Cactoideae are the four subfamilies. Over 90% of the species belong to the Opuntioideae and Cactoideae subfamilies (Ortega-Baes *et al.*, 2010). The ecological characteristics are connected to morphological and physiological changes that enable them to retain more water and survive in arid environments (Edwards & Diaz, 2006; Edwards & Donoghue, 2006).

2.2.1 General description of *Pereskia bleo*

P. bleo belongs to the plant botanical family Cactaceae, commonly known as "Pokok Jarum Tujuh Bilah" in Malaysia (Abdul-Wahab et al., 2012), while in Chinese, it is called "Cak Sing Cam" or "Qi Xing Zhen" (Wahab et al., 2009). It is recognised by many English names, including rose cactus, wax rose, and leaf cactus (Yen et al., 2013). The genus Pereskia belongs to the Pereskioideae (Hunt, 2016). Except for P. bleo, which lives in areas with higher annual rainfall, most Pereskia species are found in dry forests or thorny scrubs in tropical regions with a dry season and very arid forest life (Edwards & Donoghue, 2006). P. bleo is used as a medicinal plant that originated from South America, Brazil, Mexico, and Central America. It is widely distributed and cultivated in many tropical and subtropical regions, including Malaysia, China, and India (Zareisedehizadeh et al., 2014). P. bleo can reach a height of 0.8-8 m. It has bright green and large leaves, orangish-red blooms (depending on the species; flowers might be white, yellow, or red) that can occur singly or in groups, and long spiky stems (5 to 7 black spines of 1 cm in length). However, there are only 1 to 4 young shoots. The formation of areoles may be seen in these spines, which have 1 to 5 cm diameter and resemble roses (Figure 2.1). Fruits are commonly waxy, round, and green with dark shading, which changes to yellow when ripe (Yen et al., 2013). P. bleo has been shown to have anti-cancer, anti-rheumatic, anti-ulcer, anti-inflammatory, antioxidant, and anti-microbial properties in previous research (Zareisedehizadeh et al., 2014).



Figure 2.1: A: The stem of *P. bleo* (Zareisedehizadeh *et al.*, 2014), B. leaves and the orangish-red flowers of *P. bleo* (Abdul-Wahab *et al.*, 2012).

2.2.2 Classification of *P. bleo*

Table 2.4:	P. bleo categorization	(Butterworth &	Wallace.	2005).

Division	Class
Kingdom	Plantae
Phylum	Tracheophyta
Class	Magnoliopsida
Order	Caryophyllales
Family	Cactaceae
Subfamily	pereskioideae
Genus	Pereskia
Species	Pereskia bleo

2.2.3 Medicinal uses of *P. bleo*

The leaves of *P. bleo* contain carbon (50.6%), magnesium (0.4%), sulphur (1.5%), oxygen (35.4%), chloride (1.2%), potassium (10.16%), phosphorus (0.4%), and calcium (0.3%) (Abbdewahab *et al.*, 2009); hence, it is used either in healthy soups or eaten raw as salad (Garcia *et al.*, 2019). *P. bleo* leaves are commonly used in traditional medicine for cancer, high blood pressure, diabetes, rheumatism and inflammation, atopic dermatitis, ulcers, gastric pain, headaches, and haemorrhoids (Vijayablan *et al.*, 2021). Potassium is abundant in *P. bleo* leaves (10.16%), and a high-potassium diet has been demonstrated to impact blood pressure reduction significantly (Geleijnse *et al.*, 1994). *P. bleo* is also used to cure gastrointestinal

disorders in Panama (Gupta *et al.*, 1996). The plant has been reportedly prepared in various ways, commonly consumed raw or as a decoction made from the leaves. The traditional uses and preparation methods of *P. bleo* are shown in Table 2.5.

Purpose	Method of preparation			
Detoxification and prevention of cancer	Boiling the leaves or fruit to make tea (Rahmat <i>et al.</i> , 2013)			
Dietary purposes and health maintenance	Eating the raw leaf, flower, and fruit (Malek <i>et al.</i> , 2009)			
Health maintenance and revitalising the body	Boiling the leaves in water and extracting the juice (Rahmat <i>et al.</i> , 2013)			
To relieve muscular pain	Making decoction from the leaves and then using as a warm bath for muscle ache (Gupta <i>et al.</i> , 1993)			
To treat haemorrhoids, hypertension, diabetes, infections, headaches, and inflammatory conditions (rheumatism and asthma).	No information is available in the literature (Malek <i>et al.</i> , 2009; Er <i>et al.</i> , 2007)			

Table 2.5:Traditional usage and methods of preparation of P. bleo.

2.2.4 Chemical constituents of *P. bleo*

Alkaloids (namely 3,4-dimethoxy- β -phenethylamine, mescaline, 3methoxytyramine, and tyramine), carotenoids, flavonoids, terpenoids, sterols (e.g., beta-carotene), fatty acids, lactone, phytosterol glycoside, phenolic compounds, and alpha-tocopherol, are phytochemical compounds of *P. bleo* leaves (Johari & Khong, 2019). On the other hand, Goh verified that stigmasterol, β -sitosterol, dihydroactinidiolide, and campesterol compounds were also isolated from *P. bleo* (Goh, 2000). Meanwhile, β -sitosterol, 2,4-ditert-butylphenol, and phytol were isolated through ethyl acetate extraction of *P. bleo* in another investigation (Malek *et al.*, 2007) (Table 2.6).

Alkaloids	3,4-Dimethoxy- β - phenethylamine	3-Methoxytyramine
	Tyramine HO NH ₂	Mescaline $\bigvee_{0}^{0} \bigvee_{1}^{NH_2}$
Fatty acids	Methyl palmitate	Methyl linoleate
Flavonoids	Vitexin How	
Phytosterol glycoside	β-Sitosterol glucoside β_{Ho}	3~5
Lactone	Dihydroactinidiolide	≥
	2,4- Ditert-butylphenol	α-Tocopherol
Phenolic compounds	Catechin HO Catechin	Epicatechin
	Quercetin Ho	Myricetin HO CONTRACTOR
Sterols	Campesterol H_{H_0C}	Stigmasterol
	β -Sitosterol	
Terpenoids	β- Carotene	Phytol Y

Table 2.6:Components of P. bleo leaves (Vijayablan et al., 2021).

2.2.5 Bioactivity of *P. bleo*

The *P. bleo* plant was found to have a wide range of pharmacological activities, such as anti-nociceptive, anti-bacterial, antioxidant, anti-microbial, anti-diabetic, and anti-cancer (Azizan et al., 2024). The bioactive compounds discovered in the extract are linked to the powerful pharmacological properties of the plant. For instance, researchers have shown that flavonoid chrysin isolated from P. bleo extracts has medicinal properties for treating diabetes. It may lower triglyceride and glucose levels (Mat Darus & Mohamad, 2017), and downregulate the expression of pro-inflammatory cytokines linked to the onset of diabetes and its complications, such as atherosclerosis and other heart disorders (Ramírez-Espinosa et al., 2017). Additionally, chrysin and Apigenin 6-C-glucoside, discovered in *P. bleo* extract, play a major role in elevating insulin production (Mat Darus & Mohamad, 2017). Chan and co-workers reported that the extract from P. bleo leaves was found to exhibit antifungal activity on Trichoderma mentagrophytes, Cryptococcus neoformans, Aspergillus brasiliensis, Candida albicans, Issatchenkia orientalis, and Candida parapsilosis (Chan et al., 2018). Previous research findings show that the P. bleo leaves extract exhibited antibacterial properties on Staphylococcus aureus, Escherichia coli, Pseudomonas aeruginosa, and Streptococcus pyogenes (Lee HongLim et al., 2009). Apart from the leaves, hexane, ethyl acetate, and dichloromethane extracts of P. bleo exhibited notable antinociceptive properties (Abdul-Wahab et al., 2012). Research conducted in vitro revealed that plants abundant in phenolic compounds have antioxidant properties. P. bleo leaves, which are abundant in phenolic compounds, were shown to have significant antioxidant properties according to DPPH test findings (Hassanbaglou et al., 2012). Phytol extracted from P. bleo leaves also exhibited anti-cancer activity in various cancer cells (Malek et al., 2009). The EtOAc fraction (\beta-sitosterol, 2,4-di-tertbutylphenol, α-tocopherol and phytol) exhibits specific anti-cancer effects on human carcinoma cells. Furthermore, the ethyl acetate fraction was more potent against human colon cancer (HCT116) and breast cancer cells (MCF-7) (Malek *et al.*, 2007). Based on their research, Mohd-Salleh *et al.* concluded that *P. bleo* leaf extracts had significant anti-cancer activity towards HeLa cervical cancer cells and MDA-MB-231 breast cancer cells (Mohd-Salleh *et al.*, 2020a).

2.3 Cancer

2.3.1 Cancer and prevalence

Cancer is derived from the Latin term carcinoma, which means crab. Previous cancer research witnessed tremendous improvement in the knowledge of cancer biology. Cancer originates from changes or mutations in genetic material (DNA). Moreover, cells in an organ proliferate uncontrollably, resulting in the formation of aberrant or abnormal cells known as cancer cells (Sitki-Copur, 2019). Tobacco, environmental contaminants, exposure to ionising radiation, infectious diseases, such as hepatitis B/C, helicobacter pylori and Human immunodeficiency virus (HIV), poor nutrition, and inherited genetic defects are among the causes of cancer (Blackadar, 2016).

Globally, the most frequent cancer therapies are surgery, radiation therapy, chemotherapy, hormone therapy, and immunotherapy (American Cancer Society, 2015; Siegel *et al.*, 2021). Radiation therapy and chemotherapy are currently prestigious medical specialties, encompassing subspecialties such as medical oncology. Chemotherapy and radiation therapy, on the other hand, can destroy DNA material while also harming normal cells, leading to a variety of side effects, such as vomiting, fatigue, nausea, hair loss, and in some cases, death (Aslam *et al.*, 2014).

21

According to the data from the WHO in 2020, cancer is the second most significant cause of death worldwide (Ferlay *et al.*, 2020). According to available data, in 2021, there were approximately 192,000 new cases of cancer and 61,000 fatalities attributed to cancer (Siegel *et al.*, 2021). Globally, 2020 witnessed a significant increase in the number of cancers (Figure 2.2A), with around 19.29 million and 10 million cancer-related deaths (Figure 2.2B) (World Health Organization, 2022; Gonzalez-Valdivieso *et al.*, 2021).

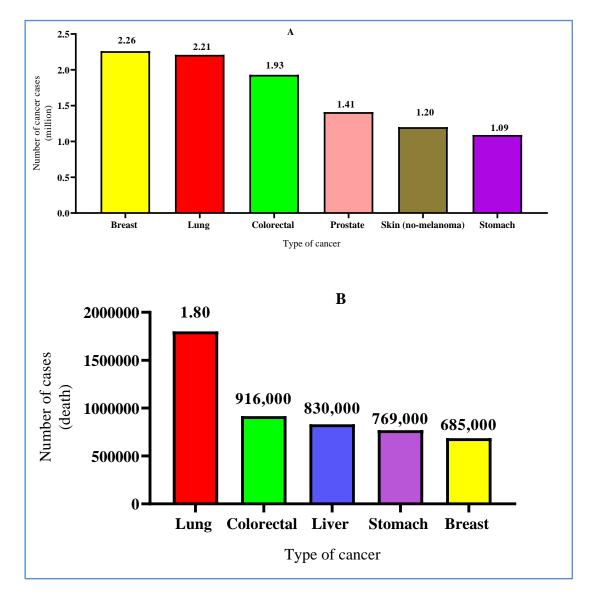


Figure 2.2: Incidence (A) and mortality rate of cancer (B) worldwide, 2020 (World Health Organization, 2022; Gonzalez-Valdivieso *et al.*, 2021).

Non-communicable diseases (NCDs) are on the rise and are responsible for approximately 73% of fatalities in Malaysia. These ailments include cardiovascular diseases, diabetes, cancers, and respiratory diseases (Institute for Public Health, 2020). Nevertheless, the most prevalent ailment in Malaysia is cancer. According to the WHO, cancer incidence rate and mortality are increasing year by year in Malaysia and worldwide. For instance, around 103,507 new cases in 2007 to 2011 (MALAYSIA National Cancer Registry Report, 2007–2011), 115,238 new cases in 2012 to 2016 (MALAYSIA National Cancer Registry Report, 2012–2016), and 48,639 new cancer cases in 2020 were reported. Cancer incidence and mortality rates are expected to grow by the year 2040. The incidence rate of cancer in 2020 for both sexes of all ages is illustrated in Table 2.7 (World Health Organization, 2022). 2022).

				S	exes				
Rank	Both sexes		Females			Males			
	Cancer	New cases	% of cancers	Cancer	New cases	% of cancers	Cancer	New cases	% of cancers
1	Breast	8,418	17.0%	Breast	8,418	32.9%	Lung	3,925	17%
2	Colorectal	6,597	10.6%	Colorectal	3,057	11.9%	Colorectal	3,540	15.4%
3	Lung	5,139	10.6%	Ovary	1,836	7.2%	Prostate	2,146	9.3%
4	Nasopharynx	2,222	4.6%	Cervix uteri	1,740	6.8%	Nasopharynx	1,703	7.4%
5	Liver	2,149	4.4%	Corpus uteri	1,401	5.5%	Liver	1,553	6.7%
6	Other cancers	24,114	49.6%	other cancers	9,135	35.7%	Other cancers	10,185	44.2%