# EVALUATION OF ANTI-HYPERTENSIVE AND VASORELAXANT EFFECTS OF GYNURA PROCUMBENS MERR. EXTRACTS

# ZAFAR IQBAL

# **UNIVERSITI SAINS MALAYSIA**

2020

# EVALUATION OF ANTI-HYPERTENSIVE AND VASORELAXANT EFFECTS OF GYNURA PROCUMBENS MERR. EXTRACTS

by

# ZAFAR IQBAL

Thesis submitted in fulfillment of the requirements for the degree of Doctor of Philosophy

February 2020

Dedicated To my Family and Teachers Whom I love the most

#### ACKNOWLEDGEMENT

All praises for the Almighty Allah. I would like to express my deep gratitude to Professor Dr. Mohd. Zaini Asmawi and Dr. Yam Mun Fei for their helping attitude. I have learned a great deal under your guidance, and I appreciate all the challenging works you have put into helping me complete this degree. You have been supportive, understanding, and positive. Thank you very much for making my time in the lab extremely fulfilling.

I would also like to thank Professor Dr. Amirin Sadikun and Associate Professor Mariam Binti Ahmad for their constant assistance and patience with me. I am grateful for your encouragement, advice, and support.

I am very grateful to all my colleagues and friends for their encouragement and kind support. Last but not the least my family has been there sound and firm at my back. I have no words to express thanks to them. My father Malik Gul Muhammad (late), my mother Kaneez Fatima (late) and my sister Zahida Perveen (late) who dreamt this day their whole life but today cannot see their dream coming true. May Allah bless them the highest place in heaven.

I would like to thank my wife, Arifa Khan for all his patience and encouragement. Finally, I would like to thank my children, Fakiha Zafar and Anaiba Zafar for being such loving and independent, enabling me to complete this work.

Zafar Iqbal

ii

## **TABLE OF CONTENTS**

| ACK            | NOWLEDG     | EMENT                                    | ii   |
|----------------|-------------|--|------|
| TABI           | LE OF CON   | ITENTS                                   | iii  |
| LIST OF TABLES |             |  | viii |
| LIST           | OF FIGUR    | ES                                       | ix   |
| LIST           | OF ABBRE    | EVIATIONS                                | xvi  |
| ABST           | <b>'RAK</b> |  | xx   |
| ABST           | RACT        |  | xxii |
| CHAI           | PTER 1 IN   | TRODUCTION                               | 1    |
| 1.1            | Backgrou    | and                                      | 1    |
| 1.2            | Problem     | statement                                | 1    |
| 1.3            | Hypothes    | sis                                      | 2    |
| 1.4            | Objective   | es                                       | 2    |
| CHAI           | PTER 2 LI   | TERATURE REVIEW                          | 4    |
| 2.1            | Hyperten    | sion                                     | 4    |
| 2.2            | The Card    | liovascular system                       | 5    |
|                | 2.2.1       | The Heart                                | 5    |
|                | 2.2.2       | Arteries and arterioles                  | 7    |
| 2.3            | Vascular    | Smooth Muscle Contraction and Relaxation | 8    |
|                | 2.3.1       | Endothelium                              | 10   |
| 2.4            | Enzyme-     | Linked Receptors                         | 15   |
|                | 2.4.1       | Soluble Guanylyl Cyclase pathway         | 16   |
|                | 2.4.2       | Serine-Threonine Protein Kinases         | 16   |
| 2.5            | G-Protein   | n-Coupled Receptors (GPCRs)              | 17   |
|                | 2.5.1       | Gqα-Protein-Coupled Receptors            | 18   |
|                | 2.5.2       | Gia-Protein-Coupled Receptors            | 19   |
|                | 2.5.3       | Gsα-Protein-Coupled Receptors            | 19   |

| 2.6   | Channel           | -Linked Receptors                             | 19 |
|-------|-------------------|---|----|
|       | 2.6.1             | Potassium Channels                            | 20 |
|       | 2.6.2             | Calcium Channels                              | 23 |
| 2.7   | Regulati          | ion of Blood Pressure                         | 26 |
| 2.8   | Autonor           | nic nervous system (ANS)                      | 28 |
|       | 2.8.1             | Sympathetic nervous system                    | 29 |
|       | 2.8.2             | α-Adrenoceptors                               | 30 |
|       | 2.8.3             | β-adrenoceptors                               | 32 |
| 2.9   | Renin -           | Angiotensin System (RAS)                      | 34 |
|       | 2.9.1             | Angiotensin receptors                         | 36 |
|       | 2.9.2             | Local (Tissue) RAS                            | 37 |
|       | 2.9.3             | Kidney RAS                                    | 37 |
| 2.10  | Convent           | tional antihypertensive drugs                 | 38 |
| 2.11  | Herbal r          | nedicine                                      | 42 |
|       | 2.11.1            | Investigation of traditional medicinal plants | 43 |
|       | 2.11.2            | Medicinal plants for cardiovascular disorders | 44 |
| 2.12  | Classific         | cation of <i>G. procumbens</i> :              | 50 |
| СНАРТ | TER 3 N           | IATERIALS AND METHODS                         | 51 |
| 3.1   | List of t         | ools and equipment                            | 51 |
| 3.2   | List of chemicals |   | 52 |
| 3.3   | Study design      |   | 53 |
| 3.4   | Plant Materials   |   | 55 |
| 3.5   | Preparat          | ion of crude extracts                         | 55 |
| 3.6   | Preparat          | ion of drugs and solutions                    | 56 |
|       | 3.6.1             | For In vitro studies                          | 56 |
|       | 3.6.2             | For <i>in vivo</i> studies                    | 57 |

|      | 3.6.3  | Preparation of ursolic acid standard and GPPE<br>samples for High Performance Liquid<br>Chromatography analysis | 57 |
|------|--|---|----|
|      | 3.6.4  | Preparation of reagent and extract for Antioxidant assays   | 57 |
| 3.7  | Experim  | nental animals  | 58 |
| 3.8  | Vasorela   | axant activity of G. procumbens   | 58 |
|      | 3.8.1  | Preparation of isolated rat aortic rings  | 58 |
|      | 3.8.2  | Vasorelaxation by G. procumbens extracts  | 60 |
|      | 3.8.3  | Vasorelaxation mechanism of <i>G. procumbens</i> extracts in rat aortic ring preparations                       | 61 |
|      | 3.8.4  | Involvement of endothelium-dependent pathways   | 61 |
|      | 3.8.5  | Effect of potassium channel blockers  | 62 |
|      | 3.8.6  | Involvement of $\beta$ -adrenergic receptors  | 62 |
|      | 3.8.7  | Inhibition of extracellular influx through the L-type $Ca^{2+}$ channel   | 62 |
| 3.9  | Acute to   | exicity study in the rat  | 63 |
| 3.10 | Anti-hyj   | Anti-hypertensive studies of <i>G. procumbens</i> extracts in SHR   |    |
| 3.11 | 11 Blood pressure measurements in anesthetized normotensive rats |   | 66 |
|      | 3.11.1   | Effects of <i>G. procumbens</i> extracts in anesthetized normotensive rats                                      | 67 |
|      | 3.11.2   | Effect on cardiovascular muscarinic receptors   | 67 |
|      | 3.11.3   | Effect on cardiovascular $\beta$ -adrenergic receptors  | 68 |
|      | 3.11.4   | Effect on cardiovascular $\alpha$ -adrenergic receptors   | 68 |
| 3.12 | Termina  | tion of the experiment  | 68 |
| 3.13 | Phytoch  | emical Analysis   | 69 |
|      | 3.13.1   | GC-MS analysis of petroleum ether extract   | 69 |
|      | 3.13.2   | High-performance liquid chromatography analysis   | 69 |
| 3.14 | Antioxidant assays   |   | 70 |

|       | 3.14.1                       | Total phenolic content  | 70 |
|-------|------------------------------|---|----|
|       | 3.14.2                       | Total flavonoid content   | 70 |
|       | 3.14.3                       | 2,2-diphenyl-1-picrylhydrazyl (DPPH) assay  | 71 |
|       | 3.14.4                       | 2,2'-azino-bis (3-ethylbenzothiazoline-6-sulphonic acid) or ABTS radical scavenging assay   | 71 |
|       | 3.14.5                       | Ferric reducing antioxidant power (FRAP) assay  | 71 |
| 3.15  | Statistic                    | cal analysis  | 71 |
| CHAPT | rer 4 r                      | RESULTS   | 73 |
| 4.1   | Endothe                      | elium-intact isolated aortic ring preparation   | 73 |
| 4.2   | Endothe                      | elium-denuded isolated aortic ring preparation  | 74 |
| 4.3   | The vas                      | orelaxation effects of G. procumbens extracts   | 74 |
| 4.4   | The effective endothe        | ects of petroleum ether extract of <i>G. procumbens</i> in elium-intact isolated aortic ring preparation  | 76 |
| 4.5   | The eff<br>endothe           | fects of chloroform extract of <i>G. procumbens</i> in elium-intact isolated aortic ring preparation  | 77 |
| 4.6   | The effection of the endothe | fects of methanol extract of <i>G. procumbens</i> in elium-intact isolated aortic ring preparation  | 78 |
| 4.7   | The ef<br>endothe            | ffects of water extract of <i>G. procumbens</i> in elium-intact isolated aortic ring preparation  | 79 |
| 4.8   | The eff<br>endothe           | fects of <i>G. procumbens</i> extracts and fractions in elium-intact isolated aortic ring preparation   | 80 |
|       | 4.8.1                        | The effects of petroleum ether extract of <i>G</i> . <i>procumbens</i> in endothelium-intact isolated aortic ring preparation                       | 81 |
|       | 4.8.2                        | The effects of acetone-insoluble fraction of petroleum ether extract of <i>G. procumbens</i> in endothelium-intact isolated aortic ring preparation | 82 |
|       | 4.8.3                        | The effects of acetone-soluble fraction of petroleum ether extract of <i>G. procumbens</i> in endothelium-intact isolated aortic ring preparation   | 83 |
| 4.9   | Vasorel                      | axation mechanisms of GPPE  | 84 |
|       | 4.9.1                        | The effect of GPPE on endothelium-intact and endothelium-denuded rat aortic ring preparations   | 84 |

|       | 4.9.2           | Endothelium-dependent vasorelaxation mechanisms<br>of GPPE in endothelium-intact rat aortic ring<br>preparations in the presence of antagonists                   | 86  |
|-------|-----------------|---|-----|
|       | 4.9.3           | Endothelium-independent vasorelaxation<br>mechanisms of GPPE in endothelium-denuded rat<br>aortic ring preparations in the presence and absence<br>of antagonists | 90  |
|       | 4.9.4           | The effect of potassium channel blockers on GPPE induced vasorelaxation in endothelium-denuded rat aorta  | 94  |
|       | 4.9.5           | The effect of GPPE on Calcium Channels  | 100 |
| 4.10  | Phytoch         | nemical analysis of G. procumbens   | 105 |
|       | 4.10.1          | Gas Chromatography-Mass Spectrometry (GC-MS)  | 105 |
| 4.11  | Antioxi         | dant assay  | 108 |
| 4.12  | High-Pe         | erformance Liquid Chromatography of GPPE  | 109 |
| 4.13  | The <i>in</i> - | vivo effects of G. procumbens   | 112 |
|       | 4.13.1          | Acute oral toxicity of G. procumbens  | 112 |
|       | 4.13.2          | The effects of daily oral administration of <i>G</i> . <i>procumbens</i> extracts on systolic blood pressure of Spontaneously Hypertensive Rats (SHR)             | 114 |
|       | 4.13.3          | Effects of <i>G. procumbens</i> extracts on blood pressure of anesthetized rats   | 117 |
|       | 4.13.3(a        | a) Effect on muscarinic receptors   | 119 |
|       | 4.13.3(t        | b) Effect on $\beta$ -adrenergic receptors  | 121 |
|       | 4.13.3(0        | Effect on $\alpha$ -adrenergic receptors  | 123 |
| CHAP  | TER 5 D         | DISCUSSION  | 126 |
| CHAP  | FER 6 C         | CONCLUSION  | 152 |
| REFER | RENCES          |   | 154 |

## LIST OF TABLES

| Table 2.1 | Endothelium-derived vasoactive factors. Summary of<br>major vasoactive factors (dilators and constrictors)<br>found in endothelium, their synthesizing enzyme,<br>target, the effect on tone and mechanism of action  |
|-----------|---|
| Table 4.1 | Main phytocomponents related to the vasorelaxant activity in petroleum ether extract of <i>G. procumbens</i> identified by GC-MS  |
| Table 4.2 | Chemical analysis of GPPE showed the presence of phenolic compounds, flavonoids, free radical scavenging activity and total antioxidant activity. Values are expressed as mean $\pm$ SEM ( $n = 3$ )  |
| Table 4.3 | Limit of detection, Limit of quantification and Linearity<br>of standard curves for ursolic acid  |
| Table 4.4 | Effect of a single dose of 2000 mg/kg of <i>G.</i><br><i>procumbens</i> crude extract on the physiological<br>parameters of rats. The - sign indicates the absence of<br>toxic signs. Whereas + Sign shows regular activity.<br>Weights are presented as Mean $\pm$ SEM (n = 5) 113 |

## LIST OF FIGURES

| Figure 2.1  | The cardiovascular system  |
|-------------|--|
| Figure 2.2  | Structure and layers of a typical blood vessel in the circulatory system   |
| Figure 2.3  | Mechanisms, by which contraction of vascular smooth muscle takes place   |
| Figure 2.4  | The production, of the endothelial nitric oxide and its effects in the cells of vascular smooth muscle   |
| Figure 2.5  | Flow chart diagram describe the vasodilation by the production of $PGI_2$ and NO in the endothelial cell   |
| Figure 2.6  | The signalling pathways in vascular endothelium and vascular smooth muscle acts synergistically to maintain the vasocontractions and vasorelaxations in the blood vessels  |
| Figure 2.7  | The role of Enzyme-Linked Receptors in vasorelaxation  |
| Figure 2.8  | G protein-coupled receptor signalling 18   |
| Figure 2.9  | Potassium channels are controlling the tone of the vascular smooth muscle cells  |
| Figure 2.10 | Function of the potassium channel in the blood vessels   |
| Figure 2.11 | Ion channels and the blood vessel  |
| Figure 2.12 | Types of Voltage-gated calcium channels (VGCCs) or VOCC  |
| Figure 2.13 | Types of Ligand-gated calcium channels (ROCC) 25   |
| Figure 2.14 | Regulation of Blood Pressure   |
| Figure 2.15 | Mechanisms for regulating mean arterial blood pressure   |
| Figure 2.16 | Mechanism of action of $\beta$ - adrenoceptor blockers and<br>pathways through which beta-adrenergic receptor ( $\beta$ -<br>AR) antagonists exert beneficial effects in<br>hypertension. 34   |
| Figure 2.17 | Effects of combined stimulation of renal angiotensin II<br>(Ang II) $AT_1 (AT_1R)$ receptors, vasopressin (AVP) $V_1a$<br>(V1aR) and V2 (V <sub>2</sub> R) receptors, and aldosterone (Aldo)<br>receptors (MR) on body fluid volume and pressure |

| Figure 3.1 | A flow chart describing the preparation of <i>G</i> . <i>procumbens</i> crude extracts  | 53 |
|------------|---|----|
| Figure 3.2 | A Flow chart describing the overall study design  | 54 |
| Figure 3.3 | The preliminary evaluation of the antihypertensive effect of <i>G. procumbens</i> extracts.   | 65 |
| Figure 3.4 | Anaesthetized animal experimental design in normotensive rats   | 66 |
| Figure 4.1 | Representative traces of isometric force recordings of 1 $\mu$ M phenylephrine - induced vasocontraction and 1 $\mu$ M acetylcholine - induced vasorelaxation in endothelium - intact aortic ring preparations.   | 73 |
| Figure 4.2 | The effect of 1 $\mu$ M phenylephrine and 1 $\mu$ M acetylcholine on endothelium - denuded aortic ring preparations in representative traces of isometric force recording.  | 74 |
| Figure 4.3 | The concentration response curves of petroleum ether (GPPE), chloroform (GPCE), methanol (GPME) and water (GPWE) extracts of <i>G. procumbens</i> on phenylephrine (1 $\mu$ M) pre-contracted endothelium-intact (+E) rat aortic ring preparation. Results are expressed as means $\pm$ S.E.M (n = 8).  | 75 |
| Figure 4.4 | Representative traces of isometric force recording of 1 $\mu$ M phenylephrine-induced vasocontraction, 1 $\mu$ M acetylcholine-induced vasorelaxation and concentration dependent vasorelaxation of phenylephrine pre-<br>contracted endothelium-intact aortic ring preparations with <i>G. procumbens</i> petroleum ether extract (GPPE) 0.25, 0.5, 1, 2 and 3 mg/mL (Concentrations in organ bath). | 76 |
| Figure 4.5 | The effect of 1 $\mu$ M phenylephrine, 1 $\mu$ M acetylcholine<br>and <i>G. procumbens</i> chloroform extract (GPCE) 0.25,<br>0.5, 1, 2 and 3 mg/mL (concentrations in organ bath) on<br>phenylephrine pre-contracted endothelium-intact aortic<br>ring preparations in representative traces of isometric<br>force recording.  | 77 |
| Figure 4.6 | Representative traces of isometric force recording of 1 $\mu$ M phenylephrine-induced vasocontraction, 1 $\mu$ M acetylcholine-induced vasorelaxation and concentration dependent vasocontraction in phenylephrine pre-<br>contracted endothelium-intact aortic ring preparations with <i>G. procumbens</i> methanol extract (GPME) 0.25, 0.5, 1, 2 and 3 mg/mL (concentrations in organ bath)        | 78 |

| Figure 4.7  | Representative traces of isometric force recording of 1 $\mu$ M phenylephrine-induced vasocontraction, 1 $\mu$ M acetylcholine-induced vasorelaxation and concentration dependent vasocontraction in phenylephrine pre-<br>contracted endothelium-intact aortic ring preparations with <i>G. procumbens</i> water extract (GPWE) 0.25, 0.5, 1, 2 and 3 mg/mL (concentrations in organ bath).                                  | 79 |
|-------------|---|----|
| Figure 4.8  | The concentration response curves of petroleum ether<br>extract of G. procumbens (GPPE), its acetone soluble<br>and acetone insoluble fractions on phenylephrine (1<br>$\mu$ M) pre-contracted endothelium-intact (+E) rat aortic<br>ring preparation. Results are expressed as means ±<br>S.E.M (n = 8).   | 80 |
| Figure 4.9  | Representative traces of isometric force recording of 1 $\mu$ M phenylephrine-induced vasocontraction, 1 $\mu$ M acetylcholine-induced vasorelaxation and concentration dependent vasorelaxation in phenylephrine pre-<br>contracted endothelium-intact aortic ring preparations with <i>G. procumbens</i> petroleum ether extract (GPPE) 0.125, 0.25, 1 and 2 mg/mL (concentrations in organ bath).                          | 81 |
| Figure 4.10 | Representative traces of isometric force recording of 1 $\mu$ M phenylephrine-induced vasocontraction, 1 $\mu$ M acetylcholine-induced vasorelaxation and concentration dependent vasorelaxation in phenylephrine pre-<br>contracted endothelium-intact aortic ring preparations with acetone insoluble fraction of <i>G. procumbens</i> petroleum ether extract 0.125, 0.25, 1 and 2 mg/mL (concentrations in organ bath).   | 82 |
| Figure 4.11 | Representative traces of isometric force recording of 1 $\mu$ M phenylephrine-induced vasocontraction, 1 $\mu$ M acetylcholine-induced vasorelaxation and concentration dependent vasorelaxation in phenylephrine pre-<br>contracted endothelium-intact aortic ring preparations with acetone soluble fraction of <i>G. procumbens</i> petroleum ether extract 0.125, 0.25, 1 and 2 mg/mL (concentrations in organ bath).     | 83 |
| Figure 4.12 | The concentration dependent vasorelaxation of <i>G.</i><br><i>procumbens</i> petroleum ether extract (GPPE) on<br>endothelium-intact (+E) and endothelium denuded (-E)<br>rat aortic ring preparation pre-contracted with<br>phenylephrine (1 $\mu$ M). ** indicates <i>p</i> < 0.01. Results<br>are expressed as means ± S.E.M (n = 8). Statistics were<br>analysed by one-way ANOVA followed by Dunnett's<br>post-Hoc test. | 85 |

Figure 4.13 The concentration dependent vasorelaxation of G. procumbens petroleum ether extract (GPPE) on endothelium-intact (+E) rat aortic ring preparation precontracted with phenylephrine  $(1 \mu M)$  in the absence and in the presence of L-NAME (10  $\mu$ M). \* indicates p < 0.05. Results are expressed as means  $\pm$  S.E.M (n = 8). Statistics were analysed by one-way ANOVA followed Figure 4.14 The concentration dependent vasorelaxation of G. procumbens petroleum ether extract (GPPE) on endothelium-intact (+E) rat aortic ring preparation precontracted with phenylephrine  $(1 \mu M)$  in the absence and presence of atropine (10 µM). Results are expressed as means  $\pm$  S.E.M (n = 8). Statistics were analysed by one-way ANOVA followed by Dunnett's Figure 4.15 The concentration dependent vasorelaxation of G. procumbens petroleum ether extract (GPPE) on endothelium-intact (+E) rat aortic ring preparation precontracted with phenylephrine  $(1 \mu M)$  in the absence and presence of indomethacin (1  $\mu$ M). \*\* indicates p <0.01. Results are expressed as means  $\pm$  S.E.M (n = 8). Statistics were analysed by one-way ANOVA followed Figure 4.16 The concentration dependent vasorelaxation of G. procumbens petroleum ether extract (GPPE) on endothelium-denuded (-E) rat aortic ring preparation pre-contracted with phenylephrine (1 µM) in the absence and presence of propranolol (10 µM). Results are expressed as means  $\pm$  S.E.M (n = 8). Statistics were analysed by one-way ANOVA followed by Dunnett's Figure 4.17 The concentration dependent vasorelaxation of G. procumbens petroleum ether extract (GPPE) on endothelium-denuded (-E) rat aortic ring preparation pre-contracted with phenylephrine (1 µM) in the absence and presence of methylene blue (10 µM). Results are expressed as means  $\pm$  S.E.M (n = 8). Statistics were analysed by one-way ANOVA followed Figure 4.18 The concentration dependent vasorelaxation of G. procumbens petroleum ether extract (GPPE) on endothelium-denuded (-E) rat aortic ring preparation pre-contracted with phenylephrine  $(1 \mu M)$  in the absence and presence of barium chloride (10 µM). Results are expressed as means  $\pm$  S.E.M (n = 8).

|             | Statistics were analysed by one-way ANOVA followed by Dunnett's post-Hoc test   |
|-------------|---|
| Figure 4.19 | The concentration dependent vasorelaxation of <i>G.</i><br><i>procumbens</i> petroleum ether extract (GPPE) on<br>endothelium-denuded (-E) rat aortic ring preparation<br>pre-contracted with phenylephrine (1 $\mu$ M) in the<br>absence and presence of glibenclamide (10 $\mu$ M). *<br>indicates $p < 0.05$ . Results are expressed as means $\pm$<br>S.E.M (n = 8). Statistics were analysed by one-way<br>ANOVA followed by Dunnett's post-Hoc test |
| Figure 4.20 | The concentration dependent vasorelaxation of <i>G.</i><br><i>procumbens</i> petroleum ether extract (GPPE) on<br>endothelium-denuded (-E) rat aortic ring preparation<br>pre-contracted with phenylephrine (1 $\mu$ M) in the<br>absence and presence of 4-aminopyridine (1 $\mu$ M).<br>Results are expressed as means $\pm$ S.E.M (n = 8).<br>Statistics were analysed by one-way ANOVA followed<br>by Dunnett's post-Hoc test                         |
| Figure 4.21 | The concentration dependent vasorelaxation of <i>G.</i><br><i>procumbens</i> petroleum ether extract (GPPE) on<br>endothelium-denuded (-E) rat aortic ring preparation<br>pre-contracted with phenylephrine (1 $\mu$ M) in the<br>absence and presence of tetraethyl ammonium (1 $\mu$ M).<br>Results are expressed as means $\pm$ S.E.M (n = 8).<br>Statistics were analysed by one-way ANOVA followed<br>by Dunnett's post-Hoc test                     |
| Figure 4.22 | The effect of different concentrations of <i>G. procumbens</i> petroleum ether extract (GPPE) on calcium chloride-<br>induced vasocontraction in isolated denuded (-E) aortic<br>ring preparation in calcium free high potassium Krebs<br>solution. ** indicates $p < 0.01$ and *** indicates $p < 0.001$ . Results are expressed as the Mean $\pm$ SEM (n =<br>8). Statistics were analysed by one-way ANOVA<br>followed by Dunnett's post-Hoc test      |
| Figure 4.23 | The effect of different concentrations of verapamil on calcium chloride-induced vasocontraction in isolated denuded (-E) aortic ring preparation in calcium free high potassium Krebs solution. ** indicates $p < 0.01$ and *** indicates $p < 0.001$ . Results are expressed as the Mean $\pm$ SEM (n = 8). Statistics were analysed by one-way ANOVA followed by Dunnett's post-Hoc test  |

| Figure 4.24 | GC-MS chromatogram of GPPE. The peaks are labeled<br>to indicate the retention time (RT). Peak RT 27.09<br>indicates the presence of ursolic acid   |
|-------------|---|
| Figure 4.25 | GC-MS chromatogram of GPPE. The peaks are labeled<br>to indicate the retention time (RT). Peak RT 8.86<br>indicates the presence of spathulenol   |
| Figure 4.26 | Calibration curve of standard (ursolic acid) 110  |
| Figure 4.27 | Spectrum of (A) GPPE (5000 $\mu$ g/mL) and (B) standard ursolic acid (500 $\mu$ g/mL) in Ethyl Acetate at 223nm 111   |
| Figure 4.28 | The effect of daily oral administration of <i>G.</i><br><i>procumbens</i> petroleum ether (GPPE), chloroform<br>(GPCE), methanol (GPME) and water (GPWE) extracts<br>on systolic blood pressure (SBP) of Spontaneously<br>Hypertensive Rats (SHR). The values are the mean $\pm$<br>SEM (n = 6)   |
| Figure 4.29 | The effect of daily oral administration of <i>G</i> .<br><i>procumbens</i> water extract on systolic blood pressure<br>(SBP) of spontaneously hypertensive rats. ** indicates<br>p < 0.01 and *** indicates $p < 0.001$ . The values are the<br>mean $\pm$ SEM (n = 6). Statistics were analysed by one-<br>way ANOVA followed by Dunnett's post-Hoc test   |
| Figure 4.30 | The reduction in mean arterial pressure (MAP) elicited<br>by intra-venous (i.v.) administration of <i>G. procumbens</i><br>petroleum ether (GPPE), chloroform (GPCE), methanol<br>(GPME) and water (GPWE) extracts in anesthetized<br>normotensive rats. The results are presented as mean $\pm$<br>SEM whereas n = 6   |
| Figure 4.31 | The effect of intravenous (i.v.) administration of <i>G</i> .<br><i>procumbens</i> water extract (GPWE), in the absence and<br>presence of atropine 1 mg/kg on the mean arterial<br>pressure (MAP) of anesthetized normotensive rats. The<br>results are presented as mean $\pm$ SEM whereas (n = 6).<br>Statistics were analysed by one-way ANOVA followed<br>by Dunnett's post-Hoc test                     |
| Figure 4.32 | The effect of intravenous (i.v.) administration of <i>G.</i><br><i>procumbens</i> water extract (GPWE), in the absence and<br>presence of propranolol 2 mg/kg on the mean arterial<br>pressure (MAP) of anesthetized normotensive rats. **<br>indicates $p < 0.01$ . The results are presented as mean $\pm$<br>SEM (n = 6). Statistics were analysed by one-way<br>ANOVA followed by Dunnett's post-Hoc test |

## LIST OF ABBREVIATIONS

| °C               | Degree celsius                                      |
|------------------|---|
| %                | Percent   |
| ±                | Plus, minus   |
| AA               | Arachidonic acid                                    |
| ACE              | Angiotensin converting enzyme                       |
| ACEI             | Angiotensin converting enzyme inhibitor             |
| ACh              | Acetylcholine                                       |
| Ang              | Angiotensin   |
| ANOVA            | Analysis of variance                                |
| AT1              | Angiotensin type I receptor                         |
| ATP              | Adenosine triphosphate                              |
| AV               | Atrioventricular                                    |
| BK <sub>ca</sub> | Big-conductance calcium-sensitive potassium channel |
| BP               | Blood pressure                                      |
| Ca <sup>2+</sup> | Calcium   |
| cAMP             | Cyclic adenosine monophosphate                      |
| cGMP             | Cyclic guanosine monophosphate                      |
| CHF              | Congestive heart failure                            |
| Cl-              | Chloride  |
| CNS              | Central nervous system                              |
| COX              | Cyclooxygenase                                      |
| CVD              | Cardiovascular disease                              |
| DAG              | Diacylglycerol                                      |
| DBP              | Diastolic blood pressure                            |
| DMSO             | Dimethylsulfoxide                                   |

| EDHF             | Endothelium-derived hyperpolarizing factor                   |
|------------------|--|
| EDRF             | Endothelium-derived relaxation factor                        |
| EP               | Epinephrine  |
| eNOS             | Endothelial nitric oxide synthase                            |
| GC-MS            | Gas chromatography mass-spectrometry                         |
| G. procumbens    | Gynura procumbens  |
| GPPE             | Gynura procumbens petroleum ether extract                    |
| GPCE             | Gynura procumbens chloroform extract                         |
| GPME             | Gynura procumbens methanol extract                           |
| GPWE             | Gynura procumbens water extract                              |
| g                | Gram   |
| HPLC             | High performance liquid chromatography                       |
| i.p              | Intraperitoneal  |
| i.v              | Intravenous  |
| IK <sub>ca</sub> | Intermediate-conductance calcium-sensitive potassium channel |
| IP               | PGI <sub>2</sub> receptor                                    |
| IP <sub>3</sub>  | 1,4,5-inositol triphosphate                                  |
| $\mathbf{K}^+$   | Potassium  |
| KATP             | ATP-sensitive potassium channel                              |
| K <sub>ca</sub>  | Calcium-activated potassium channel                          |
| K <sub>IR</sub>  | Inward rectifier potassium channel                           |
| Kv               | Voltage-gated K <sup>+</sup> channel                         |
| L-NAME           | $N\omega$ -Nitro-L-arginine methyl ester                     |
| μg               | Microgram  |
| μL               | Microliter   |
| М                | Molar  |

| MAP              | Mean arterial pressure                   |
|------------------|--|
| mg               | Milligram                                |
| MI               | Myocardial infarction                    |
| mL               | Milliliter                               |
| MLC              | Myosin light chain                       |
| mM               | Millimolar                               |
| K <sub>IR</sub>  | Inward rectifier potassium channel       |
| Kv               | Voltage-gated K <sup>+</sup> channel     |
| L-NAME           | $N\omega$ -Nitro-L-arginine methyl ester |
| μg               | Microgram                                |
| μL               | Microliter                               |
| М                | Molar                                    |
| MAP              | Mean arterial pressure                   |
| mg               | Milligram                                |
| MI               | Myocardial infarction                    |
| mL               | Milliliter                               |
| MLC              | Myosin light chain                       |
| mM               | Millimolar                               |
| GC-MS            | Gas chromatography-Mass spectrometry     |
| Na <sup>+</sup>  | Sodium                                   |
| NE               | Norepinephrine                           |
| NO               | Nitric oxide                             |
| NOS              | Nitric oxide synthase                    |
| PEG              | Polyethylene glycol                      |
| PGI <sub>2</sub> | Prostacyclin                             |
| PIP <sub>2</sub> | Phosphatidyl inositol-(4,5)-bisphosphate |
| РКС              | Protein kinase A                         |

| РКС   | Protein kinase C               |
|-------|--------------------------------|
| PLC   | Phospholipase C                |
| PVR   | Peripheral vascular resistance |
| ROS   | Reactive oxygen species        |
| S.E.M | Standard error of mean         |
| SBP   | Systolic blood pressure        |
| SD    | Sprague Dawley                 |
| SNP   | Sodium nitroprusside           |
| SNS   | Sympathetic nervous system     |
| SVR   | Systemic vascular resistance   |
| TEA   | Tetraethyl ammonium            |
| TPR   | Total peripheral resistance    |
| VSM   | Vascular smooth muscle         |

# PENILAIAN KESAN ANTI-HIPERTENSI DAN KESAN VASORELAKSAN EKSTRAK GYNURA PROCUMBENS MERR

#### ABSTRAK

Penyakit kardiovaskular terus meningkat di kedua-dua negara maju dan membangun. Tekanan darah tinggi merupakan "pembunuh senyap" yang sedia diketahui menjadi punca utama kepada komplikasi kardiovaskular. Kawalan tekanan darah yang tidak mencukupi boleh membina komplikasi yang berbeza seperti strok, infarksi miokardium, hipertrofi ventrikel kiri, kegagalan buah pinggang dan jantung. Gynura procumbens (G. procumbens) secara tradisinya digunakan untuk rawatan hipertensi. Ia telah dihipotesiskan bahawa, G. procumbens mungkin mempunyai ciriciri anti-hipertensi, hipotensi dan vasorelaksan yang dihasilkan oleh beberapa molekul aktif, yang akan menurunkan tekanan darah dan vasodilasi. Daun-daun G. procumbens kering dikizar menjadi serbuk halus. Serbuk daun ini telah diekstrak secara bersiri menggunakan eter petroleum, kloroform, metanol dan air melalui proses maserasi. Setiap ekstrak telah dikeringkan di bawah tekanan yang dikurangkan. Empat jeniz ekstrak, iaitu eter petroleum (GPPE), kloroform (GPCE), metanol (GPME) dan air (GPWE) daripada G. procumbens diperolehi. Kesan setiap ekstrak diperiksa pada cincin aortik tikus yang terpencil dan persediaan tikus yang dibius. Objektif kajian ini adalah untuk mengkaji aktiviti anti-hipertensi secara dalam tikus berhipertensi sponttan, aktiviti vasorelaksan dalam persediaan cincin aortik tikus, aktiviti penuninan tekanan darah dan mekanisme tindakan tikus normotif anestetik bagi ekstrak G. procumbens. Pemberian harian oral GPWE mempunyai lebih banyak kesan anti-hipertensi dalam SHR berbanding ekstrak lain. Pemberian intravena GPWE, berbanding ekstrak lain, juga mempunyai lebih banyak kesan

pengurangan tekanan darah dalam tikus normotif anestetik. GPPE dan pecahannya menunjukkan lebih banyak vasorelaksasi dalam persediaan cincin aorta. Penguncupan yang disebabkan oleh kalsium adalah ketara (p < 0.01) dan (p < 0.001) dihalang oleh GPPE dalam lengkungan tindak balas kalsium bergantung kepekatan cincin aorta. GPPE mempunyai kesan yang sama seperti verapamil pada saluran kalsium bergantung voltan cincin aorta. Saluran kalium sensitif dengan ketara (p < p0.05) menghalang vasorelaksasi GPPE. Pra-rawatan dengan propranolol menghalang ketara (p < 0.01) kesan-kesan hipotensi GPWE dalam tikus normotif anestetik. Kesan penurunan GPWE pada tikus anestetik mungkin disebabkan oleh kesan β-adrenergik. GPWE boleh bertindak sebagai  $\beta_2$ -agonis. Ekstrak yang lebih polar (GPWE) di dapati telah mengekalkan beberapa aktiviti anti-hipertensi dan penurunan tekanan darah, manakala ekstrak bukan kutub (GPPE) mempunyai lebih banyak aktiviti vasorelaksan. Keputusan ini menunjukkan bahawa G. procumbens menginduksi kesan kardiovaskularnya pada reseptor  $\beta_2$ -adrenergik dalam otot licin vaskular secara merangsang laluan adenosine monophosphate kitaran, saluran potassium sensitif ATP dan menghalang saluran kalsium jenis-L. Analisis Kromatografi gasspektrometri massa dan kromatografi cecair berprestasi tinggi menunjukkan kehadiran spatulenol dan asid ursolik, yang menunjukkan bahawa kesan kardiovaskular mungkin berkaitan dengan kehadiran sebatian-sebatian ini.

# EVALUATION OF ANTI-HYPERTENSIVE AND VASORELAXANT EFFECTS OF *GYNURA PROCUMBENS* MERR. EXTRACTS

#### ABSTRACT

The cardiovascular diseases are continually increasing in both developed and developing countries. High blood pressure is a well-known "silent killer" which is the leading cause of cardiovascular complications. The inadequate control of blood pressure can develop different complications such as stroke, myocardial infarction, left ventricular hypertrophy, renal and heart failure. Gynura procumbens (G. procumbens) traditionally used for treatment of hypertension. It was hypothesised that, G. procumbens may have anti-hypertensive, hypotensive and vasorelaxant properties due to some active molecules that would lower blood pressure and vasodilations. The dried G. procumbens leaves were ground into a fine powder. The powdered leaves material was serially extracted with petroleum ether, chloroform, methanol and water by maceration process. Each extract was dried under reduced pressure. Four extracts, petroleum ether (GPPE), chloroform (GPCE), methanol (GPME) and water (GPWE) of G. procumbens were obtained. The effect of each extract was examined on isolated rat aortic ring and anesthetized rat preparations. The aims of the present study were to investigate the anti-hypertensive activity in spontaneously hypertensive rats (SHR), vasorelaxant activity in rat aortic ring preparations and blood pressure lowering activity and mechanism of action in anesthetized normotensive rats of G. procumbens extracts. Daily oral administration of GPWE has more anti-hypertensive effects in SHR compared to other extracts. The intravenous (i.v.) administration of GPWE among the extracts also has more blood pressure lowering effects in anesthetized normotensive rats. GPPE and its fractions

show more vasorelaxation in aortic ring preparations. Calcium-induced contraction was significantly (p < 0.01) and (p < 0.001) inhibited with GPPE in concentration dependent calcium response curve of aortic rings. GPPE has similar effects as verapamil on voltage dependent calcium channel (VDCC) of aortic rings. ATP sensitive potassium channel ( $K_{ATP}$ ) significantly (p < 0.05) inhibit the vasorelaxation of GPPE. Pre-treatment with propranolol inhibit significantly (p < 0.01) the hypotensive effects of GPWE in anesthetized normotensive rats. The blood pressure lowering effects of GPWE in anesthetized rat may be due to  $\beta$ -adrenergic effects. GPWE may act as  $\beta_2$ -agonist. The more polar extract (GPWE) appeared to retain some anti-hypertensive and blood pressure lowering activities, while non-polar extract (GPPE) has more vasorelaxant activity. The results suggest that G. procumbens induced its cardiovascular effects on  $\beta_2$ -adrenergic receptors in the vascular smooth muscle by stimulating cyclic adenosine monophosphate (cAMP) pathway, sensitive potassium channel (K<sub>ATP</sub>) and inhibiting L-type calcium channels. Gas chromatography mass-spectrometry (GC-MS) and high-performance liquid chromatography (HPLC) analysis shows the presence of spathulenol and ursolic acid, which suggest that the cardiovascular effect could be related to the presence of these compounds.

#### **CHAPTER 1**

#### **INTRODUCTION**

#### 1.1 Background

In 2013, cardiovascular illness was the most common underlying cause of death in the world. According to an assessment, 17.3 million of total deaths in the world, was due to cardiovascular illness (Benjamin, 2017). The cardiovascular diseases are continually increasing in both developed and developing countries (Balakumar et al., 2016). Globally in 2008, the incidence of hypertension among adults aged 25 and above was almost 40 %. In 2009, the total financial load of hypertension in the United States was almost USD 73.4 billion. Even in the presence of clinical practice guidelines (CPGs), optimal hypertension control is not achieved. According to a study, a decrease of 10 mm Hg in systolic blood pressure and 5 mm Hg in diastolic blood pressure, causes a 20 percent decrease of coronary heart disease and a 32 percent decrease of stroke in one year (Al-Ansary et al., 2013). According to the Centres for Disease Control and Prevention (CDC), even a very slight rise in BP increases the risk for cardiovascular disease. Appropriate treatment of hypertension can decrease the chances of heart attacks and strokes.

#### **1.2 Problem statement**

In spite of there are many antihypertensive drugs available, but there are many research publications on their quite little effectiveness in the form of monotherapy as well as long-lasting side effects (Jarari et al., 2016, Guerrero-García and Rubio-Guerra, 2018). Therefore, the finding of novel antihypertensive drugs is still a hot issue. Agents that can act on vascular tone in addition to lower blood pressure may lead to a good approach in the treatment of hypertension and prevention of cardiovascular morbidities (Yannoutsos et al., 2016). Therefore, the discovery of new molecules is highly desired.

The importance of *G. procumbens* in the cardiovascular studies has been reported (Abrika et al., 2013, Kaur et al., 2012, Kaur et al., 2013, Hoe et al., 2011). Previous studies mostly have investigated the cardiovascular activity of the polar portion of *G. procumbens* extracts, still non-polar part of the plant for its cardiovascular effects have not been validated in the laboratory. This background offers an opportunity to study the plant both pharmacologically and phytochemically.

#### 1.3 Hypothesis

It was hypothesised that, *G. procumbens* may have anti-hypertensive, hypotensive and vasorelaxant properties due to some active molecules that would lower blood pressure and vasodilations. The tail cuff experimental model was proposed to explore the anti-hypertensive effect of different extracts of leaves of *G. procumbens* in spontaneously hypertensive rats (SHR). The anesthetized rat experimental model was suggested for the possible blood pressure lowering mechanism of action for the most active *G. procumbens* extract. The isolated rat aortic ring model was proposed to find the vasorelaxant activity and the possible mechanism of action of different extracts of *G. procumbens*.

#### 1.4 Objectives

1. To investigate the vasorelaxant activity of different extracts obtained from the leaves of *G. procumbens* and to explore the possible mechanism of action of the most active *G. procumbens* extract using isolated rat aortic ring model.

- 2. To evaluate the possible blood pressure lowering mechanism of action for the most active *G. procumbens* water extract using anesthetized rat experimental model in normotensive rats.
- 3. To determine the anti-hypertensive effect of different extracts of leaves of *G. procumbens* in spontaneously hypertensive rats (SHR) using tail cuff experimental model.
- 4. To characterize, the main possible components responsible for the cardiovascular effects of *G. procumbens* by using Gas chromatographymass spectrometry (GC-MS) and high-performance liquid chromatography (HPLC).

#### **CHAPTER 2**

#### LITERATURE REVIEW

#### 2.1 Hypertension

Hypertension can be well-defined as if a person experiences systolic blood pressure (SBP)  $\geq$ 140 mmHg or diastolic blood pressure (DBP)  $\geq$ 90 mmHg or taking the medication for hypertension (CDC, 2012). The Seventh Report of the Joint National Committee (JNC) USA, classified hypertension as below:

- 1. Normal blood pressure (SBP <120 mmHg, DBP < 80mmHg)
- 2. Prehypertension (SBP 120 to 139 mmHg, DBP 80 to 89 mmHg)
- 3. Stage 1 hypertension (SBP 140 to 159 mmHg, DBP 90 to 99 mmHg)
- 4. Stage 2 hypertension (SBP 160 mmHg, DBP 100 mmHg) (Holm et al., 2006).

High blood pressure is a well-known "silent killer" which is the leading cause of cardiovascular complications. Approximately, 13 percent of world population experiences high blood pressure, and mostly belong to developing countries. In 2001, higher than 32 percent population in Malaysia, aged 18 years and above, and 43 percent, aged 30 years and above, were hypertensive (Loh et al., 2018). High blood pressure is one of the deadly causes of mortality in the world, due to its symptomless behavior and complications which can cause accompanying illnesses, for example, stroke (Loh et al., 2016).

The inadequate control of BP can develop different complications such as stroke, myocardial infarction, left ventricular hypertrophy, renal and heart failure.

The blood pressure depends upon:

- 1. The force of received blood, by which, the heart muscles are stretched.
- 2. The resistance to blood flow in the blood vessels.
- 3. The volume of blood in the blood vessels.
- 4. The autonomic nervous system.
- 5. The kidney increases the blood pressure by:
  - Inducing the contraction of the blood vessels.
  - Increasing the volume of blood in the blood vessels (Lionakis et al., 2012).

### 2.2 The Cardiovascular system

#### 2.2.1 The Heart

The heart works as a pump, which supplies blood throughout the body by constant and rhythmical contractions.

The heart wall comprises three layers:

- 1. Pericardium (the outermost layer)
- 2. Myocardium (the middle layer)
- 3. Endocardium (the innermost layer).

Between the two ventricles an interventricular septum, and between the atrium and ventricle, the atrioventricular (AV) valves are present, which permit only one-way blood flow. The right AV valve is tricuspid, and the left AV valve is bicuspid (Weinhaus and Roberts, 2009).



Figure 2.1 The cardiovascular system.

The deoxygenated blood accumulates in the right atrium via the superior venae cavae and inferior venae cavae, and then pushed into the right ventricle. The right ventricle pumps the deoxygenated blood to the lungs, via the pulmonary artery. The oxygenated blood comes to the left atrium, through pulmonary veins. The left atrium pushes the blood into the left ventricle. The left ventricle supplies blood, to the body. The exchange, of oxygen and carbon dioxide, takes place via capillaries. The cardiac muscle is an involuntary striated muscle, like skeletal and smooth muscles. The cardiac muscles are highly resistant, to the tiredness, because of a large number of mitochondria, myoglobin and adequate blood supply (Weinhaus and Roberts, 2009).

#### 2.2.2 Arteries and arterioles

Arterial architecture is increasingly recognized as an essential determinant in the pathophysiology of hypertension and cardiovascular diseases (Mayet and Hughes, 2003). Agents that can act to modulate arterial wall structure and function, in addition to lower blood pressure may lead to a novel approach in the treatment of hypertension and prevention of cardiovascular morbidities (Yannoutsos et al., 2016).

There are three different layers, in the wall of an artery:

- 1. Tunica intima
- 2. Tunica media
- 3. Tunica adventitia

Tunica intima is inner layer, which comprises a single layer of endothelial cells (Figure 2.2). Tunica media is the middle layer and tunica adventitia is the outer layer. There is an internal elastic lamina of the connective tissue between the tunica media and the tunica intima (Loh et al., 2018). Tunica adventitia comprises fibroblasts, collagen, and sympathetic nerves. The arterial resistance depends upon the diameter of the lumen. In diastole phase, the occurrence of elastic tissues in the vessels, decrease resistance (Widmaier et al., 2006). In the walls of the arteriole, there is less elastic tissue so causes more resistance to the blood flow. The slight changes, in the diameter of the arteriole, can cause a significant change in the total peripheral resistance (TPR) (Mayet and Hughes, 2003).



Figure 2.2 Structure and layers of a typical blood vessel in the circulatory system. (Diagram adapted from McGraw-Hill companies.inc).

#### 2.3 Vascular Smooth Muscle Contraction and Relaxation

The vascular smooth muscle cell (VSM) forms an integral structural element of the blood vessels and involve in the regulatory processes of the vascular system (Saddouk et al., 2017). It is necessary to understand the signaling mechanism pathways before the start of vasculature-associated research. The smooth muscle contraction takes place, due to increased intracellular calcium. There are numerous signal transduction mechanisms such as G-protein-coupled pathway, nitric oxidecGMP pathway, voltage-dependent Ca<sup>2+</sup> channels (VDCC), and receptor-operated Ca<sup>2+</sup> channels (ROCC) which control intracellular calcium concentration and consequently the state of vascular tone. These signal transduction mechanisms take place in the vascular endothelium and the VSM. The relaxation and contraction of the blood vessels occurred by these signal transduction mechanisms and finally maintained by the actin and the myosin filament. The vascular contraction occurs by the sliding of the actin and myosin filaments over one another. The contraction in the VSM takes place by the opening of voltage-dependent calcium channels (L-type calcium channels); the electrical depolarization takes place, which increases intracellular calcium concentration (Figure 2.3). Many chemicals act as a stimulant, for instance, epinephrine, norepinephrine, vasopressin, endothelin-1, thromboxane A2, and angiotensin II can induce contraction. The receptors are present on the endothelium and vascular smooth muscle, by which these chemicals bind and induce contraction in the blood vessel (Webb, 2003).



Figure 2.3 Mechanisms, by which contraction of vascular smooth muscle takes place. (Webb, 2003).

The free intracellular calcium binds with the specific protein calmodulin. This calcium-calmodulin complex activates an enzyme myosin light chain kinase (MLCK). The phosphorylation of myosin light chain (MLC) takes place in the presence of ATP. The removal of calcium takes place by the ATP-dependent calcium pump and the sodium-calcium exchanger. The chemical mechanism which modifying the  $Ca^{2+}$  metabolism is very important, in the stability of vascular smooth muscle tone (Somlyo et al., 1999).

A myosin light chain phosphatase (MLCP) enzyme is present in myosin light chain, which catalysis  $Ca^{2+}$ , consequently intracellular  $Ca^{2+}$  level comes down to the resting stage, and dephosphorylation takes place, and relaxation of vascular smooth muscle occurs. The small G protein RhoA and its downstream target Rho kinase play an important role in the regulation of MLC phosphatase activity. Rho kinase phosphorylates the myosin binding subunit of MLC phosphatase, inhibiting its activity and thus promoting the phosphorylated state of MLC (Somlyo et al., 1999, Webb, 2003). For the elimination of cytosolic  $Ca^{2+}$ , there are several mechanisms. The relaxation of smooth muscle cells occurs, because of the inhibition of  $Ca^{2+}$  /  $Mg^{2+}$  - ATPase action, in the sarcoplasmic reticulum, which diminishes cytosolic  $Ca^{2+}$ . Furthermore, the blockage of receptor-operated and voltage-operated  $Ca^{2+}$ channels, causes the relaxation of smooth muscle, because of a reduction in intracellular  $Ca^{2+}$  (Webb, 2003).

#### 2.3.1 Endothelium

The endothelium is present in the whole vascular system. The endothelium regulates the vascular function by neurotransmitters, hormones, and vasoactive factors (Sandoo et al., 2010). For the protection of the blood vessels, an optimum

level of these factors is required, while imbalance of these factors causes endothelial dysfunction. and atherosclerosis (Lerman and Zeiher, 2005, Sandoo et al., 2010). The endothelium releases many vasoactive factors, for example, nitric oxide (NO) and prostacyclin (PGI<sub>2</sub>), which induce vasorelaxation and endothelin-1 (ET-1) and thromboxane (TXA<sub>2</sub>), induce contraction (Figure 2.4 and Figure 2.5) (Loh et al., 2018).

Figure 2.4 describes the production, of the endothelial nitric oxide and its effects in the cells of vascular smooth muscle. Acetylcholine (ACh), adenosine diphosphate (ADP), bradykinin (BK) and adenosine triphosphate (ATP) are examples of NO synthesis, by the depletion of intracellular Ca<sup>2+</sup>stores (Lambert et al., 1986, Schilling and Elliott, 1992, Schilling et al., 1992, Moncada and Higgs, 2006). When intracellular levels of Ca<sup>2+</sup> increase, eNOS detaches from caveolin and is activated. Ca<sup>2+</sup> attaches to the protein calmodulin in the cytoplasm of the cell, after which it undergoes structural changes which allows it to bind to eNOS (Fleming and Busse, 1999). Consequently, the eNOS changes L-arginine to NO (Palmer et al., 1988). When the Ca<sup>2+</sup> level decreases, then break down of calcium-calmodulin complex takes place and deactivated by binding with caveolin (Sandoo et al., 2010).



Figure 2.4 The production, of the endothelial nitric oxide and its effects in the cells of vascular smooth muscle (Sandoo et al., 2010).

Abbreviations: ACh= acetylcholine; BK= bradykinin; ATP= adenosine triphosphate; ADP= adenosine diphosphate; SP= substance P; SOCa<sup>2+</sup>= store-operated Ca<sup>2+</sup> channel; ER= endoplasmic reticulum; NO= nitric oxide; sGC= soluble guanylyl cyclase; cGMP= cyclic guanosine-3', 5-monophosphate; MLCK= myosin light chain kinase. When Ca<sup>2+</sup> stores of the endoplasmic reticulum are depleted a signal is sent to SOCa<sup>2+</sup> channel which allows extracellular Ca<sup>2+</sup> into the endothelial cell.



Figure 2.5 Flow chart diagram describe the vasodilation by the production of PGI<sub>2</sub> and NO in the endothelial cell.

A high blood pressure, in the blood vessel, induces shear stress. This shear stress causes the production of NO, by the process of phosphorylation (Figure 2.4). The extent of the shear stress is, directly proportional to the production of the NO. Whenever, the shear stress is for the short period, then the intracellular  $Ca^{2+}$  is released, while if shear stress is for a longer period (>30 min), then the production of the NO takes place due to the phosphorylation of the eNOS (Pittner et al., 2005, Sandoo et al., 2010, Mount et al., 2007).

From the endothelial cell, the NO penetrates in to the smooth muscle. Then the NO attaches with the enzyme soluble guanylyl cyclase (sGC) and stimulates the enzyme. The conversion of guanosine triphosphate (GTP) to cGMP increases by the stimulated enzyme. Consequently, the contraction of the smooth muscle decreases. From the sarcoplasmic reticulum, the secretion of the  $Ca^{2+}$  decreases and  $Ca^{2+}$ restores in the sarcoplasmic reticulum. Thus, the relaxation of smooth muscle cells occurs (Davignon and Ganz, 2004, Sandoo et al., 2010). With a continuous production of the NO, the vasodilator tone is maintained (Gladwin et al., 2004). The N $\infty$ -nitro-1-arginine methyl ester (L-NAME) is a nitric oxide synthase (NOS) inhibitor (Dawes et al., 2001). Disturbance of vascular homeostasis can lead to the development of endothelial dysfunction. The steady production of the endothelin (vasoconstrictors) and the NO (vasodilator) maintains vascular tone (Sandoo et al., 2010). One of the leading causes of endothelial dysfunction is the decreased level of NO in the blood. There is an association, between increased age, the endothelium dysfunction, and cholesterol (Gimbrone and García-Cardeña, 2016).

There are many elements, which regulate the tone of the blood vessels, which are given below in **Error! Reference source not found.**.

Table 2.1 Endothelium-derived vasoactive factors. Summary of major vasoactive factors (dilators and constrictors) found in endothelium, their synthesizing enzyme, target, the effect on tone and mechanism of action.

| zes<br>iscle |
|--------------|
|              |
|              |
|              |
|              |
|              |
|              |
|              |
|              |
|              |
|              |

Abbreviations: NO - Nitric oxide, EDHF - Endothelium-derived hyperpolarizing factor, PGH2 - Prostaglandin H2 (endoperoxide), ET - Endothelin, Ang II - Angiotension II, PGI - Prostacyclin, TXA2 - Thromboxane A2,  $\uparrow$  or  $\downarrow$  - increase or decrease.

The interaction among endothelium and VSM is closely related. Both act synergistically in a complex manner to maintain the normal arterial tone.



Vascular endothelium

Vascular smooth muscle cells

Figure 2.6 The signalling pathways in vascular endothelium and vascular smooth muscle acts synergistically to maintain the vasocontractions and vasorelaxations in the blood vessels.

#### 2.4 Enzyme-Linked Receptors

Enzyme-Linked Receptors are situated on the membrane and known as catalytic receptors, which are stimulated by catalytic enzymes and ligand-receptors. The guanylyl cyclase is essential enzyme-linked receptors that play a key role in the maintenance of the vascular tone. The NO diffuses into the VSM after its synthesis in the endothelium and attaches with soluble guanylyl cyclase (sGC). Activation of sGC by NO in VSM leads to the conversion of guanosine 5' – triphosphate (GTP) to cyclic guanosine 3', 5' -monophosphate (cGMP) as shown in Figure 2.7. The cGMP activates the protein kinase G (PKG) which reduces the intracellular Ca<sup>2+</sup> release

from sarcoplasmic reticulum store and causes vascular smooth muscle relaxation (Loh et al., 2018, Fellner and Arendshorst, 2002).



Figure 2.7 The role of Enzyme-Linked Receptors in vasorelaxation.

#### 2.4.1 Soluble Guanylyl Cyclase pathway

In the cytosol of the VSMCs, the sGC is freely available. The NO attaches, with the heme group of the sGC. On the activation of the sGC, the conversion of the GTP into the cGMP takes place, which stimulates the PKG. As a result, vasodilation occurs (Ko et al., 2008, Loh et al., 2018). The 1H- [1,2,4] oxadiazole [4,3-a] quinoxaline-1-one (ODQ) is dissolved in dimethylsulfoxide (DMSO) and block the sGC, by oxidizing the heme group of the sGC (Loh et al., 2018). The methylene blue (MB) is another cGMP blocker (Kontos and Wei, 1993, Evora, 2016).

#### 2.4.2 Serine-Threonine Protein Kinases

These are kinase enzymes, which are triggered when attached to their second messenger, and phosphorylation of their hydroxyl (OH) group takes place. The protein kinase A (PKA), protein kinase C (PKC), and protein kinase G (PKG) is involved in the functioning of the blood vessels. The PKA is a cAMP-dependent and is activated when attaches with cAMP. The vasodilation occurs, when its

phosphorylation takes place. By phosphodiesterase 3 (PDE<sub>3</sub>), the breakdown of cAMP takes place, into adenosine monophosphate (Loh et al., 2018, Bouschet et al., 2003). In the endothelium of the blood vessels and VSMCs, the PKC is triggered by DAG and attaches with  $Ca^{2+}$  ions at  $C_1$  and  $C_2$  domain respectively. Phosphorylation of the serine or threonine sites takes place when the PKC is activated. Then, contraction of the blood vessels takes place (Huang, 1989). The PKG is activated, on the attachment of the cGMP. Then dilation of the blood vessels takes place (Wall et al., 2003, Paul and Snyder, 2012).

#### 2.5 G-Protein-Coupled Receptors (GPCRs)

G-Protein-Coupled Receptors are present on intracellular surface of cell membranes and known as the seven-transmembrane domain receptors, which are activated when attached to their ligand and transmit signals into the cell. Three types of guanine nucleotide binding protein (G-protein) are present on both endothelium and VSMC. G $\alpha$ -, G $\beta$ -, and G $\gamma$ -proteins are subunits of G-protein. G $\alpha$  performs the main role in the blood vessels, which is further sub-divided into three types—Gq $\alpha$ , Gi $\alpha$ , and Gs $\alpha$ . The activation of the G-protein is started, when GPCRs attach to a ligand and makes the G-protein perform as guanine nucleotide exchange factor (GEF) and replace its guanosine diphosphate (GDP) into GTP. When attached with GTP, the G-protein trimer detaches into G $\alpha$ -GTP monomer and G $\beta\gamma$ -dimer. G $\alpha$ -GTP monomer initiates to act with intracellular proteins for signal transduction, and then G $\beta\gamma$ -dimer tends to activate numerous types of signaling molecules, together with ion channels, phospholipases, lipid kinases, and its particular signaling cascades (Walaas et al., 1992, Dorsam and Gutkind, 2007, Yuen et al., 2010).



Figure 2.8 G protein-coupled receptor signalling (Lynch and Wang, 2016).

#### 2.5.1 Gqa-Protein-Coupled Receptors

The stimulated Gq $\alpha$ -protein splits the phosphatidylinositol 4,5-bisphosphate (PIP<sub>2</sub>) into second messengers, inositol triphosphate (IP<sub>3</sub>) and DAG by binding its Gq $\alpha$ -subunit to the phospholipase C (PLC). The IP<sub>3</sub> binds to the intracellular receptor, IP<sub>3</sub> receptor (IP<sub>3</sub>R), on the SR to activate the intracellular release of the Ca<sup>2+</sup> ions from the SR into the cytosol. While the DAG activates the PKC. As a result, the Ca<sup>2+</sup> concentration in the cytosol increases. In the blood vasculature, the Gq $\alpha$ -protein-coupled receptors are present in endothelium, which contains angiotensin-2 receptor (AT<sub>2</sub>), serotonin receptor (5-HT<sub>1D</sub>), bradykinin receptor (B<sub>2</sub>), muscarinic-3 receptor (M<sub>3</sub>), endothelin-B receptor (ET<sub>B</sub>R), and calcitonin receptor-like receptor, angiotensin-1 receptor (AT<sub>1</sub>), endothelin receptors (ET<sub>A</sub>R and ET<sub>B</sub>R),

serotonin receptor (5-HT<sub>2</sub>), and TXA<sub>2</sub> receptor in VSMCs (Jakala et al., 2009, Loh et al., 2018, Bockaert et al., 2006, Ishii and Kurachi, 2006).

#### 2.5.2 Gia-Protein-Coupled Receptors

The Gia-protein-coupled receptors are present in the VSMCs, for instance, a  $\alpha$ 2-adrenergic receptor ( $\alpha$ 2). When the receptor is stimulated, the activity to convert ATP into cAMP is inhibited, as a result, vasoconstriction occurs (Qin et al., 2008).

#### 2.5.3 Gsa-Protein-Coupled Receptors

The Gs $\alpha$ -protein-coupled receptors are opposite, to the Gi $\alpha$ -protein-coupled receptors. When the receptor is stimulated, it activates AC to produce cAMP from ATP, the cAMP will increase, the activation of PKA. As a result, vasodilation occurs. There are two important Gs $\alpha$ -protein-coupled receptors in VSMCs, for instance, the  $\beta$ 2-adrenergic receptor ( $\beta$ <sub>2</sub>) and PGI<sub>2</sub> receptor (IP) (Jakala et al., 2009).

#### 2.6 Channel-Linked Receptors

The Channel-Linked Receptors are also known as ion channel-linked receptors or ionotropic receptors or ligand-gated receptors. These are triggered when attached to their ligand and allowing ions, for example, Na<sup>+</sup>, K<sup>+</sup>, Cl<sup>-</sup>, and Ca<sup>2+</sup> to pass through the membrane. The action potential in VSMCs is controlled, by these receptors through depolarization or hyperpolarization. There are two important channel-linked receptors, playing critical roles in vascular tone regulation, which are K<sup>+</sup> and Ca<sup>2+</sup> channels (Loh et al., 2018).

#### 2.6.1 Potassium Channels

The Potassium channel is widely distributed, in living organisms. There are four types of K<sup>+</sup> channels in the blood vessels:

- 1. Calcium-activated K<sup>+</sup> channel (Kca)
- 2. ATP-sensitive  $K^+$  channel (K<sub>ATP</sub>)
- 3. Inwardly-rectifying K<sup>+</sup> channel (Kir)
- 4. Voltage-gated K<sup>+</sup> channel (Kv)

In the human blood vessels, Kca channel is sub-divided into three types:

- a. Big-conductance (BKca)
- b. Intermediate-conductance (IKca)
- c. Small-conductance (SKca)

BKca channel is widely spread in VSMCs, while IKca and SKca channels are mostly present, in the endothelium (Jakala et al., 2009, Chen et al., 2012, Eichler et al., 2003). The electric conductance is 2–25 Pico Siemens (Ps) for SKca, 25–100 Pico Siemens (Ps) for IKca and 100–300 Pico Siemens (Ps) for BKca channels. In the VSMCs, the BKca channels are  $Ca^{2+}$  and voltage-dependent. The high level of  $Ca^{2+}$  in the cells, trigger these channels and allows the efflux of K<sup>+</sup> ions, at that time hyperpolarization and the closing of the  $Ca^{2+}$  channels occurred. As a result, vasodilation takes place (Gautam et al., 2006). Furthermore, the BKca channels are indirectly activated by the PKA and PKG (Loh et al., 2018). SKca and IKca channels are not voltage-dependent (Burnham et al., 2006, Barfod et al., 2001). These channels are susceptible, to the calmodulin and  $Ca^{2+}$  (García-Pascual et al., 1995, Schumacher et al., 2001). The stimulation of the Kv channels, in the blood vessels, is voltage dependent and linked with the voltage-operated  $Ca^{2+}$  channel (VOCC). The steady state of the membrane potential is reversed by Kv channel. The 4aminopyridine (4-AP) is a Kv channel blocker (Nelson and Quayle, 1995, Loh et al., 2018). By the attachment with PIP<sub>2</sub>, the Kir channel is activated, and the inwards movement of the K<sup>+</sup> ions takes place. Consequently, recovery of the resting membrane potential occurs (Tucker and Baukrowitz, 2008). The Kir channel is activated when the hyperpolarization state occurs, and the influx of K<sup>+</sup> ions takes place. The barium chloride (BaCl<sub>2</sub>) is the only selective blocker, for the Kir channel (Edwards and Weston, 1995, Loh et al., 2018). The K<sub>ATP</sub> channel acts as a weak inwardly rectifying K<sup>+</sup> channel, in resting membrane potential, because of that it is included in the classification of Kir channel family. On the activation, of K<sub>ATP</sub> channel, the K<sup>+</sup> efflux takes place, to keep a negative resting potential and vasorelaxation occurs. The glibenclamide is a selective, K<sub>ATP</sub> channel inhibitor (Jakala et al., 2009).



Figure 2.9 Potassium channels are controlling the tone of the vascular smooth muscle cells (Loh et al., 2018).



Figure 2.10 Function of the potassium channel in the blood vessels (Sobey, 2001).



Figure 2.11 Ion channels and the blood vessel.

In a vascular smooth muscle cell, the  $K_{IR}$ ,  $K_{ATP}$ ,  $K_V$ , and  $BK_{Ca}$  are shown on the top. The voltage-gated Ca<sup>2+</sup> channels are also present, two types of Cl<sup>-</sup> channels, SOC channels (SOCC), and SAC channels (SACC). In the sarcoplasmic reticulum (SR) there are ryanodine receptors (RyR) and inositol 1,4,5-trisphosphate receptors (IP<sub>3</sub>R). The adenylate cyclase (AC), PKA, cAMP-dependent protein kinase, soluble guanylate cyclase (sGC), PKG, cGMP-dependent protein kinase, epoxyeicostetraenoic acid (EETs), phospholipase (PLC), diacylglycerol (DAG), protein kinase C (PKC) on the bottom (Jackson, 2000).

#### 2.6.2 Calcium Channels

There are  $Ca^{2+}$  Channels, in the vascular smooth muscle cell, which are selectively permeable for  $Ca^{2+}$  ions and allows  $Ca^{2+}$  ions to enter the cytosol, depolarization occurs and causing vasoconstriction. There are three main types, of  $Ca^{2+}$  channel:

- 1. Voltage-operated Ca<sup>2+</sup> channels (VOCC)
- 2. Receptor-operated Ca<sup>2+</sup> channels (ROCC)
- 3. Store-operated Ca<sup>2+</sup> channels (SOCC)

Usually, the cytosolic Ca<sup>2+</sup> concentration is raised by two ways

- 1. The influx of  $Ca^{2+}$  ions from the outside
- 2. The intracellular release of  $Ca^{2+}$  from the SR store

The Ca<sup>2+</sup> ions are the most important second messenger, in the blood vessels. The membrane depolarization occurs by the increased level of Ca<sup>2+</sup> ions, in the cells and the up-regulation of Ca<sup>2+</sup>- calmodulin complexes takes place. The activated calmodulin trigger, the MLC kinases (MLCK) to phosphorylate the MLC. As a result, the formation of actin-myosin protein (AMP) occurs, and the contraction of VSMCs takes place, via the mechanism of filament sliding (Jakala et al., 2009, Marchenko and Sage, 1996, Gao et al., 2003, Webb, 2003). The membrane potential, of the VSMCs, is maintained by the VOCC. Generally, the VOCC is known as an Ltype Ca<sup>2+</sup> channel. The concentration of the Ca<sup>2+</sup> ions outside of the cell is thousandtime higher when compared to inside the cell, in normal physiological condition (McFadzean and Gibson, 2002). During the depolarization phase, the Ca<sup>2+</sup> ions from the outside move into the cytosol via VOCC, as a result, vasoconstriction takes place (Patrick, 2002). Other than the VOCC, there is another way for calcium, in the VSMCs, via the ROCC. The intracellular  $Ca^{2+}$  release via ROCC causes the membrane depolarization. The ROCC can induce the intracellular release of  $Ca^{2+}$  ions from the SR store into the cytosol. There are three main types of ROCC receptors such as:

- 1. IP<sub>3</sub>R,
- 2. RyRs, and
- 3. Store-operated Ca<sup>2+</sup> channels (SOCC)

The IP<sub>3</sub>R are present on the SR and triggered by the second messenger, IP<sub>3</sub>, which is formed by stimulated Gq $\alpha$ -protein-coupled receptors. It is the important site, for the release of intracellular Ca<sup>2+</sup> from the SR store, which upsurges the formation of Ca<sup>2+</sup>- calmodulin complexes (McFadzean and Gibson, 2002, Landsberg and Yuan, 2004, Putney et al., 2001). When the concentration of Ca<sup>2+</sup> ions in the cell increases, the RyRs are activated and releases more Ca<sup>2+</sup> ions from the SR store, which is necessary, for muscle contraction. The main function of SOCC is to the replacement of Ca<sup>2+</sup> ions, so it is known as a capacitive-dependent calcium entry channel. In the SR store, the Ca<sup>2+</sup> ions attach to calsequestrin and reducing the concentration of free Ca<sup>2+</sup> ions. Consequently, more calcium is stored (McFadzean and Gibson, 2002, Loh et al., 2018, Swietach et al., 2008).