

**CHARACTERIZATION OF HYDROPHOBIC  
SEAWEED FILM REINFORCED WITH KENAF  
BAST CELLULOSE NANOFIBERS**

**CHE MOHAMAD HAZWAN BIN CHE ISMAIL**

**UNIVERSITI SAINS MALAYSIA**

**2022**

**CHARACTERIZATION OF HYDROPHOBIC  
SEAWEED FILM REINFORCED WITH KENAF  
BAST CELLULOSE NANOFIBERS**

by

**CHE MOHAMAD HAZWAN BIN CHE ISMAIL**

**Thesis submitted in fulfilment of the requirements  
for the degree of  
Doctor of Philosophy**

**September 2022**

## ACKNOWLEDGEMENT

Foremost, I would like to express my sincere gratitude to Allah (SWT) for giving me the chance to go through these years of study. I would also like to extend my deepest gratitude to my family. Without their encouragement and support, I would not have finished this study. My most precious thanks to my advisor Prof. Datuk Ts. Dr. Abdul Khalil Shawkataly for the continuous support on this study and research, for his patience, motivation, enthusiasm, and immense knowledge. His guidance helped me in all times of research also in writing this thesis. It is such a big pleasure having a better advisor and mentor for my study. Besides my advisor, I would like to thank those who help me a lot on road to completing this thesis for their encouragement, insightful comments, and hard questions. I would like to thank my co-supervisor Asso. Prof. Dr. Mohamad Haafiz Mohamad Kassim and our senior research officer Ts. Dr. Che Ku Abdullah Bin Che Ku Alam. Thank you very much to USM Fellowship program for the financial support on my study. Not to forget our laboratory assistant Mr. Azhar Mohamad Noor Mr. Shamsul Zoolkiffli, Mrs. Noorhasni Othman, Mrs. Noraida Bukhari for their guidance in preparing apparatus and machines to complete this study. Thank you to my fellow Ph.D and Master lab-mates, for all the knowledge they had shared with me, also to my friend Tunku Syed Iskandar Al-Qadri for being with me since my first day at the university. For the most important part, I would like to wish a special thanks to my family members, especially to my late lovely mother, for all the sacrifices they had made for me to complete this study ever since. Thank you, my dear father, for making me a better person. Thank you to all the people I had met on this journey. Last but not least, to lina, my dear consort, eternal. May Allah bless all of us here and after.

## TABLE OF CONTENTS

<b>ACKNOWLEDGEMENT</b> .....	<b>ii</b>
<b>TABLE OF CONTENTS</b> .....	<b>iii</b>
<b>LIST OF TABLES</b> .....	<b>viii</b>
<b>LIST OF FIGURES</b> .....	<b>ix</b>
<b>LIST OF SYMBOLS</b> .....	<b>xii</b>
<b>LIST OF ABBREVIATIONS</b> .....	<b>xiii</b>
<b>ABSTRAK</b> .....	<b>xiv</b>
<b>ABSTRACT</b> .....	<b>xvi</b>
<b>CHAPTER 1 INTRODUCTION</b> .....	<b>1</b>
1.1 Overview .....	1
1.2 Problem statement .....	5
1.3 Research Objectives .....	8
1.4 Thesis Layout .....	9
<b>CHAPTER 2 LITERATURE REVIEW</b> .....	<b>10</b>
2.1 Macro algae .....	10
2.1.1 <i>Kappaphycus alvarezii</i> .....	12
2.2 Chemical and rheological properties of carrageenan .....	18
2.3 Extraction of carrageenan from seaweed .....	24
2.4 Biodegradable polymers and plastics .....	28
2.5 Natural biodegradable polymers: Polysaccharides.....	31
2.6 Hydrocolloids in red seaweed <i>Kappaphycus alvarezii</i> .....	36
2.7 Applications of seaweed in biocomposites .....	40
2.8 Natural fibers.....	44
2.9 Non-wood fibres: Kenaf fiber .....	48
2.10 Cellulose nanofiber .....	52

2.11	Extraction of Cellulose nanofiber .....	58
2.12	Hydrophobic chemical modification .....	61
2.13	Silane surface treatment .....	62
2.14	Filler as reinforcement in composite .....	67
2.15	Cellulose as reinforcement materials for biocomposites.....	69
2.15.1	Filler effect on mechanical properties .....	72
2.16	Biodegradation .....	75
2.17	Wettability .....	79
<b>CHAPTER 3    METHODOLOGY.....</b>		<b>83</b>
3.1	Materials and chemicals used.....	83
3.2	Extraction of Cellulose Nanofiber (CNF) .....	86
3.3	Fabrication of seaweed/CNF composite film.....	87
3.4	Silane surface treatment of seaweed/CNF composite film .....	88
3.5	Characterization of <i>Kappaphycus alvarezii</i> seaweed.....	89
3.5.1	FT-IR characterization of seaweed .....	89
3.6	Characterization of cellulose nanofiber (CNF) .....	90
3.6.1	Crystallinity (%) of cellulose nanofiber (CNF).....	90
3.6.2	Transmission Electron Microscopy (TEM) .....	90
3.6.3	Particle Size and Zeta Potential.....	90
3.6.4	FT-IR characterization of CNF .....	91
3.7	Physical characterization of seaweed/CNF composite film.....	91
3.7.1	Moisture absorption capacity (MAC) .....	91
3.7.2	Water vapour permeability (WVP) .....	92
3.7.3	Water Solubility (WS).....	92
3.7.4	Contact angle (CA).....	93
3.7.5	Crystallinity index .....	94
3.7.6	Weight percent gain (WPG).....	94

3.8	Chemical characterization of seaweed/CNF biocomposite film .....	95
3.8.1	FT-IR characterization .....	95
3.9	Mechanical characterization of seaweed/CNF composite film .....	96
3.9.1	Tensile strength (TS) .....	96
3.9.2	Young modulus (YM) .....	96
3.9.3	Elongation at break (EAB) .....	97
3.10	Morphological characterization of seaweed/CNF composite film .....	97
3.10.1	Surface morphology characterization (SEM) .....	97
3.10.2	Atomic Force Microscopy (AFM) .....	97
3.11	Thermal characterization of seaweed/CNF composite film .....	98
3.11.1	Thermogravimetric analyzer (TGA) .....	98
3.12	Biodegradable characterization of seaweed/CNF composite film .....	99
3.12.1	Soil burial testing .....	99
<b>CHAPTER 4 RESULTS AND DISCUSSIONS .....</b>		<b>100</b>
4.1	Characterization of cellulose nanofiber (CNF) .....	100
4.1.1	Morphological characterization of CNF .....	100
4.1.2	FT-IR characterization of CNF .....	101
4.1.3	Particle size and Zeta potential of CNF .....	104
4.1.4	Crystallinity index of cellulose nanofiber .....	107
4.2	Chemical characterization of seaweed .....	109
4.2.1	FT-IR characterization of seaweed and kappa carrageenan .....	109
4.3	Physical characterization of modified seaweed film .....	111
4.3.1	Moisture absorption capacity (MAC) .....	111
4.3.2	Water solubility (WS) .....	113
4.3.3	Water vapour permeability (WVP) .....	115
4.3.4	Contact angle (CA) .....	118
4.3.5	Weight percent gain (WPG) .....	122

4.4	Chemical characterisation of modified seaweed film .....	124
4.4.1	FT-IR characterisation of modified seaweed film.....	124
4.5	Preliminary studies optimum parameter .....	127
4.6	Physical characterization of modified and unmodified seaweed/CNF composite film.....	130
4.6.1	Moisture absorption capacity (MAC) .....	130
4.6.2	Water Solubility (WS).....	133
4.6.3	Water vapour permeability (WVP) .....	135
4.6.4	Contact angle (CA).....	138
4.6.5	Crystallinity index (XRD).....	141
4.7	Chemical characterization of modified and unmodified seaweed/CNF composite film.....	144
4.7.1	FT-IR characterization .....	144
4.8	Mechanical characterization of modified and unmodified seaweed/CNF composite film.....	146
4.8.1	Tensile strength (TS).....	146
4.8.2	Young modulus (YM) .....	149
4.8.3	Elongation at break (EAB).....	152
4.9	Morphological characterization of modified and unmodified seaweed/CNF composite film.....	154
4.9.1	Scanning electron microscopy (SEM).....	154
4.9.2	Atomic force microscopy (AFM).....	159
4.10	Thermal characterization of modified and unmodified seaweed/MCC composite film.....	162
4.10.1	Thermogravimetric analyzer (TGA) .....	162
4.11	Biodegradable characterization of modified and unmodified seaweed/CNF composite film.....	167
4.11.1	Soil-burial testing .....	167
4.12	Film applications .....	174

<b>CHAPTER 5</b>	<b>CONCLUSION AND RECOMMENDATIONS .....</b>	<b>177</b>
5.1	Conclusion.....	177
5.2	Recommendations for further works.....	179
<b>REFERENCES</b> .....		<b>180</b>
<b>LIST OF PUBLICATIONS</b>		



## LIST OF TABLES

	<b>Page</b>
Table 2.1	Types and varieties of <i>Kappaphycus alvarezii</i> in Sabah, Malaysia (Ling, 2014). ..... 14
Table 2.2	Chemical compositions of <i>Kappaphycus alvarezii</i> . ..... 17
Table 2.3	Incorporation of carrageenan/seaweed with other components. .... 42
Table 2.4	Macrofibril size and content of kenaf stem (Wambua et al., 2003). ..... 51
Table 2.5	Terminologies related to cellulose units (Sehaqui et al., 2011). ..... 71
Table 2.6	Comparison of nanocellulose with other selected fillers. .... 74
Table 3.1	List of chemicals. .... 84
Table 4.1	Droplet images and contact angle (°) of silane modified seaweed film. .... 119
Table 4.2	Droplet images and contact angle (°) of seaweed/CNF composite film. .... 139
Table 4.3	AFM and surface roughness of seaweed/CNF composite film. .... 160
Table 4.4	Major thermal decomposition step of seaweed/CNF composite films. .... 165
Table 4.5	Soil burial degradation of seaweed/CNF composite film. .... 170

## LIST OF FIGURES

	<b>Page</b>
Figure 2.1	Chemical structure carrageenans (Cunha & Grenha, 2016).....20
Figure 2.2	Double helix model of $\iota$ -carrageenan(Campo et al., 2009).....22
Figure 2.3	(a) $1C_4$ conformation of $\beta$ -D-galactoyranose (b) $4C_1$ conformation of $\beta$ -D galactoyranose (Campo et al., 2009). ....23
Figure 2.4	Essential steps involved in extraction of carrageenan powder and preparation of carrageenan fibers. (A. preparation of carrageenan powder; B. preparation of carrageenan fibers; C. preparation of carrageenan fibers with extraction.) (Dong et al., 2018).....25
Figure 2.5	Flow chart of refined-carrageenan (Hernandez-Carmona et al., 2013). ....26
Figure 2.6	Flow chart of semi-refined carrageenan production (Hernandez-Carmona et al., 2013). ....27
Figure 2.7	Classification of Biodegradable polymers (Doppalapudi et al., 2014). ....30
Figure 2.8	Structures of some polysaccharides (Chandra & Rustgi, 1998). .....34
Figure 2.9	Gelling mechanism for carrageenan in seaweed (Khalil et al., 2018). ....38
Figure 2.10	Gelling mechanism for carrageenan in seaweed (Khalil, Y. Tye, et al., 2017). ....39
Figure 2.11	Classification of natural fibres (Sanjay et al., 2018).....45
Figure 2.12	Kenaf plantation (Akil et al., 2011). ....49
Figure 2.13	Schematic picture of cell wall of the natural plans (Rouison et al., 2004). ....50
Figure 2.14	Schematic representation of the cellulose (Gupta et al., 2016).....53

Figure 2.15	Wood hierarchical structure and depiction of amorphous and crystalline components with cellulose, hemicellulose, and lignin chemical structures (Dufresne, 2013). .....	56
Figure 2.16	Schematic representation of extraction of nanocellulose from lignocellulosic biomass (Phanthong et al., 2018).....	57
Figure 2.17	Mechanism of silane surface treatment on seaweed/CNF biocomposite film. ....	66
Figure 2.18	Influence of surface energy and surface tension in wettability alteration (Jothi Prakash & Prasanth, 2021).....	81
Figure 3.1	Raw seaweed ( <i>Kappaphycus alvarezii</i> ).....	83
Figure 3.2	Experimental design flow chart. ....	85
Figure 3.3	The pulping, bleaching and isolation process of kenaf cellulose nanofiber. ....	87
Figure 3.4	Schematic presentation of biocomposite film fabrication.....	88
Figure 3.5	Contact angle analyser. ....	94
Figure 3.6	Fourier Transforms Infrared Spectroscopy (FT-IR). ....	95
Figure 3.7	Thermogravimetric analyzer (TGA). ....	98
Figure 4.1	TEM images of CNF isolated from kenaf bast with (a) 200 nm scale and (b) 100nm scale. The red arrows point to CNF indicate average fiber diameter size.....	100
Figure 4.2	FT-IR spectra for experimental kenaf CNF and commercial kenaf CNF.....	102
Figure 4.3	Kenaf bast CNF (a) Zeta potential value and (b) particle size distribution. ....	105
Figure 4.4	The XRD patterns experimental kenaf CNF and commercial kenaf CNF.....	108
Figure 4.5	FT-IR spectra of raw seaweed and kappa-carrageenan.....	110
Figure 4.6	MAC of silane modified seaweed film. ....	112
Figure 4.7	Water solubility of silane modified seaweed film.....	114

Figure 4.8	WVP of silane modified seaweed film.....	116
Figure 4.9	Silane modification effect on seaweed films contact angle. ....	121
Figure 4.10	Weight percent gain of silane modified seaweed film. ....	122
Figure 4.11	Separate FT-IR spectra for silane modified seaweed film. ....	125
Figure 4.12	Overlap FT-IR spectra for silane modified seaweed film.....	126
Figure 4.13	Effect of silane concentration on SF moisture resistance testing.....	128
Figure 4.14	Effect of silane treatment time on SF moisture resistance testing. ..	129
Figure 4.15	Moisture absorption capacity of seaweed/CNF composite film. ....	131
Figure 4.16	Water solubility of seaweed/CNF composite film. ....	133
Figure 4.17	Water vapour permeability of seaweed/CNF composite film.....	136
Figure 4.18	Crystallinity index of seaweed/CNF composite film. (a) Unmodified films, (b) silane modified films.....	142
Figure 4.19	FT-IR spectra of seaweed/CNF composite film.....	144
Figure 4.20	Overlap FT-IR spectra of seaweed/CNF composite film.....	145
Figure 4.21	Tensile strength of seaweed/CNF composite film. ....	147
Figure 4.22	Young modulus of seaweed/CNF composite film. ....	150
Figure 4.23	Elongation at break of seaweed/CNF composite film. ....	152
Figure 4.24	SEM images of unmodified seaweed/CNF composite film.....	156
Figure 4.25	SEM images of modified seaweed/CNF composite film.....	157
Figure 4.26	Thermogravimetric analyzer of seaweed/CNF composite film. (a) For unmodified seaweed composite film and (b) for modified seaweed composite films.....	163
Figure 4.27	Weight loss (%) of seaweed/CNF composite film.....	168

## LIST OF SYMBOLS

$\pm$	Plus-minus
$<$	Less than
$>$	Greater than
$^{\circ}\text{C}$	Degree Celsius
$^{\circ}$	Degree
$\%$	Percentage
$\text{mm}^2$	Millimetre square
Pa	Pascal
g	Gram
s	Second
$\text{m}^2$	Meter square
m	Meter
$\alpha$	Alfa
$T_m$	Melting temperature
$H_m$	Enthalpy of melting
$T_{\text{onset}}$	Initial degradation temperature
$T_{\text{max}}$	Maximum degradation temperature
$M_o$	Initial weight
$M_f$	Final weight
$l_o$	Original length
e	Extension
$\theta$	Tangent

## LIST OF ABBREVIATIONS

CA	Contact angle
CF	Cellulose fibres
CNC	Cellulose nanocrystals
CNF	Cellulose nanofibres
CNW	Cellulose nanowhiskers
EAB	Elongation at break
FT-IR	Fourier Transforms Infrared Spectroscopy
GPa	Gigapascal
HCl	Hydrochloric acid
KCl	Potassium chloride
KBr	Potassium bromide
MAC	Moisture absorption capacity
MCC	Microcrystalline cellulose
MPa	Megapascal
NCC	Nanocrystalline cellulose
RC	Refined carrageenan
RH	Relative humidity
Rpm	Revolutions per minute
SEM	Scanning electron microscopy
SF	Seaweed film
TGA	Thermogravimetric analysis
TS	Tensile strength
WS	Water solubility
WVP	Water vapour permeability
WVTR	Water vapour transmission rate
XRD	X-ray diffraction
YM	Young modulus

**PENCIRIAN FILEM RUMPAI LAUT HIDROFOBİK DIPERKUAT  
DENGAN SELULOSA GENTIAN NANO DARI KULIT KENAF**

**ABSTRAK**

Plastik amat kukuh dan tahan kepada proses biodegradasi dalam persekitaran semula jadi. Oleh itu pelbagai kajian telah dijalankan pada masa lalu untuk membangunkan filem dari sumber semula jadi. Dalam hal ini, filem berasaskan rumpai laut menunjukkan ciri-ciri fungsian yang sangat baik, jadi terdapat keperluan untuk menyiasat prestasi dan kesannya terhadap persekitaran untuk menentukan jangka hayat serta kesan pada persekitaran. Penggunaan nanoselulosa sebagai tetulang pengisi amat terkenal untuk meningkatkan sifat mekanikal filem. Sifat hidrofilik rumpai laut boleh diatasi dengan rawatan silane menggunakan triethoxymethyl silane untuk memberikan sifat hidrofilik kepada filem. Filem rumpai laut berasaskan nanoselulosa telah disediakan menggunakan proses tuangan pelarut. Dalam projek penyelidikan ini, siasatan telah dibuat untuk mengkaji sifat fiziko-kimia filem terbiodegradasi dan prestasinya. Kajian ini merangkumi pemahaman tentang sifat mekanikal, fizikal, kimia, haba, morfologi, biodegradasi dan hidrofilik filem. Kajian terperinci telah dijalankan untuk mengenal pasti ukuran optimum hidrofilik filem untuk memeriksa tahap rintangan pada lembapan. Jumlah nanoselulosa yang berbeza yang digabungkan dalam filem rumpai laut juga dikaji dan dibandingkan antara satu sama lain. Kadar interaksi air akan diukur dengan mengira pertambahan berat film yang selari dengan masa sewaktu kajian. Selain itu, maklumat tentang perubahan fiziko-kimia yang berlaku dalam sampel filem akibat rawatan silane di bawah keadaan terkawal telah diperoleh oleh analisis spektroskopi inframerah transformasi. Mengimbas analisis imej mikroskop elektron memberikan bukti yang sangat baik pada tahap mikroskopik yang

menjelaskan sifat morfologi filem. Juga dilaporkan bahawa kehabluran sangat mempengaruhi prestasi mekanikal oleh itu analisis pembelauan sinar-X bagi semua sampel telah disiasat. Projek ini membantu memahami dengan teliti terhadap kesan penggabungan nanoselulosa dan kesan rawatan silane ke atas prestasi filem rumpai laut. Filem rumpai laut berasaskan nanoselulosa yang diubah suai menggunakan silane telah berjaya direka dengan sifat hidrofobisiti yang baik tanpa mengurangkan sifat mekanikal dan fizikal filem tersebut.



# **CHARACTERIZATION OF HYDROPHOBIC SEAWEED FILM REINFORCED WITH KENAF BAST CELLULOSE NANOFIBERS**

## **ABSTRACT**

Plastics are durable and resistant to biodegradation processes in the natural environment. Intensive researches have been conducted in past to develop the film from the natural source. The seaweed-based film showed excellent functional properties, so there is a need to investigate its performance and impact on surroundings to determine its longevity and harmfulness. Using kenaf nanocellulose as filler reinforcement is well known to increase the mechanical properties of films. The hydrophilicity nature of the seaweed can be countered by silane treatment using triethoxymethyl silane to give hydrophobicity properties to the film. The nanocellulose based seaweed film were prepared using solvent casting process. In this research project, study on physicochemical properties of the biodegradable film and its performance has been completed. This study include the understanding of the mechanical, physical, chemical, thermal, morphology, biodegradability, and hydrophilicity properties of the film. Detail study has been carried out to identify the optimum measure of silane surface treatment on seaweed films hydrophobicity. Different amounts of nanocellulose incorporated within the seaweed film also has been studied and compared to one another. The rate of water interaction with the seaweed films were measure by calculate the weight increase with respect to time. Information about physicochemical changes that occurred in film samples due to silane treatment has also been determined by Fourier transform infrared spectroscopy analysis. Scanning electron microscopy image analysis provide very good evidence at the microscopic level in favor of film morphological properties. It was also reported that

crystallinity strongly affects mechanical performance based on X-ray diffraction analysis on films sample. This project contribute to understanding the effect of incorporating nanocellulose and the effect of silane treatment on seaweed films performance. The silane-modified nanocellulose based seaweed film has successfully fabricated to have good hydrophobicity properties without reducing the mechanical and physical properties of the film.

# CHAPTER 1

## INTRODUCTION

### 1.1 Overview

The development of a chemical procedure for creating synthetic polymers (plastics) from crude oil was a breakthrough in both chemistry and material sciences, and it led the way for the manufacture of one of the most versatile groups of materials ever created in the history of humanity. These novel materials combined characteristics such as strength, flexibility, lightweight, ease of manufacture, and cheap cost of manufacture (Sivan, 2011). However, it was discovered that these materials were exceedingly durable and were believed to be among the most non-biodegradable synthetic materials available. These characteristics made it possible to use plastics in practically every industrial, agricultural, or residential sector.

The handling of synthetic plastics is also one of the most serious problems facing the world today. It causes serious environmental problems through contaminated natural resources, such as water quality and soil fertility (Bhatnagar & Kumari, 2013). To date, more than 267 species in the marine environment have been affected by the entanglement or ingestion of synthetic plastic fragments (Derraik, 2002). Pollutants leached from synthetic plastics tend to accumulate in marine organisms that absorb or ingest them. This synthetic plastic pollutant poses a threat to the foundation of every food chain, and has a profound impact on non-marine species such as polar bears, seabirds, and humans who consumed the marine-grown resource (Hammer et al., 2012).

Data show that, due to population growth and huge demand, approximately 140 million tons of synthetic plastics are delivered globally every year (Rao et al.,

2014). Approximately 50% of synthetic plastics are used in packaging applications, and 90% of them are used as municipal waste (Sangawar & Gadre, 2013). Compared with other plastics such as polyvinyl chloride, polypropylene, and polystyrene, polyethylene such as low-density polyethylene (LDPE) and high-density polyethylene (HDPE) is one of the most commonly used synthetic plastics. It is estimated that the global production of polyethylene, polypropylene, polyvinyl chloride and polystyrene in 2012 will be 30%, 19%, 11% and 7% respectively (Pathak, 2017). The production of synthetic plastics is increasing, and its consumption is increasing at a rate of 12% per year (Koutny et al., 2006; Sivan, 2011). Plastic like polyethylene is the most widely used synthetic polymer, having a worldwide manufacturing capacity of around 140 million tonnes per year at the time of writing. Plastic trash accumulates in the environment due to a lack of effective techniques for properly disposing of it, providing an ever-growing ecological hazard to both land and aquatic wildlife (Shimao, 2001).

Naturally occurring biopolymers include starch, cellulose, gum, alginate, carrageenan, chitosan, and pectin. Among these, starch and cellulose are two of the most well-known polysaccharide-based biopolymers due to their widespread availability, abundance, and low cost (Hills, 2012). Seaweed is a versatile organism that generates a variety of polysaccharides including agar, carrageenan, and alginate. These polysaccharides are widely used in the development of biopolymers. Biopolymers generated from seaweed polysaccharides have several advantages, including their renewable nature, biodegradability, biocompatibility, and environmentally friendly (Jumaidin et al., 2018). Seaweeds (or marine macro-algae) are plant-like creatures that often reside in coastal locations adhering to rock or other hard substrates. Due to its high carbohydrate content, seaweed species have been

increasingly used industrially as a source of hydrocolloids (seaweed derivatives) in industries such as food technology, biotechnology, microbiology, and medicine, as well as the plastics industry (El-Said & El-Sikaily, 2013; Gade et al., 2013). Carrageenan is a naturally occurring polymer that is derived from edible red seaweeds of the *Rhodophyceae* family. It has been utilized as a thickener or gelling agent in a variety of sectors, including food, pharmaceuticals, and other industries (Zia et al., 2017).

There are a few disadvantages in using natural polymers such as carrageenan and cellulose. Because of their hydrophilic nature, these natural materials are only suitable for a limited range of applications (Thakur et al., 2013). Biopolymers must possess physico-chemical characteristics that are equivalent to those of synthetic polymers in order to be considered viable prospective substitutes for synthetic polymers in the marketplace (Thakur et al., 2014). The current study has thus been carried out by the use of chemical modification such as silane surface treatment in order to improve the films performance, especially in term of their hydrophobicity. Their mechanical performance, however, is often poor in terms of high-end applications, which might restrict their potential for usage. Plasticizers can be added to biopolymers to give them the workability they need in order to overcome the problem (Vieira et al., 2011). With further enhancement of the biopolymer, the performance of the resulted materials is comparable with the conventional synthetic polymer.

Nanocellulose as a reinforcing material in nanocomposites has been a significant area of study. Apart from its numerous benefits, such as affordable, low density, renewability, high specific properties, biodegradability, and relatively good

surface reactivity, nanocellulose exhibits superior properties as a reinforcing phase in nanocomposites when compared to micro- or macro-cellulose composites. The tailorability, design flexibility, and processability of nanocellulose–polymer composites enable their widespread use in a variety of sectors, including automotive, packaging, electronics, and biotechnology (Kargarzadeh et al., 2017). The term "nanocellulose" refers to nanocellulose-derived nanomaterials. Nanocellulose is classified into three basic types: nanocrystalline cellulose (NCC), nanofibrillated cellulose/cellulose nanofiber (CNF), and bacterial nanocellulose (BNC). Although these nanocelluloses are chemically identical, their shape, particle size, and crystallinity may vary owing to their varying sources and extraction techniques (Lavoine et al., 2012; Phanthong et al., 2018). CNF which is made by breaking cellulose fibres, is an example of a biodegradable, renewable nanofiller (biopolymer) with a minimal carbon footprint. Each year, plants produce around 75 billion tonnes of cellulose, making it a highly abundant resource (Feng, Cheng, et al., 2018).

Natural cellulosic fibres, like kenaf, have the potential to be used in polymeric composites in replacement of synthetic fibres. The increasing depletion of synthetic resources such as petroleum and increased awareness of global environmental concerns associated with synthetic materials all contribute to the acceptability of natural fibres as biocomposite components. Kenaf (*Hibiscus cannabinus L.*) is a versatile hibiscus species that is used to manufacture engineered timber, garments, packaging material, rope, and twine. Kenaf is mostly composed of cellulose (about 70%), which explains for its superior mechanical properties (Asyraf et al., 2021). Kenaf is a fast-growing plant that is very adaptable to its surroundings. It is one of the most traditional sources of natural cellulosic fibre. However, the high lignin concentration and tiny fibre size exclude its use in the manufacturing of textile fibres

and bioenergy refineries (Song et al., 2018). This study focuses on examining the properties of the used materials and the modified/unmodified composite films in relation to their water barrier and mechanical performance.

Packaging materials comprised of edible components are known as edible films. Due to their environmental friendliness, safety of consumption, and convenience of use, edible films are increasingly in demand nowadays (Maftoonazad & Badii, 2009). This is due to the fact that edible films can lengthen the quality, freshness, and shelf life of meals. The edible films cover the packed food product to form a semipermeable barrier, increasing its barrier characteristics by reducing moisture, lipid, gas, and volatile exchange (Kumar et al., 2021).

## **1.2 Problem statement**

*Kappaphycus alvarezii* is the world's fifth most farmed macro algae, thanks to its significant contribution in the production of carrageenan, which is extensively utilized in the food sector. However, *Kappaphycus alvarezii* is an underdeveloped biomass for green bio refinery conversion, further study is needed since it contains important proteins, lipids, and phenolic chemicals (Rudke et al., 2020). Carrageenan is a naturally occurring polymer produced from seaweed that has been extensively utilized as a food ingredient due to its appealing physicochemical features during the last decade (Liu et al., 2015). The extraction and purification of carrageenan are expensive, time intensive, and need a large amount of water and chemicals throughout the process (Hernandez-Carmona et al., 2013). Raw seaweed was utilized instead of purified carrageenan in the production of composite films due to its lower cost and ease of processing. Several prior investigations have shown the capability of fabricating composite films from raw seaweed (Khalil, Tye, Saurabh, et al., 2017).

Despite this, their low mechanical strength and poor barrier qualities have restricted their application possibilities.

Seaweed based films offer a number of benefits, including excellent transparency, good tensile strength, gelling ability, and film forming capabilities (Sánchez-García et al., 2010). Due to the rising need for raw materials throughout time, k-carrageenan is now mostly extracted from the cultivated species *Kappaphycus alvarezii* (Rhein-Knudsen et al., 2015). The use of carrageenan based film from seaweed as packaging material gave several benefit because of the suitable properties the materials had to offer especially being eco-friendly and biodegradable. However, in comparison to plastics, they have lower mechanical and water barrier properties, limiting their industrial use. As a result, research is being conducted to discover ways for optimizing the mechanical and barrier qualities of these biodegradable packaging materials (Savadekar et al., 2012). By utilizing the CNF as a filler, the functionality especially mechanical properties of the film can be enhanced. In comparison to other inorganic reinforcing fillers, CNF provide additional benefits, including a low ecological impact, low density, ease of recycling, and relative simplicity of processing owing to their nonabrasive nature (Azizi Samir et al., 2005; Podsiadlo et al., 2005). For hydro-soluble matrices, such as starch are used, the nanocomposites are usually formed by the casting process. Numerous researches have reported that solution cast nanocomposites based on starch and cellulose nanofibers exhibit excellent dispersion and overall performance (Angles & Dufresne, 2000; Chen et al., 2009; Mathew et al., 2008). Incorporating the CNF as a filler for carrageenan film also exhibit similar characteristic and properties because of their chemical compatibility shared by the carbohydrate.



The utilization of kenaf bast as the resource for the nanocellulose give several advantages. Kenaf is a fibrous plant, like wood and bamboo, is professionally grown as a fibre crop attributed to its beneficial properties (Khalil et al., 2010). Nanocellulose fibres have a high mechanical strength, a high strength-to-weight ratio, a huge surface area, and are very flexible. For these reasons, it's' become the focus of study and industrial use, as the novel advanced material promotes the development of promising biodegradable polymers for a variety of applications, including textiles, paper, engineering, medicine, and construction (Dufresne, 2013). Kenaf bast fibre is a non-woody lignocellulosic substance that is an excellent precursor material for CNF extraction (Kargarzadeh et al., 2012). Kenaf bast fibre is high in cellulose and has a low specific gravity also have superior properties as reinforcement agents (Anuar & Zuraida, 2011). The high specific strength and modulus of the fibre have attracted considerable attention because to its superior mechanical qualities, large specific surface area, and low coefficient of thermal expansion, including a high aspect ratio (Sulaiman et al., 2020).

The issue of having poor water barrier properties is a crucial task to be settle. Packaging materials are supposed to have a good barrier properties which become a challenge for bio-based material which usually very hydrophilic. Carrageenan is a water soluble polymer comprised of a linear chain of partly sulfonated galactans. These hydrocolloids are long-chain hydrophilic polymers (polysaccharides) that when dispersed in water create viscous dispersions and gels (Khalil, Tye, Saurabh, et al., 2017). Chemical modification can be made to increase the hydrophobicity of the material. Performing silane surface treatment using triethoxymethylsilane on carrageenan based film can improve the water barrier properties of the film. There are previous studies that have reported the fabrication of hydrophobic cellulose based

material by silane modifications which tend to improve the performance and properties of the material (Chen et al., 2015; Jiang & Hsieh, 2014; Jiménez-Saelices et al., 2017; Zhang et al., 2014). Hence, this study is designed to explore comprehensively on the optimum measure of silane surface treatment condition and different reinforcement filler loading quantity in producing better hydrophobic macro algae film on physical, mechanical, morphology, thermal and biodegradability properties for packaging applications.

### **1.3 Research Objectives**

The present research embarks on the following objectives:

- 1) To analyze the chemical properties of raw seaweed and cellulose nanofiber for biocomposite films fabrications.
- 2) To study the effect of silane surface treatment with different concentration and treatment period on the water barrier properties of seaweed films.
- 3) To investigate the effect of CNF loading on physical, mechanical, morphological, thermal and biodegradability properties of the modified and unmodified seaweed biocomposite films.

## 1.4 Thesis Layout

This thesis was dividing into six respective chapters, as shown below:

**Chapter 1: Introduction:** provide a summary of the research's background, justification, and objectives.

**Chapter 2: Literature review:** studied prior research and literature on a variety of topics relating to the overall area of the research.

**Chapter 3: Methodology:** described in detail the procedures and materials used in this study, as well as the experimental design. The production of nanocellulose, seaweed films and the silane surface treatment was discussed in detail. Techniques for characterizing raw materials and composite films were addressed in detail.

**Chapter 4: Results and Discussion:** provide the results of the investigation based on variety of scientific approaches and analysis for the raw materials, silane modification parameter and the modified/unmodified CNF seaweed film. Each study's results were carefully analyzed and discussed.

**Chapter 5: Conclusions and Recommendations:** summarized the research's overall findings and made positive future projections and suggestions for further research.

## CHAPTER 2

### LITERATURE REVIEW

#### 2.1 Macro algae

Macro algae or seaweed are found in marine ecosystems ranging from the arctic to tropical latitudes and from the intertidal zone to the euphotic zone's deepest depths. These algae provide vital ecological functions, including facilitating a variety of invertebrate recruitment processes, acting as autogenic ecosystem engineers by providing three-dimensional habitat structure, and providing critical structural strength to coral reef ecosystems. Calcified macro algae play a crucial role in carbonate deposition in coastal areas (Nelson, 2009). Apart from other biomass sources, algae produce a high amount of biomass per unit of light and area, may include a large amount of starch or oil, does not need fresh water or agricultural land, and can be sustained with either wastewater or ocean. Algae produce a diverse range of chemical compounds, most notably carbohydrates and lipids (Rajkumar et al., 2014).

Macro algae grow rapidly and may reach lengths of up to 60 m (McHugh, 2003b). Macro algae grow at a pace that is far faster than terrestrial plants. For example, brown algae biomass production ranged between 3.3 and 11.3 kilograms dry weight  $m^{-2} yr^{-1}$  in non-cultured algae and up to 13.1 kg dry weight  $m^2$  during a seven month period in cultured algae, compared to 6.1 to 9.5 kg fresh weight  $m^{-2} yr^{-1}$  in sugar cane, the most productive land plant (Kraan, 2013). They are only obtainable seasonally in natural water basins. Macro algae cultivation at sea, which does not need arable land or fertilizer, may provide a solution to the energy issue. macro algae are primarily used for food production and hydrocolloid extraction (Goh & Lee, 2010). In marine ecosystems, macro algae are at the bottom of the food chain and contribute between 30% and 50%

of the estimated net quantity of world oxygen through photosynthesis (Conti & Iacobucci, 2008).

Macro algae is a generic term that encompasses red algae (*Rhodophyta*), green algae (*Chlorophyta*), and brown algae (*Ochrophyta*). Although these algae are evolutionarily distinct, they are linked by the endosymbiotic processes (for which numerous hypotheses exist) that resulted in the formation of plastids (Lobban et al., 1994). Multicellular algae often grow vertically, toward the source of light, in order to generate larger and larger thalli and gather inorganic nutrients from a larger volume of water. Between the top of the mediolittoral belt and the greatest depth to which sufficient light (to fuel photosynthesis and growth) may penetrate, benthic microalgae reside attached to the rocky bottom by a basal holdfast (Lobban et al., 1994). Due to the absence of roots in algae, only a few species can survive on sedimentary substrates, such as certain siphonous green algae (e.g., *Halimeda*, *Caulerpa*, and *Udotea*) that develop root-like penetrating holdfasts that are also employed for nutrient absorption (Littler & Littler, 1988).

At the present, deterioration of benthic habitats is regarded as the greatest significant danger to micro algae biodiversity. It is well acknowledged that human-induced coastal changes, sediment loading, and seawater pollution have diminished the area of critical benthic habitats such as seagrass meadows and micro algae beds, hence impairing the ecosystem's functioning (Mangialajo et al., 2008; Stefani & Vincenzi, 2005; Turley, 1999). However, considerable variation in micro algae assemblages might be attributed to the interaction of human and natural ecological factors (Rinne et al., 2011). Marine plant and algae species are also critical for global carbon and nitrogen cycling, and they offer several ecosystem services, such as support for invertebrate and

fish populations that are consumed by humans (Conti & Iacobucci, 2008; Hemminga & Duarte, 2000).

Rhodophyta's red colour is due to the predominance of phycoerythrin and phycocyanin pigments, which obscure other pigments such as chlorophyll a, beta-carotene, and a number of xanthophylls. Coralline algae (orders Corallinales and Peyssonneliales) are a vast group of red algae that are capable of mineralizing the cell wall with calcium and, to a lesser degree, magnesium carbonate (Mann & Jahns, 1996). According to the six kingdoms of life (Cavalier-Smith, 2004), Rhodophyta and Chlorophyta are classified in kingdom Plantae, whilst Ochrophyta is classified in kingdom Chromista. Rhodophyta, or red algae, may be grown in temperate, subtropical, and tropical climates. *Eucheuma denticulatum*, *Kappaphycus alvarezii*, *Chondrus crispus*, and *Sarcothalia crispata* are the most significant red algae species for industrial use, since they are sources of carrageenan (Rudke et al., 2020). They represent a large array of species that are found in tropical, temperate, and cold-water coastal and continental shelf environments. Red algae are ecologically crucial because they are primary producers, offer structural habitat for other marine animals, and are critical in the formation and preservation of coral reefs (Kumar et al., 2008).





### **2.1.1 Kappaphycus alvarezii**

*Kappaphycus alvarezii*, more often referred to as "cottoni," is the primary source of kappa carrageenan, whereas *Eucheuma denticulatum*, more commonly referred to as "spinosum," is the primary source of iota carrageenan. Both species account for roughly 88 percent of raw material processed for carrageenan production, generating around 120,000 dry tonnes per year<sup>-1</sup>, with the majority of output coming from the Philippines, Indonesia, and Tanzania (McHugh, 2003b). *Kappaphycus* is a genus of red algae (Rhodophyta). There are six species of *Kappaphycus*, namely *Kappaphycus alvarezii*,

*Kappaphycus cottonii*, *Kappaphycus inermis*, *Kappaphycus malesianus*, *Kappaphycus procrusteanus* and *Kappaphycus striatus* (Liao, 1996; Nguyen & Huynh, 1995; Tan et al., 2014). The global production of *Kappaphycus alvarezii* for commercial extraction of kappa carrageenan reveals that the BIMP-EAGA region produces 96.5% of the total production, of which 55% comes from the Philippines, followed by Indonesia (38%) and Malaysia (2.5%) (Hayashi et al., 2010).

Over the last five decades, much attention has been paid to the red seaweed *Kappaphycus alvarezii* (Rhodophyta, Gigartinales, Areschougiaceae), a carrageenophyte of economic significance. According to reports, seaweed cultivation began in southern Mindanao in the latter part of the 1960s in the Philippines, adopting indigenous species selected from the wild (Krauss, 1977). *Kappaphycus alvarezii* provides for around 80% of Philippine seaweed exports, which are sold fresh and dried; although dried seaweed is more in demand, fresh seaweed is highly prized in restaurants (Hurtado, 2003). The Philippines is home to Asia's biggest carrageenan refinery, which facilitates the product's entry into other Asian nations such as Indonesia, Malaysia, Japan, China, India, and Vietnam. Farming continues to expand into new areas. At the mid-1990s, an Indian *Kappaphycus alvarezii* agriculture was established in Okha on the west coast with the purpose of producing kappa carrageenan (Mairh et al., 1995).

Table 2.1 Types and varieties of *Kappaphycus alvarezii* in Sabah, Malaysia (Ling, 2014).

Types/ Varieties	Processing method
<p data-bbox="325 338 863 376">"White" <i>Kappaphycus alvarezii</i> var. giant</p> 	<p data-bbox="890 432 1412 651">Seaweed was placed in a clear plastic bag and left it under the sun until it turns yellow-white colour. Then, the seaweed was further placed under the sun until constant weight obtained. The technique was known as "sauna- dried."</p>
<p data-bbox="325 748 863 786">"Purple" <i>Kappaphycus alvarezii</i> var. giant</p> 	<p data-bbox="890 869 1412 1014">Seaweed was hanged and dried under the sun until the seaweed turned into purplish colour. Then, the seaweed was dried to constant weight obtained.</p>
<p data-bbox="363 1144 863 1211">"Green" <i>Kappaphycus alvarezii</i> var. tambalang hijau</p> 	<p data-bbox="890 1283 1412 1429">This type of "green" seaweed was obtained by drying the seaweed directly under the sun until a constant weight achieved.</p>
<p data-bbox="331 1576 863 1615"><i>Kappaphycus alvarezii</i> var. green flower</p> 	<p data-bbox="890 1686 1412 1832">This type of "green" seaweed applied the same processing method as "white seaweed," but an artificial food-grade dye was mixed with this seaweed</p>



Locally, red seaweed *Kappaphycus alvarezii* is also referred to as "white," "purple," and "green" seaweeds, depending on their drying technique, as shown in Table 2.1 (Limantara & Brotosudarmo, 2015). *Kappaphycus alvarezii* may be classified according to their pigmentation into red, brown, and green forms. It is feasible to use the pigments to make food dyes that are safe for humans and provide considerable functional health benefits (Limantara & Brotosudarmo, 2015). Pharmaceutical substances such as fucoxanthin, carotenoid, zeaxanthin, and astaxanthin, polyphenols such as phenolic, flavonoids, and tannins are abundant in seaweed and have been shown to have significant antioxidant and anti-prostate cancer activities (Ling, 2014).

*Kappaphycus alvarezii* is a robust, dense, and hard marine algae/seaweed that may reach a height of two meters. Its thallus, a plant body that is not separated into root, stem, and leaf, but rather comprises of the holdfast, stipe, and blade, is coarse, with axes and branches ranging in diameter from 1-2 cm. Meanwhile, its principal axes are rather straight and devoid of secondary branches around their apices/tips. *Kappaphycus alvarezii* are one of the most abundant red algae in the tropics, growing at a fast rate that may double its biomass in 15 to 30 days (Trono Jr, 1992). These species may spread by vegetative fragmentation as well as sexual reproduction by spreading spores into the water column and allowing them to be carried by prevailing water currents (Azanza-Corrales et al., 1992). This species of alga may be reddish, yellowish, brown, or green in color, depending on the quantity of the phycoerythrin pigment. It is relatively suitable for the cultivation and grows rapidly, at a rate of around 4.5 percent daily (Gereniu et al., 2017).

This plant has been introduced internationally into a variety of maritime nations for experimental and commercial production as a viable alternative livelihood for

coastal communities. Along with the introduction, there is growing concern about the impact of species on the biodiversity of endemic habitats. The introduction of non-endemic cultivars required the adoption of quarantine techniques to prevent more invasive species (Bindu & Levine, 2011). These marine red algae, specifically *Kappaphycus alvarezii*, are economically significant because they generate the hydrocolloid carrageenan, which is widely used in the food, pharmaceutical, cosmetic, baking, and daily product industries due to its unique functional properties such as gelling, thickening, stabilizing, and water-holding capabilities (Gade et al., 2013; Rhein-Knudsen et al., 2015). This polysaccharide makes up the majority to 37% of the algae on dry base (Lechat et al., 1997). Concerns about *Kappaphycus alvarezii* non-endemic nature, impact on biodiversity, and disruption of local ecological community health and balance have been raised as a consequence of its worldwide introduction into new environments. Commercial invasive cultivars are gaining attention and are being seen as a possible threat to algae culture as a source of economic growth for coastal fishing communities (Barrios, 2005; Rodgers & Cox, 1999; Woo et al., 1989).

*Kappaphycus alvarezii* has a high concentration of minerals, vitamins, bioactive compounds, proteins, lipids, and polyphenols that have antibacterial, antiviral, antifungal, and other medical benefits (Hayashi & Reis, 2012). Carrageenan is the most abundant chemical component in *Kappaphycus alvarezii*, accounting for between 35.3 and 61.7 % of the total (de Góes & Reis, 2012). Kappa-carrageenan is obtained mostly from *Kappaphycus alvarezii*, whereas iota- and lambda-carrageenan are isolated from *Eucheuma*, *Gigartina*, and *Chondrus* species (Bixler & Porse, 2011). Carrageenan was originally obtained from wild red seaweed *Chondrus crispus*, which is still used in small amounts today (Abdou & Sorour, 2014). Over time, the increased demand for the raw material (carrageenan) resulted in the cultivation of the *Kappaphycus alvarezii* and

*Eucheuma denticulatum* species (McHugh, 2003a). *Kappaphycus alvarezii* is mostly used for the extraction of kappa-carrageenan, while *Eucheuma denticulatum* is used primarily for the extraction of iota-carrageenan (Cunha & Grenha, 2016).

Previous (Matanjun et al., 2009) reported a studies on the nutritional content of Malaysian *Kappaphycus alvarezii*, formerly known as *Eucheuma cottonii*, where its proximate, fatty acid, and amino acid compositions. The tendency of *Kappaphycus alvarezii* to absorb and accumulate trace elements in its tissue is well documented (Kumar et al., 2007). This indicates that the mineral and trace element composition of micro propagated *Kappaphycus alvarezii* may differ from that of field-grown specimens. (Graham, 2009) discovered that the amount of compounds contained in seaweeds might vary depending on a variety of parameters such as the season, the age of the population, the species, the geographic location, and the temperature. Table 2.2 shows the differences in chemical compositions in *Kappaphycus alvarezii* based on prior investigations.

Table 2.2 Chemical compositions of *Kappaphycus alvarezii*.

Composition(wt %)	<i>Kappaphycus alvarezii</i> (%)	<i>Kappaphycus alvarezii</i> (%)	<i>Kappaphycus alvarezii</i> (%)	References
Carbohydrate	59.58	57.30	65.20	(Abd-Rahim et al., 2014) (Abirami & Kowsalya, 2011) (Abdul Khalil et al., 2018)
Protein	5.74	4.5	3.40	
Lipid	0.75	0.89	1.10	
Ash	19.70	28.9	11.57	

Generally, the red seaweeds *Kappaphycus alvarezii*, *Eucheuma denticulatum*, and *Betaphycus gelatinae* are the three most important red seaweeds for the manufacture of commercial carrageenan (Kılınç et al., 2013). Nonetheless, some species from the *Gigartina*, *Iridae*, *Hypnea*, and *Furcellaria* genera have also been

utilised, resulting in the production of several kinds of carrageenan extracts from these plants (De Ruiter & Rudolph, 1997; McHugh, 2003a). *Chondrus crispus* is less commercially viable than *Kappaphycus alvarezii* because they tend to generate a combination of kappa and lambda carrageenan, which is difficult to extract and separate from the rest of the carrageenan mixture (Cunha & Grenha, 2016). The decline of gel strength that occurs when either kappa or iota carrageenan is mixed with lambda carrageenan occurs throughout applications, resulting in a weaker gel that is less suited for industrial applications (McHugh, 2003b).

A strong demand for carrageenan has resulted in the expansion of the cultivation of the seaweed *Kappaphycus alvarezii* in Malaysia, which has led in the cultivation of the seaweed. In response to rising demand for *Kappaphycus alvarezii*, a shortfall of seedling supply has emerged while micro propagation is one of the possible solutions (Yong et al., 2015). Since 1978, the cultivation of *Kappaphycus alvarezii* has been established in East Malaysia, specifically in Sabah (Sade et al., 2006). Since then, seaweed has emerged as one of the most commercially valuable natural resources in the country. Meanwhile, two plants for semi-refined carrageenan manufacture have been founded in Semporna and Tawau, both in the Sabah province. Also in Peninsular Malaysia, the cultivation of *Kappaphycus* (of the tambalang variety) has been introduced and successfully established (Phang et al., 2010).

## **2.2 Chemical and rheological properties of carrageenan**

Carrageenan, formerly referred to as 'carrageenin,' was discovered in 1862 by British chemist Stanford, who isolated it from Irish moss (*Chondrus crispus*). Later, the name was altered to carrageenan to conform to the '-an' suffix required for polysaccharide names (Pereira, 2011). Carrageenan, a polysaccharide derived from

marine algae, is rapidly gaining recognition as a potentially renewable biomaterial with significant promise as a replacement for current synthetic plastics. Carrageenan-derived materials have been extensively researched in recent decades for applications ranging from pharmacological and biomedical to culinary films and coatings. Carrageenans have limitations in the field of flexible films, principally due to its inherent hydrophilicity (Sedayu et al., 2019).

Carrageenan is a hydrophilic linear sulfated galactan found only in the cell walls of red sea algae (Rhodophyceae). It is derived predominantly from the genera *Chondrus*, *Eucheuma*, *Gigartina*, and *Iridaea* (Jouanneau et al., 2010). Carrageenan has a rather large molecular weight, ranging between 100 and 1000 kDa on average. Along with the ester sulphate and galactose groups that form the primary backbone of the structure, it contains other polysaccharide residues and substituents such as glucose, xylose, uronic acids, methyl ethers, and pyruvate groups (Aldalbahi, 2012). Carrageenan is a hydrocolloid mostly composed of potassium, sodium, magnesium, and calcium salts of sulfated galactose esters and copolymers of 3,6-anhydro-galactose (De Ruiter & Rudolph, 1997). Carrageenan is a sulfated polygalactan that contains between 15% and 40% ester-sulfate and an average relative molecular mass of well over 100 kDa. It is composed of alternating units of d-galactose and 3,6-anhydro-galactose (3,6-AG), which are linked by 1,3 and 1,4-glycosidic linkages.

Carrageenan is categorized into six basic forms: Iota ( $\iota$ -), Kappa ( $\kappa$ -), Lambda ( $\lambda$ -), Mu ( $\mu$ -), Nu ( $\nu$ -) and Theta ( $\theta$ -) carrageenan. This nomenclature is essential both chemically and commercially, since the various carrageenans subtypes are isolated from different weed sources (Campo et al., 2009). This categorization is based on the fact that it is soluble in potassium chloride. The key changes affecting the carrageenan type's

characteristics are the quantity and location of ester sulphate groups, as well as the amount of 3,6-AG (Necas & Bartosikova, 2013). These designations do not refer to particular chemical structures but rather to generic changes in the composition and degree of sulfation at various sites within the polymer. The sulphate groups on the disaccharide repeating unit indicate the 3 major carrageenan type: lambda ( $\lambda$ ), kappa ( $\kappa$ ), and iota ( $\iota$ ) (Al-Alawi et al., 2011) as depicted in Figure 2.1 below. Increased ester sulphate content results in a lower solubility temperature and a weaker gel. Carrageenan of the kappa type contains around 25%–30% ester sulphate and approximately 28–35% 3,6-AG. Carrageenan of the iota type contains around 28 to 30% ester sulphate and approximately 25 to 30% 3,6-AG. Lambda-type carrageenan contains about 32–39% ester sulphate and no 3,6-AG (Barbeyron et al., 2000).

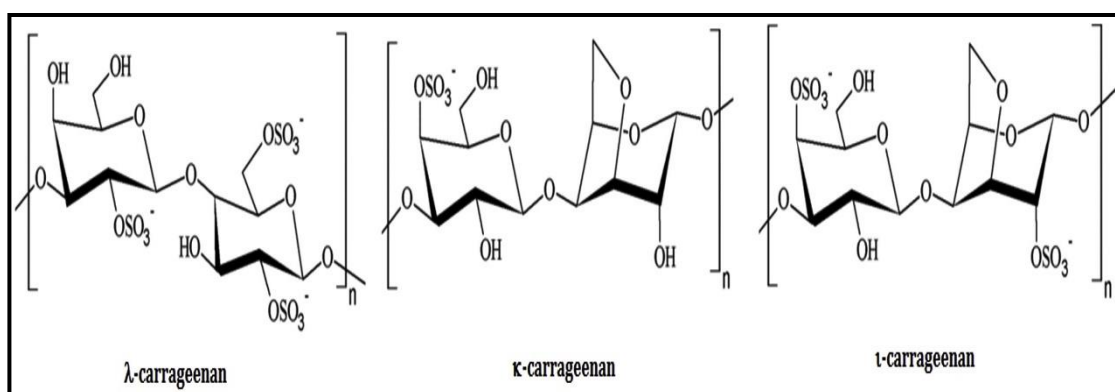


Figure 2.1 Chemical structure carrageenans (Cunha & Grenha, 2016).

Kappa ( $\kappa$ )-carrageenan is a polysaccharide comprised of alternating 3-linked  $\beta$ -D-galactose 4-sulfate and 4-linked 6-anhydro- $\alpha$ -galactopyranose is a disaccharide repeating unit with one negative charge (Campo et al., 2009). Each disaccharide repeating unit of Iota ( $\iota$ )-carrageenan contains two sulphate groups (Pavli et al., 2011). Iota and kappa carrageenan exhibit similar properties, including the ability to undergo thermoreversible conformational rearrangement in aqueous solution at high temperatures, while combining and forming networks at lower temperatures via

sulphate groups and the 3,6-anhydro-d-galactopyransyl ring (Gu et al., 2005). Additionally,  $\iota$  and  $\kappa$  carrageenans exhibit gelling capabilities in the presence of cations, which is affected by the concentration and type of the cations (i.e.  $K^+$  and  $Ca^{2+}$ , respectively) in the solution, as well as the concentration of the biopolymer (Uruakpa & Arntfield, 2004). Additionally, various salts have different impacts on the phase transitions and gelation of  $\iota$ - and  $\kappa$ -carrageenan gels. It has been observed that in the presence of KCl, stronger  $\kappa$ -carrageenan gels were formed compared in the presence of the other salts, particularly NaCl,  $MgCl_2$ , LiCl,  $SrCl_2$ , and  $CaCl_2$  (Kara et al., 2006). The gel-sol transition temperatures of  $\kappa$ -carrageenan were strongly related to KCl, NaCl, and  $CaCl_2$  contents (Pekcan & Tari, 2008).

Lambda ( $\lambda$ )-carrageenan contains three sulphate groups per disaccharide unit, but unlike  $\kappa$  and  $\iota$ -carrageenan, it lacks a 3, 6-anhydride bridge (Pavli et al., 2011). Throughout all temperatures,  $\lambda$ -carrageenan is incapable of forming gel and has a random coil shape (Gu et al., 2005). The addition of salts may reduce the viscosity of a solution by lessening the electrostatic repulsion between the sulphate groups, which is permitted by an increase in the concentration of macromolecules in the solution. For instance, this is what occurs for lambda fraction (Campo et al., 2009). In the presence of sulphate groups orientated towards their exterior section, the kappa- and iota-carrageenans form a network of three-dimensional double helices, which is formed by the "crosslinking" of adjacent spiral chains that include sulphate groups directed towards their external part as shown in Figure 2.2. The 2-sulfate group of  $\lambda$ -carrageenan is directed towards the interior half of the molecule, preventing this "crosslinking" from occurring (Morais et al., 1989).

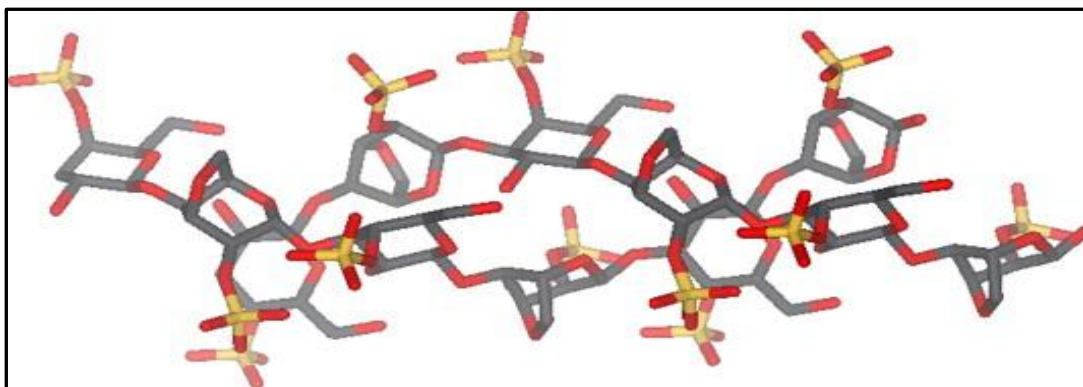


Figure 2.2 Double helix model of  $\iota$ -carrageenan(Campo et al., 2009).

Along with the distinct carrageenan kinds discussed above, there are also various hybrid carrageenan types to consider. In nature, seaweeds do not often generate pure carrageenan, but rather a variety of diverse sulfated polysaccharides in their structures, including hybrid carrageenans, which are a kind of sulfated polysaccharide (van de Velde, 2008). The amount of carrageenans present in a particular seaweed is governed by a variety of variables, including the seaweed's biological stage (Gómez-Ordóñez & Rupérez, 2011), gender of the gametophyte, environmental conditions, as well as the extraction process used in the recovery of carrageenan (Hilliou, 2014; Larotonda et al., 2016; Souza et al., 2011). The vegetative phase of the Gigartinales seaweed order generates highly sulfated forms of  $\mu$ -carrageenan, which are non-gelling, but the reproductive stage produces  $\kappa$ - and  $\iota$ -carrageenans that are gelling, as well as  $\nu$ - and  $\eta$ -carrageenans. Additionally, the immature seaweed of *Kappaphycus striatum* has less gelling carrageenan types than the matured seaweed (Hilliou, 2014).

Regarding carrageenan's rheological characteristics, all three commercially significant hydrocolloids exhibited significantly diverse features. Kappa- and iota-carrageenan shared the characteristic structure of a 4-linked galactose residue's anhydro-galactose bridge, which adopts the  ${}^1C_4$  – chair conformation, as seen in Figure



2.3(a). This conformation is critical for the production of helical structures and gels (Campo et al., 2009). Also non-gelling carrageenans are the naturally occurring precursors (Mu- and Nu-carrageenan) of kappa- and iota-carrageenan, which include D-galactopyranosil units in the  ${}^4C_1$  conformation as shown in Figure 2.3(b), which are non-gelling carrageenans. The occurrence of disaccharide units without the 3, 6-anhydro ring and possessing a  ${}^4C_1$  conformation, according to (Campo et al., 2009), results in twists in the regular chains, which eventually prevents the development of helical strands. Carrageenan is prevented from gelling as a result of this effect.

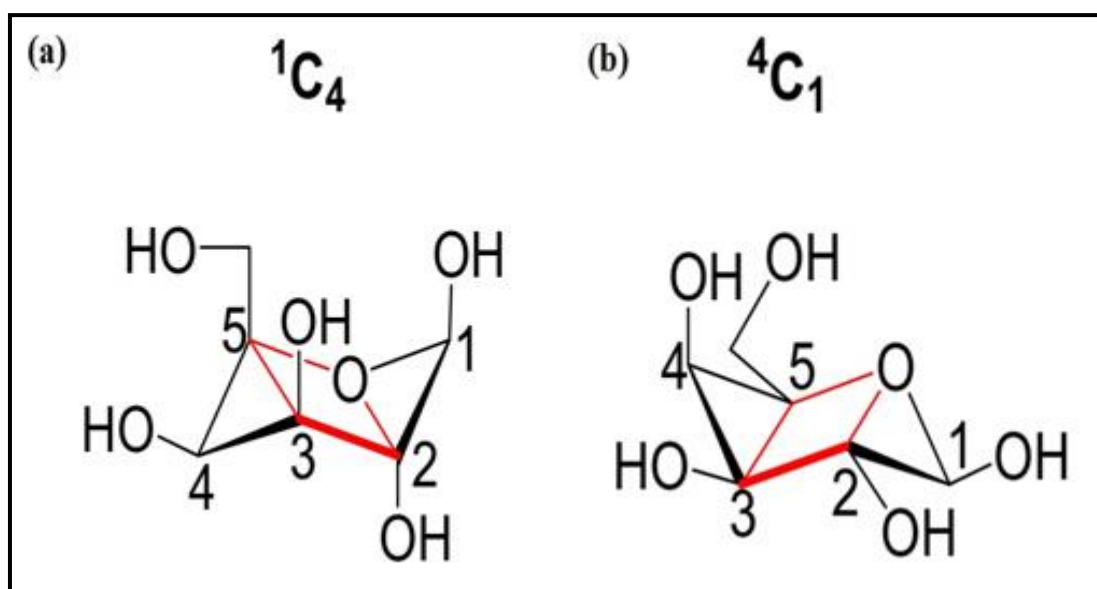


Figure 2.3 (a)  ${}^1C_4$  conformation of  $\beta$ -D-galactopyranose (b)  ${}^4C_1$  conformation of  $\beta$ -D-galactopyranose (Campo et al., 2009).

The conformations ( ${}^1C_4$  and  ${}^4C_1$ ) shown in Figures 2.3(a) and 2.3(b) are defined by the use of numbers to indicate ring atoms that are either above or below a specified reference plane (red) that contains the carbon atoms C-2, C-3, C-5, and O. Figure 2.5 (a) shows that the C-1 is positioned above the reference plane and the C-4 is placed below it, resulting in the symbol  ${}^1C_4$  being used. Meanwhile, in Figure 2.5(b), the C-4 plane is located above the reference plane and the C-1 plane is located below the plane, resulting in the sign  ${}^4C_1$  for the C-4 plane. There is a possibility that the carrageenan's

conformations contribute to the rheological characteristics of the substance in distinct ways.

### **2.3 Extraction of carrageenan from seaweed**

The extraction of seaweed hydrocolloids is a time-consuming and complex procedure that involves many phases of processing in order to be effective. Due to the water soluble nature of ordinary seaweed hydrocolloids, hot water is often the most preferred solvent for their extraction, with the exception of alginate/alginate acid extractions, which need hot alkali as the primary solvent (Minghou, 1990). Traditionally, carrageenan powder was obtained from red seaweed by following the stages of impurity removal, hot water extraction, and drying to get a fine powder as shown in figure 2.4. The primary disadvantage of this approach is that an extraction solvent is required during the extraction process to precipitate the carrageenan, and then the precipitate is dried and crushed to get carrageenan powder. Carrageenan powder must be dissolved prior to the manufacturing of carrageenan fibres. Adding the solvent during the extraction process and dissolving the carrageenan powder during the preparation of carrageenan fibre uses a significant quantity of water, ethanol, and energy (Dong et al., 2018).