PRODUCTION OF NOVEL RECOMBINANT ANTI-PFHRP2 V_{NAR} -G1 PROTEIN USING ESCHERICHIA COLI BL21(DE3) EXPRESSION SYSTEM

KOK BOON HUI

UNIVERSITI SAINS MALAYSIA

JUNE 2020



PUSAT PENGAJIAN TEKNOLOGI INDUSTRI UNIVERSITI SAINS MALAYSIA

BORANG PENYERTAAN DISERTAI MUTAKHIR SATU (1) NASKAH

Nama Penyelia: Pn. Wan Zafira Ezza Binti Wan Zakaria	
Bahagian: <u>Teknologi Bioproses</u>	
Saya telah menyemak semua pembetulan/pindaan yang dilaksanakan oleh Encik/Puan/Cik Kok Boon Hui	
mengenai disertasinya sebagaimana yang dipersetujui oleh Panel Pemerik Vocenya.	sa di Viva
2. Saya ingin mengesahkan bahawa saya berpuashati dengan pembet dilaksanakan oleh calon.	ulan/pindaan yang
Sekian, terima kasih.	
40.30 le.	
(WAN ZAFIRA EZZA BINTI WAN ZAKARIA) PENSYARAH DS45 PUSAT PENGAJIAN TENNOLOGI INDUSTRI UNIVERSITI SAINS MALAYSIA, 11800 USM PULAU PINANG	17/7/2020
(Tandatangan dan cop)	Tarikh



PRODUCTION OF NOVEL RECOMBINANT ANTI-PFHRP2 V_{NAR} -G1 PROTEIN USING ESCHERICHIA COLI BL21(DE3) EXPRESSION SYSTEM

by

KOK BOON HUI

A dissertation submitted in the partial fulfillment of the requirements for the degree of Bachelor of Technology (B. Tech) in the field of Bioprocess

Technology

School of Industrial Technology

Universiti Sains Malaysia

June 2020

DECLARATION BY AUTHOR

This dissertation is composed of my original work, and contains no material previously published or written by another person except where due reference has been made in the text. The content of my dissertation is the result of work I have carried out since the commencement of my research project and does not include a substantial part of work that has been submitted to qualify for the award of any other degree or diploma in any university or other tertiary institution.

bommi

KOK BOON HUI

JUNE 2020

ACKNOWLEDGEMENTS

This study was completed with the support and guidance from some organization and individuals. First of all, I would like to express my greatest and sincerest gratitudes to my main supervisor, Pn Wan Zafira Ezza Wan Zakaria from School of Industrial Technology, my co-supervisors, Dr. Leow Chiuan Herng from Institute for Research in Molecular Medicine (INFORMM) and Dr. Warren Lee Xian Liang from Usains Biomics Laboratory Testing Services Sdn. Bhd. Pn Zafira was a supervisor full of patience and she always provide useful information and helpful guidance especially in the statistical part whereby a new software was applied in analysing my study. Besides, she also provided many useful comments while reviewing my thesis.

Next, I am very grateful and appreciate to work under supervision of Dr Leow with this challenging but interesting study. He was very responsible and patience while giving advice when I faced problems during the experiments, always follow up with my progress and gave me a lot of detailed guidance which really helped a lot in my experiment especially the part related to DNA technology. Besides, he also helped a lot in improving my thesis writing. Dr Warren, a diligent co-supervisor who spent a lot of precious time in guiding and teaching me despite his busy schedule. He always shared his opinions and some valuable suggestions especially in the downstream part based on his experiences which was really useful in my study. Also, I would like to thank him for improving my thesis writing.

On the other hand, I would like to extend my gratitude and thankfulness to

INFORMM, Universiti Sains Malaysia (USM) for allowing me to use the laboratory

equipments and facilities such as thermocycler, high speed centrifuge,

microcentrifuge, mini gel electrophoresis system, gel imager, incubator shaker,

spectrophotometer, semi-dry transfer cell and chemiluminescence scanner to

complete my final year project. Besides, I also feel grateful and appreciate for the

help and guidance given by Dr Leow's PhD student, Pn. Nor Raihan Mohammad

Shabani and medical laboratory technologists such as Pn. Izzati Zahidah Binti Abdul

Karim and En. Mohamed Qais Bin Abu Bakar.

Lastly, I would to express my heartfelt gratitude to my dearest family

members who have always been staying by my side and gave me lots of moral

support and motivations. Also, a million thanks to my friends who always support

and encourage me to move forward. My experiment and thesis would not be

successful without the help and guidance from those mentioned. Once again, many

thanks to the ones mentioned.

KOK BOON HUI

June 2020

iv

TABLE OF CONTENTS

	Page
Acknowledgements	iii
Table of Contents	v
List of Tables	X
List of Figures	xii
List of Symbols and Abbreviations	XV
Abstrak	XX
Abstract	xxi
CHAPTER 1 INTRODUCTION	
1.1 Research background	1
1.2 Problem statement	3
1.3 Research objectives	4
CHAPTER 2 LITERATURE REVIEW	
2.1 Malaria	5
2.1.1 An introduction to malaria	5
2.1.2 Plasmodium falciparum infection	6
2.1.3 Malaria rapid diagnostic tests	8
2.2 Antibody	11
2.2.1 Antibody structure	11
2.2.2 Monoclonal antibody and its limitations	12
2.3 Single domain antibody (sdAb) as an alternative to mAb	13
2.3.1 The unique characteristics of sdAbs	13
$2.3.2$ Discoveries on shark variable new antigen receptor (V_{NAR})	14
2.4 Bacteria expression system	18

2.4.1 Types of expression system	18
2.4.2 Escherichia coli strain as expression host	20
2.4.3 Escherichia coli strain as cloning host	21
2.4.4 pET system in protein expression	22
2.5 Factors affecting protein expression	23
2.5.1 Temperature	24
2.5.2 Inducer concentration	26
CHAPTER 3 MATERIALS AND METHODS	
3.1 Preparation of media	27
3.1.1 Luria-Bertani medium (LB medium)	27
3.1.2 LB agar medium	27
3.1.3 LB kanamycin medium	27
3.1.4 LB kanamycin agar medium	28
3.1.5 LB ampicillin medium	28
3.2 Preparation of stock solutions	28
3.2.1 Calcium chloride (CaCl ₂) solution	28
3.2.2 Magnesium chloride (MgCl ₂) solution	28
3.2.3 Sodium chloride (NaCl) solution at 5 M	29
3.2.4 Monosodium phosphate (NaH ₂ PO ₄) solution at 1 M	29
3.2.5 Disodium phosphate (Na ₂ HPO ₄) solution at 1 M	29
3.2.6 2-(N-Morpholino)ethanesulfonic acid (MES free acid) solution	29
at 0.5 M	
3.2.7 2-(N-Morpholino)ethanesulfonic acid sodium salt (MES sodium	29
salt) solution at 0.5 M	
3.2.8 Ammonium persulphate (APS) solution at 10% (w/v)	29

	3.2.9 Sodium dodecyl sulphate (SDS) solution at 10% (w/v)	30
	3.2.10 Bovine serum albumin (BSA) solution	30
3.3 Preparation of buffers		30
	3.3.1 Wash buffer for purification (pH 7.4)	30
	3.3.2 Elution buffer for purification (pH 7.4)	30
	3.3.3 MES buffer for resin regeneration (pH 5.0)	31
	3.3.4 Phosphate-buffered saline (PBS) solution (10x)	31
	3.3.5 Tris buffer at 1.5 M (pH 8.8)	31
	3.3.6 Tris buffer at 1.0 M (pH 6.8)	31
	3.3.7 SDS sample buffer (6x)	32
	3.3.8 Tris-glycine running buffer (10x, pH 8.3)	32
	3.3.9 Tris-buffered saline (TBS) solution (10x, pH 7.6)	32
	3.3.10 Transfer buffer (10x)	33
	3.3.11 TBST buffer (1x)	33
	3.3.12 Blocking buffer	33
3.4	Activation of recombinant <i>E. coli</i> DH5α cells in starter culture	33
3.5	Isolation of recombinant <i>E. coli</i> DH5α cells	34
3.6	Polymerase chain reaction (PCR)	35
3.7	Agarose gel electrophoresis	36
3.8	Plasmid purification and plasmid extraction	37
3.9	Determination of DNA concentration	39
3.10	Transformation of pET28a (+)-anti-PfHRP2 V _{NAR} -G1 plasmid into	39
	DH5α and BL21(DE3) competent cells	
3.11	Expression of recombinant anti-PfHRP2 V _{NAR} G1 protein	40
3.12	2 Statistical analysis on the expression factors	41

	3.13 Cell lysis and extraction of anti-PfHRP2 V _{NAR} G1	42
	3.14 Purification of anti-PfHRP2 V _{NAR} G1	43
	3.15 Sodium Dodecyl Sulfate-Polyacrylamide Gel Electrophoresis (SDS-	44
	PAGE)	
	3.15.1 Gel preparation	44
	3.15.2 Sample preparation	45
	3.15.3 Running the gel	46
	3.15.4 Gel staining and destaining	46
	3.15.5 Gel imaging	47
	3.16 Western blot	47
	3.17 Bradford assay for protein quantification	49
CHA	APTER 4 RESULTS AND DISCUSSION	
	4.1 Activation of recombinant <i>E. coli</i> DH5α cells in starter culture	52
	4.2 Isolation of recombinant <i>E. coli</i> DH5α cells	54
	4.3 Determination of DNA concentration	56
	4.4 Gel imaging on PCR product	57
	4.5 Transformation of pET28a (+1-anti-PfHRP2 V_{NAR} -G1 plasmid into	61
	DH5α and BL21(DE3) competent cells	
	4.6 Expression of recombinant anti-PfHRP2 V _{NAR} G1 protein	66
	4.7 Statistical analysis on the effects of expression factors	70
	4.7.1 Effects of expression factors on pellet weights	71
	4.7.2 Effects of expression factors on absorbance reading	77
	4.8 Sodium Dodecyl Sulfate-Polyacrylamide Gel Electrophoresis (SDS-	87
	PAGE)	
	4.9 Western blot	92

4.10 Bradford assay for protein quantification	93
CHAPTER 5 CONCLUSIONS AND RECOMMENDATIONS FOR FUTURE	
RESEARCH	
5.1 Conclusions	95
5.2 Recommendations for future research	97
REFERENCES	98
APPENDICES	120

LIST OF TABLES

Tabl	e Caption	Page
2.1	V_{HH} expression in E. coli BL21(DE3) with different expression	20
	vectors.	
3.1	The list of primers with its sequences.	35
3.2	PCR mixture for one reaction.	35
3.3	Thermocycling conditions for anti-PfHRP2 V_{NAR} -G1 gene	36
	amplification.	
3.4	Induced expression conditions for transformed BL21(DE3)	41
	subculture cultivation.	
3.5	The volumes of DNase I and B-PER reagent required to add into per	43
	gram of pellet.	
3.6	Components of 5% stacking gel.	45
3.7	Components of 14% stacking gel.	45
3.8	Components of 12% resolving gel.	45
3.9	The volumes of protein samples and sample buffer required for	46
	sample loading.	
4.1	Statistical analysis of absorbance readings of recombinant DH5 α	54
	cells cultivated in different antibiotic supplemented medium after 24	
	hours incubation.	
4.2	Statistical analysis of absorbance readings of recombinant DH5 α	56
	cells cultivated in 5 tubes of LB kanamycin medium after 24 hours	
	incubation.	
4.3	The statistical analysis of purified plasmid samples on their nucleic	57
	acid concentration (ng/ μ L) and A ₂₆₀ /A ₂₈₀ ratio.	

4.4	Means and summary statistics by group for dependent factor pellet	71
	weight.	
4.5	Analysis of temperature and IPTG concentration on the pellet	73
	weights using two-way ANOVA.	
4.6	Coefficient of multi regression model (Appendix P).	76
4.7	Means and summary statistics by group for dependent factor	78
	absorbance reading.	
4.8	Analysis of temperature and IPTG concentration on the absorbance	80
	reading using two-way ANOVA.	
4.9	Coefficient of multi regression model (Appendix P).	82
4.10	The correlation between independent variables and dependent	85
	variables $(N = 12)$.	

LIST OF FIGURES

Figu	re Caption	Page
2.1	Development stages of PfHRP2 in human and mosquito.	8
2.2	The Y-shaped basic structure of antibody.	12
2.3	The schematic diagram of human IgG (left), shark IgNAR (middle)	16
	and V_{NAR} (right).	
4.1	The turbidity in different types of antibiotic supplemented	53
	medium at 0 hour (A) and after 24 hours (B).	
4.2	Colonies of recombinant E. coli DH5α cells observed on LB	55
	kanamycin plate after 24 hours of incubation at 37°C.	
4.3	The cultivation of recombinant DH5 α cells in 10 tubes of 5.0 mL	55
	LB kanamycin medium with initial turbidity conditions at 37°C with	
	200 rpm shaking at 0 hour.	
4.4	The turbidity in the first five tubes of cultivation medium with	56
	recombinant DH5α cells after 24 hours.	
4.5	Agarose gel electrophoresis of samples with recombinant DH5 α	58
	cells grown in LB kanamycin medium.	
4.6	Agarose gel electrophoresis of samples with recombinant DH5 α	60
	cells grown in LB kanamycin medium and plasmid samples.	
4.7	Agarose gel electrophoresis of samples from recombinant DH5 α	61
	cells grown in LB kanamycin medium, recombinant DH5α cells	
	grown in LB ampicillin medium and plasmid samples.	
4.8	Colonies of transformed DH5 α and BL21(DE3) cells successfully	62
	grown on LB kanamycin plate after 24 hours incubation at 37°C.	

4.9	Agarose gel electrophoresis of colony PCR samples from	63
	transformed DH5 α and BL21(DE3) cells grown in LB kanamycin.	
4.10	Turbidity in cultivation of transformed BL21(DE3) and DH5 α cells	65
	in different antibiotic supplemented medium after 24 hours of	
	incubation at 37°C with 200 rpm shaking	
4.11	Mean of absorbance readings obtained in LB ampicillin and LB	65
	kanamycin medium cultivation of transformed BL21(DE3) cells	
	after 24 hours of incubation at 37°C with 200 rpm shaking.	
4.12	Mean of absorbance readings obtained in LB ampicillin and LB	66
	kanamycin medium cultivation of transformed DH5α cells after 24	
	hours of incubation at 37°C with 200 rpm shaking.	
4.13	Turbidity in subculture cultivation of transformed BL21(DE3) in	69
	100 mL LB kanamycin at 0 hour incubation with 200 rpm shaking.	
4.14	Turbidity in subculture cultivation of transformed BL21(DE3) in	70
	100 mL LB kanamycin after 24 hours incubation with 200 rpm	
	shaking.	
4.15	Plot of average pellet weight (g) against IPTG concentration (mM).	72
4.16	Scatterplot of residuals against fitted values.	74
4.17	Normal probability plot for pellet weight (response).	74
4.18	Plot of multi variable regression with interaction between	77
	temperature and IPTG concentration.	
4.19	Plot of average absorbance reading against IPTG concentration	79
	(mM).	
4.20	Scatterplot of residuals against fitted values.	81
4.21	Normal probability plot for absorbance reading (response).	81

4.22	Plot of multi variable regression with interaction between	83
	temperature and IPTG concentration.	
4.23	SDS-PAGE of the expressed recombinant anti-PfHRP2 VNAR-G1	88
	protein samples on 10% polyacrylamide pre-cast gel.	
4.24	SDS-PAGE of the expressed recombinant anti-PfHRP2 VNAR-G1	91
	protein samples on 14% polyacrylamide gel.	
4.25	SDS-PAGE of the expressed recombinant anti-PfHRP2 VNAR-G1	92
	protein samples on 12% polyacrylamide gel.	
4.26	Western blot analysis of purified recombinant anti-PfHRP2 VNAR-	93
	G1 protein immunodetected using TMB substrate (a) and ECL	
	substrate (b).	

LIST OF SYMBOLS AND ABBREVIATIONS

Symbol Caption Positive/plus Negative/minus Plus-minus \pm Times × More than Less than < Percentage % Infinity ∞ Degree Celsius $^{\mathrm{o}}\mathrm{C}$ Relative centrifugal force $\times g$ K Potassium Na Sodium **Abbreviation** Caption Ratio of absorbance 260 nm to A_{260}/A_{280} absorbance 280 nm Aldolase Fructose 1,6-biphosphate aldolase Analysis of variance **ANOVA** Ammonium persulphate **APS** Base pair bp **BSA** Bovine serum albumin

Calcium

Ca

CaCl₂ Calcium chloride

CBB Coomassie Brilliant Blue

cDNA Complementary DNA

CDRs Complementary determining regions

CFU/mL Colony forming unit per millilitre

C_{NAR} Constant new antigen receptor

DNA Deoxyribonucleic acid

E. coli Escherichia coli

EB Elution buffer

ECL Enhanced chemiluminescence

ELISA Enzyme-linked immunosorbent assay

Fab Antigen-binding fragment

Fc Crystallizable fragment

FR Framework region

Fv Variable fragment

g Gram

g/L Gram per litre

HC Heavy chains

HCAbs Heavy-chain-only antibodies

HCl Hydrochloric acid

His-tag Histidine-tag

HRP Horseradish peroxidase

Ig Immunoglobulin

IgNAR Immunoglobulin new antigen receptor

IMAC Immobilized metal affinity

chromatography

IPTG Isopropyl-β-D-thiogalactoside

KCl Potassium chloride

kDa Kilodalton

KH₂PO₄ Monopotassium phosphate

L Litre

LB Luria-Bertani

LC Light chains

M Molar

mA Milliampere

mAbs Monoclonal antibodies

Mean Sq Mean of square

MES free acid 2-(N-Morpholino)ethanesulfonic acid

MES sodium salt 2-(N-Morpholino)ethanesulfonic acid

sodium salt

Mg Magnesium

mg Milligram

mg/L Milligram per litre

mg/mL Milligram per millilitre

MgCl₂ Magnesium chloride

mL Millilitre

μg Microgram

μg/mL Microgram per millilitre

μL Microlitre

μm Micrometer

mM Millimolar

MWCO Molecular weight cut-off

N Number of samples

Na₂HPO₄ Disodium phosphate

NaCl Sodium chloride

NaH₂PO₄ Monosodium phosphate

NaH₂PO₄·H₂O Sodium dihydrogen phosphate

monohydrate

ng/μL Nanogram per microlitre

nm Nanometer

OD Optical density

Omp T Outer membrane protein T

P. falciparum Plasmodium falciparum

PBS Phosphate-buffered saline

PCR Polymerase chain reaction

PfHRP2 Plasmodium falciparum histidine-rich

protein 2

pLDH Plasmodium lactate dehydrogenase

PMSF Phenylmethylsulfonyl fluoride

Pr Probability

R² R-squared

RDTs Rapid diagnostic tests

RNA Ribonucleic acid

RNase Ribonuclease

rpm Rounds per minute

RSE Relative standard error

SB Super broth

scFv Single chain variable fragment

sdAbs Single-domain antibodies

SDS Sodium dodecyl sulphate

SDS-PAGE Sodium dodecyl sulfate-polyacrylamide

gel electrophoresis

SOC Super optimal broth with catabolite

repression

Sum Sq Sum of square

TAE Tris-acetate-EDTA

TB Terrific broth

TBS Tris-buffered saline

TEMED Tetramethylethylenediamine

TMB 3,3',5,5'-Tetramethylbenzidine

V Volt

v/v Volume to volume

VBNC Viable but non-culturable

V_H Heavy chain

V_{HH} Heavy chain single variable domain

V_L Light chain

 V_{NAR} Variable domain of new antigen receptor

WB Washing buffer

PENGHASILAN ANTI-PFHRP2 V_{NAR} -G1 PROTEIN REKOMBINAN NOVEL MENGGUNAKAN SISTEM EKSPRESI ESCHERICHIA COLI BL21(DE3)

ABSTRAK

Ujian diagnostik segera malaria (RDT) bertindak sebagai imunoassay berasaskan antibodi penting untuk diagnosis segera malaria. Antibodi monoklonal konvensional (mAbs) digunakan secara meluas dalam RDT tetapi ia mudah merosot pada suhu persekitaran tinggi. Oleh itu, V_{NARS} dari ikan yu mungkin merupakan alternatif yang baik untuk mAbs kerana kestabilan haba dan kekuatan gabungan dengan antigen yang lebih tinggi. Dalam kajian ini, anti-PfHRP2 V_{NAR}-G1 protein rekombinan akan dihasilkan dalam sistem ekspresi E. coli BL21(DE3) melalui pelbagai langkah seperti pengasingan sel rekombinan, PCR, elektroforesis gel agarosa, pengekstrakan plasmid, transformasi dan ekspresi protein. Selain itu, kesan gabungan suhu dan kepekatan IPTG terhadap kepadatan sel rekombinan BL21(DE3) berdasarkan bacaan serapan dan berat basah sel dianalisis menggunakan perisian R. Berdasarkan analisis statistik ANOVA 2-arah dan regresi berbilang pemboleh ubah, kedua-dua faktor ekspresi mempunyai interaksi gabungan yang sangat signifikan (p < 0.05) terhadap bacaan serapan dan berat basah sel. Analisis korelasi antara kepekatan IPTG dan bacaan serapan adalah signifikan (p < 0.05) dengan pekali korelasi Pearson yang tinggi (0.9512). Kemunculan anti-PfHRP2 V_{NAR}-G1 protein rekombinan dengan ukuran molekul sekitar 12 kDa dikesan dan disahkan melalui analisis SDS-PAGE dan western blot. Kepekatan protein ditentukan sebagai 0.209 mg/mL dari 0.406 g ekstrak sel kasar. Kesimpulannya, semua objektif dalam kajian ini tercapai dan sdAb rekombinan dari V_{NAR} ikan yu khusus untuk gabungan PfHRP2 berjaya dihasilkan dalam E. coli BL21(DE3) sebagai sumber ekspresi.

PRODUCTION OF NOVEL RECOMBINANT ANTI-PFHRP2 V_{NAR} -G1 PROTEIN USING ESCHERICHIA COLI BL21(DE3) EXPRESSION SYSTEM

ABSTRACT

Malaria rapid diagnostic tests (RDTs) act as important antibody-based immunoassays for prompt malaria diagnosis. Conventional monoclonal antibodies (mAbs) are widely used in RDTs but it can be easily degraded at high ambient temperatures. Hence, the shark V_{NARS} might be good alternatives to mAbs due to its higher thermal stability and binding affinity with antigens. In this study, the recombinant anti-PfHRP2 V_{NAR}-G1 protein was produced in E. coli expression system through various steps such as recombinant cell isolation, PCR, agarose gel electrophoresis, plasmid extraction, transformation and protein expression. Besides, the combinatorial effects of temperature and IPTG concentration towards the cell density of recombinant BL21(DE3) based on the absorbance readings and cell wet weights were investigated using software R. Based on the statistical analysis of 2-way ANOVA and multivariable regression, both expression variables had highly significant combined interactions (p < 0.05) towards absorbance readings and cell wet weights. There was significant and strong positive correlation between IPTG concentrations and absorbance readings (p < 0.05, r = 0.9512). The presence of recombinant anti-PfHRP2 V_{NAR}-G1 protein with a molecular size of about 12 kDa was detected and confirmed through SDS-PAGE and western blot analysis. The protein concentration was determined as 0.209 mg/mL from 0.406 g of crude cell extract. In conclusion, all the objectives in this study were achieved and the recombinant sdAb from shark V_{NAR} specific for PfHRP2 binding was successfully produced in E. coli BL21(DE3) as the expression host.