PREDICTING GENTAMICIN INDUCED NEPHROTOXICITY USING PHARMACOMETABONOMIC APPROACH

FATIMATUZZAHRA' ABD. AZIZ

UNIVERSITI SAINS MALAYSIA

2019

PREDICTING GENTAMICIN INDUCED NEPHROTOXICITY USING PHARMACOMETABONOMIC APPROACH

by

FATIMATUZZAHRA' ABD. AZIZ

Thesis submitted in fulfilment of requirements for the Degree of Doctor of Philosophy

March 2019

ACKNOWLEDGEMENT

My heartiest thanks go to my parents and my husband for their love and having my back throughout my PhD project. They supported me emotionally at the moments I was desperate and hopeless. Without their support, it was impossible for me to tackle the problems I encountered during the study period. I would like to specially thank my supervisor Associate Professor Dr Baharudin Ibrahim for his precious advice and guidance throughout the whole study period. It was my fortune, honour and pleasure to have Professor Dr Azmi Sarriff and Associate Professor Dr Vikneswaran Murugaiyah as my co-supervisor. My sincerest thanks to Dr. Nor Azlina Khalil as my field supervisor. I am grateful to Dr. Teh Chin Hoe who opened my horizons to the wonderful world of NMR spectroscopy. I would like to express my gratitude to Animal Research Centre (ARC) Advanced Medical and Dental Institute (AMDI) for their helpfulness and kindness during animal study. I would like to thank Orthopaedic team in Hospital Kulim, Kedah for boundless assistance in conducting human study. I also wish to thank the helpful staff of the School of Pharmaceutical Sciences for their kindness and flexibility in providing me with the facilities and their technical support: Mr. Ahmad Zainudin and his team and all the rest of the staff. My experience in the world of science would not have been pleasant without my caring and helpful colleagues, Hadzliana Zainal, Nuridah Ahamed, Dr. Balamurugun Tangiisuran. I wish to thank my co-supervisors' postgraduate team, Foroogh, Arwa, Hamza and Abu Bakar. I wish them and all the rest of my friends in the school of Pharmaceutical Sciences USM all the best of luck throughout their life.

TABLE OF CONTENTS

ACKNOWLEDGEMENT	ii
TABLE OF CONTENTS	iii
LIST OF TABLES	viii
LIST OF FIGURES	xiii
LIST OF ABBREVIATIONS	xix
LIST OF SYMBOLS	xxvii
ABSTRAK	xxix
ABSTRACT	xxxi

CHAPTER 1 - INTRODUCTION

1.1	Limitations of Current Practice	2
1.2	Statement of the Problem	2
1.3	Study Hypothesis	2
1.4	Study Objectives	3
	1.4.1 General Objective	3
	1.4.2 Specific Objective(s)	3

CHAPTER 2 - LITERATURE REVIEW

2.0	The Kidneys	4
2.1	Acute Kidney Injury (AKI)	
	2.1.1 Epidemiology	
	2.1.2 Drug Induced Nephrotoxicity	9
2.2	Aminoglycosides	
	2.2.1 Gentamicin	

2.3	3 Current Practice in Diagnosis and Management of Nephrotoxicity cause by Gentamicin		
	2.3.1	Current Practice to Detect Nephrotoxicity caused by Gentamicin17	
	2.3.2	Current Management of Nephrotoxicity Induced by Gentamicin 19	
	2.3.3	Update on Kidney Injury Biomarkers	
2.4	The Po	otential Role of Pharmacometabonomics in Nephrotoxicity24	
	2.4.1	Application of Pharmacometabonomics in Personalised Medicine25	
	2.4.2	Factors that Affect Biomarker/Metabolite Identification	
	2.4.3	Method Optimization for Pharmacometabonomics	
	2.4.4	Analytical Technologies used in Pharmacometabonomics	

CHAPTER 3 - MATERIALS AND METHODS

3.0	Introc	luction	. 44
3.1	Phase Samp	I: Impact on Different Internal Standards and Preservatives Used le Storage in Human Serum and Urine Samples	1 in . 44
	3.1.1	Assessment of Different Internal Standards	. 44
	3.1.2	Comparing the Addition of Sodium Azide Before and After Storage	. 45
	3.1.3	Sample Collection and Handling	. 45
3.2	Phase Induce	II: Discovery and Evaluation of Metabolite to Predict Gentami ed Neprotoxicity	icin . 50
	3.2.1	Chemicals and solvents	. 50
	3.2.2	Animal and sample size	. 51
3.3	Phase	III: Validation and Quantification Phase	. 56
	3.3.1	Phase III (1): Validation Phase in Sprague Dawley Rats	. 56
	3.3.2	Phase III (2): Validation and Quantification Phase in Human Group	. 61
	3.3.3	Evaluation of Patients	. 63
	3.3.4	Sample Collection and Storage	. 63
3.4	NMR	Sample Preparation and Spectral Acquisition	. 64

	3.4.1	Urine Sample Preparation	. 64
	3.4.2	Blood Sample Preparation	65
	3.4.3	NMR Spectral Acquisition	65
	3.4.4	Statistical Analyses	. 68
3.5	Data f	iltering	72
	3.5.1	Statistical Analysis	76
	3.5.2	Metabolite Identification	. 77
	3.5.3	Validation of the Pre-dose Serum and Urine Model	. 77
3.6	Ethica	l Approval	77

CHAPTER 3 - RESULTS

4.0	Introd	uction
4.1	Part I: and U	Impact of Internal Standards and Preservatives on Human Serum rine Metabolites
	4.1.1	Impact of Different Internal Standards on Human Serum and Urine Samples
	4.1.2	Impact of Preservatives Used in Sample Storage on Human Serum and Urine Samples
4.2	Part I Nephr	I: Discovery and Evaluation of Metabolite/Biomarkers to Predict otoxicity Induced by Gentamicin in Sprague-Dawley Rats93
	4.2.1	Introduction
	4.2.2	Classification of Acute Kidney Injury (AKI) in Sprague-Dawley Rats
	4.2.3	Pre-dose Serum and Urine Models to Discriminate Between SD Rats with and without Nephrotoxicity
	4.2.4	Pre-dose serum
	4.2.5	Pre-dose urine
	4.2.6	Association of Identified Pre-dose Metabolites and Model in Serum and Urine with Relevant Clinical Chemistry and Histopathology of Kidneys
	4.2.7	Changes in Metabolites of Serum and Urine prior to and after Treatment with Gentamicin

	4.2.8 Identification of Putative Biomarkers to Predict Gentamicin Induced Nephrotoxicity
4.3	Part III : Validation of Possible Metabolites in Predicting Gentamicin Induced Nephrotoxicity
	4.3.1 Validation of Pre-dose Serum Biomarkers
	4.3.2 Validation of Pre-dose Urine Biomarkers
	4.3.3 Part III (2): Validation of Possibility of Biomarkers Predicting Gentamicin Induced Nephrotoxicity in Humans

CHAPTER 5 – DISCUSSION

5.0	Introd	luction
5.1 Part I: Method Optimisation		Method Optimisation
	5.1.1	Justification of Method Optimisation for Internal Standards and Preservatives Used in Sample Storage161
	5.1.2	DSA as Internal Standard for Serum Analysis of Rats and Human Damples and TSP as Internal Standard for Urine Analysis of Rats and Human Samples
	5.1.3	Impact on Preservatives Used in Sample Storage 163
5.2	Part II	: Discovery of Metabolites 165
	5.2.1	Development of Histopathology Scoring System for Proximal Tubules
	5.2.2	Discovery of Metabolites using 40 mg Gentamicin as a Nephrotoxicity Model in SD Rats
	5.2.3	Affirmation of Nephrotoxin Model with High Dose of Gentamicin167
	5.2.4	Development of Classification of AKI in SD Rats 168
	5.2.5	Model of Pre-dose Serum and Urine Metabolites using Pharmacometabonomic Assessmen
	5.2.6	Fingerprinting Model of Pre-dose Serum and Urine Metabolites to Discriminate Rats with Gentamicin Induced Nephrotoxicity 170
	5.2.7	Discriminatory Bins 171
	5.2.8	Identification Models of Pre-dose Serum and Urine Metabolites to Discriminate Rats with Gentamicin Induced Nephrotoxicity 172

	5.2.9 Association and Variation in Metabolites of Models for Pre-dos Serum and Urine Metabolites	e 174
	5.2.10 Identified Putative Metabolites in Pre-dose Serum and Urine Samples	175
	5.2.11 Combinatory Metabolites as Diagnostic Biomarkers for Detection of Gentamicin Induced Nephrotoxicity	ion 175
	5.2.12 Anaerobic glycolisis	177
	5.2.13 Impaired Purine Metabolism and Hyperuricemia	178
	5.2.14 Tricarboxylic Acid Cycle	179
	5.2.15 Betaine and TMNO	179
	5.2.16 Depressed Urinary 4-Pyridoxic Acid	180
	5.2.17 Other Implied Pathways of the Discovered Metabolites	181
5.3	Part III: Validation of Metabolites	182
	5.3.1 Validation of AKI Models in Rats using SIMCA [®]	182
	5.3.2 Validation of AKI Models in Human using SIMCA [®]	183

REFERENCES1	190
-------------	-----

APPENDICES

LIST OF TABLES

		Page
Table 2.1	Definition of AKI	8
Table 2.2	Risk factors for drug induced nephrotoxicity	11
Table 2.3	Types of aminoglycosides	12
Table 2.4	Indication for parenteral gentamicin	15
Table 2.5	Overview table of equation to estimate kidney function	20
Table 2.6	Characteristics of ideal biomarker for acute kidney injury.	22
Table 2.7	New biomarkers to predict AKI	22
Table 2.8	Example of pharmacometabonomic studies	26
Table 2.9	Comparing of NMR and MS in pharmacometabonomics	39
Table 2.10	Acceptable values of goodness of fit (R2) and goodness of prediction (Q2) based on type of study	40
Table 3.1	Urine sample collection protocol	46
Table 3.2	Constituent of phosphate buffer	50
Table 3.3	Constituent of phosphate buffer and preservative	50
Table 3.4	Scoring system for proximal tubules	56

Table 3.5	American Society of Anaesthesiologists Physical Status (*ASA PS) Classification System.	62
Table 3.6	Nephrotoxic drugs	63
Table 3.7	NMR parameter for method optimization, animal and human experiment	69
Table 3.8	Bucket method type with description	73
Table 3.9	Type and description of scales	74
Table 4.1	Peaks with contribution plot values of more than 1 in DSA and TSP for serum samples	84
Table 4.2	Peaks of contribution plot with values of more than 1 in DSA and TSP for human urine samples	88
Table 4.3	The peaks with VIP value more than 1 in serum samples.	90
Table 4.4	The peaks with VIP value more than 1 in urine samples.	92
Table 4.5	Description of AKI toxicity based on each criterion.	94
Table 4.6	The median and interquartile range (IQR) for each criterion in 3 different groups	95
Table 4.7	Summary of all fingerprinting models of pre-dose serum for number of pre-dose metabolites with VIP of more than 1, score plot model value, accuracy and permutation values	101
Table 4.8	Summary of all identification models of pre-dose serum for number of pre-dose metabolites with VIP of more than 1, score plot model value, accuracy and permutation values	107
Table 4.9	List of possible pre-dose metabolites in serum according to identification model of pre-dose serum	108

Table 4.10	Summary of all fingerprinting models of pre-dose urine for number of pre-dose metabolites with VIP values of more than 1, score plot model value, accuracy and permutation values	113
Table 4.11	Summary of all identification models of pre-dose urine for number of pre-dose metabolites with VIP of more than 1, score plot model value, accuracy and permutation values	119
Table 4.12	List of possible pre-dose metabolites in urine according to identification modes of pre-dose urine	120
Table 4.13	Correlation table of fingerprinting models of pre-dose serum with relevant clinical chemistry and histopathology of kidneys.	121
Table 4.15	Correlation table of identification models of pre-dose serum with relevant clinical chemistry and histopathology of kidneys	123
Table 4.16	Correlation table of each metabolite in identification models of pre-dose serum with relevant clinical chemistry and histopathology of kidneys	124
Table 4.17	Correlation table of fingerprinting models of pre-dose urine with relevant clinical chemistry and histopathology of kidneys.	125
Table 4.18	Correlation table of each metabolite identification models of pre-dose urine with relevant clinical chemistry and histopathology of kidneys	127
Table 4.19	Results of Mann-Whitney test comparing PModXPS values of toxic and non toxic clusters in pre-serum models	128
Table 4.20	Results of the Mann-Whitney test comparing PModXPS values of toxic and non toxic clusters in pre-urine models	133
Table 4.21	Potential pre-dose serum metabolites and their molecular formula	145
Table 4.22	Potential pre-dose urine metabolites and their molecular formula	147

Table 4.23	Sensitivity, specificity and accuracy of serum creatinine	148
	model of pre-dose serum in SD rats using predictions set	
	in SIMCA® software	

- Table 4.24Sensitivity, specificity and accuracy of BUN model of
pre-dose serum in SD rats using predictions set in
SIMCA® software149
- Table 4.25Sensitivity, specificity and accuracy of urine output
model of pre-dose serum in SD rats using predictions set
in SIMCA® software150
- Table 4.26Sensitivity, specificity and accuracy of three AKI150criteria, except for histopathology model, of pre-dose
serum in SD rats using predictions set in SIMCA®
softwaresoftware
- Table 4.27Sensitivity, specificity and accuracy of serum creatinine
model of pre-dose urine in SD rats using predictions set
in SIMCA® software151
- Table 4.28Sensitivity, specificity and accuracy of BUN model of
pre-dose urine in SD rats using predictions set in
SIMCA® software152
- Table 4.29Sensitivity, specificity and accuracy of urine output
model of pre-dose urine in SD rats using predictions set
in SIMCA® software152
- Table 4.30Sensitivity, specificity and accuracy of three AKI153criteria, except for the histopathology model, of pre-
dose urine in SD rats using predictions set in SIMCA®
softwaresoftware
- Table 4.31Demographics and clinical findings of treatment and154control subjects
- Table 4.32Sensitivity, specificity and accuracy of SCr156identification model of pre-dose serum in humans using
predictions set in SIMCA® software156
- Table 4.33Sensitivity, specificity and accuracy of BUN156identification model of pre-dose serum in humans using
predictions set in SIMCA® software156

- Table 4.34Sensitivity, specificity and accuracy of UO157identification model of pre-dose serum in human using
prediction set in SIMCA® software.157
- Table 4.35Sensitivity, specificity and accuracy of combination of
three AKI criteria, except for histopathology
identification model, of pre-dose serum in humans using
predictions set in SIMCA® software157
- Table 4.36Sensitivity, specificity and accuracy of SCr158identification model of pre-dose urine in humans using
predictions set in SIMCA® software158
- Table 4.37Sensitivity, specificity and accuracy of BUN158identification model of pre-dose urine in humans using
predictions set in SIMCA® software158
- Table 4.38Sensitivity, specificity and accuracy of UO158identification model of pre-dose urine in humans using
predictions set in SIMCA® software158
- Table 4.39Sensitivity, specificity and accuracy of combination of
three AKI criteria, except for histopathology
identification model, of pre-dose urine in humans using
predictions set in SIMCA® software159

LIST OF FIGURES

Page

Figure 2.1	Human kidney anatomy	5
Figure 2.2	Secretion and reabsorption of various substances throughout the nephron in kidney	7
Figure 2.3	Chemical structure of Aminoglycosides antibiotics	13
Figure 2.4	Factors affecting variation in metabolic profiling and identification	30
Figure 2.5	Possible content in human urine	32
Figure 3.1	Outline of collection and handling of serum samples	47
Figure 3.2	Outline of collection and handling of urine samples	48
Figure 3.3	Flow chart for sample collection in SD rat	53
Figure 3.4 (a)	Schematic picture of grading score for proximal convoluted tubular damage	57
Figure 3.4 (b)	Schematic picture of grading score for proximal convoluted tubular damage	58
Figure 3.4 (c)	Schematic picture of grading score for proximal convoluted tubular damage	59
Figure 3.4 (d)	Schematic picture of grading score for proximal convoluted tubular damage	60

xiii

Figure 3.5	Interface of TopSpin 3.5 pl 5 software (Bruker BioSpin, Germany)	70
Figure 3.6	Interface of Amix-Viewer version 3.9.15 software (Bruker BioSpin, Germany)	71
Figure 3.7	Interface of SIMCA 14.0 software (Umetrics, Sweden)	71
Figure 3.8	Selection of data type and bucketing techniques in AMIX software	73
Figure 3.9	Steps to metabolite identification	79
Figure 4.1	A Principal component analysis (PCA-X) score plot of differences between DSA and TSP in human serum samples	82
Figure 4.2	Spectra for comparison of the chemical shift between different internal standards in human serum samples	83
Figure 4.3	Box plot of half width of DSA and TSP at 0 ppm in human serum samples	85
Figure 4.4	A Principal component analysis (PCA-X) score plot of differences between DSA and TSP in human urine samples	86
Figure 4.6	Box plot of half width of DSA and TSP at 0 ppm in human urine samples	87
Figure 4.7	A PLS-DA score plot of serum stored with and without sodium azide	90
Figure 4.8	PLS-DA score plot of urine stored with and without sodium azide	91

Figure 4.9	Step by step development of pre-dose serum and urine models to discriminate between SD rats with and without nephrotoxicity induced by gentamicin prior to administration	96
Figure 4.10 (a)	OPLS-DA score plot fingerprinting models of pre-dose serum metabolites : A, histopathology fingerprinting model; B, serum creatinine fingerprinting model	98
Figure 4.10 (b)	OPLS-DA score plot fingerprinting models of pre-dose serum metabolites: C, BUN fingerprinting model; D, urine output fingerprinting model	99
Figure 4.10 (c)	OPLS-DA score plot fingerprinting models of pre-dose serum metabolites: E, combination of three criteria fingerprinting model; F, combination of all four AKI criteria fingerprinting model	100
Figure 4.11 (a)	OPLS-DA score plot of identification models of pre- dose serum metabolites A, histopathology identification model; B, serum creatinine identification model	104
Figure 4.11 (b)	OPLS-DA score plot of identification models of pre- dose serum metabolites: C, BUN identification model; D, urine output identification model	105
Figure 4.11 (c)	OPLS-DA score plot of identification models of pre- dose serum metabolites: E, combination of three criteria identification model; F, combination of all four AKI criteria identification model.	106
Figure 4.12 (a)	OPLS-DA score plot of fingerprinting models of pre- dose urine metabolites: A, histopathology fingerprinting model; B, serum creatinine fingerprinting model	110
Figure 4.12 (b)	OPLS-DA score plot of fingerprinting models of pre- dose urine metabolites: C, BUN fingerprinting model; D, urine output fingerprinting model	111

Figure 4.12 (c)	OPLS-DA score plot of fingerprinting models of pre- dose urine metabolites: E, combination of three criteria fingerprinting models; F, combination of all four AKI criteria fingerprinting models	112
Figure 4.13 (a)	OPLS-DA score plot of identification models of pre- dose urine metabolite: A, histopathology identification model; B, serum creatinine identification model	116
Figure 4.13 (b)	OPLS-DA score plot of identification models of pre- dose urine metabolite: C, BUN identification model; D, urine output identification model	117
Figure 4.13 (c)	OPLS-DA score plot of identification models of pre- dose urine metabolite: E, combination of three criteria identification model; F, combination of all four AKI criteria identification model	118
Figure 4.14	Box plot of AUC peaks at A, 1.26 B, 1.32 C 2.03 D, 3.27 in SD rat serum samples in urine output identification models of pre-dose serum	131
Figure 4.15	Box plot of AUC peaks at A, 1.26 B, 1.33 C, 3.26 D, 5.23 E, 5.39 in SD rat serum samples in combination with three AKI criteria, except the histopathology identification model of pre-dose serum	132
Figure 4.16 (a)	Box plot of AUC peaks at A, 2.57 B, 2.72 C, 3.01 D, 3.97 in SD rat urine samples in serum creatinine identification models of pre-dose urine	135
Figure 4.16 (b)	Box plot of AUC peaks at E, 5.80 F, 7.55 G, 7.64 H, 7.84 in SD rat urine samples in serum creatinine identification models of pre-dose urine	136
Figure 4.17	Box plot of AUC peaks at A, 3.27 in SD rat urine samples in BUN identification models of pre-dose urine	137

- Figure 4.18 (a) Box plot of AUC peaks at A, 2.45 B, 2.54 C, 2.57 D, 139 2.72 in SD rat urine samples in urine output identification models of pre-dose urine
- Figure 4.18 (b) Box plot of AUC peaks at E, 3.01 F, 3.27 G, 5.40 H, 140 5.80 in SD rat urine samples in urine output identification models of pre-dose urine
- Figure 4.19 Box plot of AUC peaks at A, 3.27 B, 3.97 C,7.55 D, 141 7.84 in SD rat urine samples in combination with three AKI criteria, except for the histopathology identification model of pre-dose urine
- Figure 4.20 Box plot of AUC peaks at A, 3.27 B, 3.90 in SD rat 142 urine samples in combination with four AKI criteria identification models of pre-dose urine
- Figure 4.21 (A) Key in the respective bin in BMRD website from 143 serum metabolites; (B) Potential metabolites selected from the list
- Figure 4.22 (C) Selected potential metabolites in BMRD were 144 double-checked by searching theHMDB website; (D) & (E) Availability of the metabolites in blood checked
- Figure 4.23 Matching of representative metabolite peaks with 144 Chenomx profiler
- Figure 4.24 (A) Key in the respective bin in BMRD website from 146 urine metabolites; (B) Potential metabolites selected from the list
- Figure 4.25(C) Selected potential metabolites in BMRD double-
checked by searching the HMDB website; (D) & (E)146Availability of the metabolites in urine checked
- Figure 4.26 Matching of representative metabolites peaks with 147 Chenomx profiler

- Figure 5.1 The combinatory metabolites according to models as 176 diagnostic biomarkers to predict gentamicin induced nephrotoxicity
- Figure 5.2 Summary of the metabolic pathways involving glucose-6-phosphate, 2-phosphoglycerate lactate and citrate for normal glycolisis pathways. The anaerobic glycolisis pathways were enhanced after insults by gentamicin to the kidney caused disruption in normal glycolisis pathways

LIST OF ABBREVIATIONS

AECUSM	Animal Ethics Committee USM
AKI	Acute kidney injury
AKIN	Acute kidney injury network
AMDI	Advanced Medical and Dental Institute
AMIX	Analysis of mixture
AP	Alkaline Phosphatase
AQ	Acquisition time
ASA PS	American Society of Anaesthesiologists Physical Status
ATM	Automatic tuning and matching
ATN	Acute tubular necrosis
AUC	Area under curve
AUROC	Area under receiver operating characteristics
B-BIOREFCODE	Bruker Biofluid Reference Compound Database
BBO	Broadband observe
BMRB	Biological Magnetic Resonance Data Bank

BUN	Blood urea nitrogen
Carb Hb	Carbamylated Hb
CHF	Congestive heart failure
CKD	Chronic kidney disease
COPD	Chronic obstructive pulmonary disease
COSY	Correlation spectroscopy
CPMG	Carr-Purcell-Meiboom-Gill
CRC	Clinical Research Centre
CrCl	Creatinine clearance
CRRT	Continuous renal replacement therapy
D1	Delay parameter
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
DS	Number of dummy scans
DSA	4,4-dimethyl-4-silapentane-1-ammonium trifluoroacetate
DSS	2,2-dimethyl-2-silapentane-5-sulfonic acid
eCrCl	Estimated creatinine clearance

EDTA	Ethylenediaminetetraacetic acid
eGFR	Estimated glomerular filtration rate
ESRD	End stage renal disease
FID	Free induction decay
FSGS	Focal segmental glomerulosclerosis
FT	Fourier transform
G6P	Glucose-6-phosphate
GC-MS	Gas chromatography-mass spectrometer
GC-TOF	Gas chromatography-time-of-flight
GFR	Glomerular filtration rate
r-GST	x-gluthathione-S-transferase
GGT	x-glutamyltransferase
GST	Gluthathione-S-transferase
H&E	Haematoxylin and eosin
HDL	High density lipoprotein
HMDB	Human Metabolome Database
HSQC	Heteronuclear single quantum coherence

ICU	Intensive care unit
IL-10	Interleukin-10
IL-18	Interleukin-18
IL-6	Interleukin-6
IL-8	Interleukin-8
IRRT	Intermittent renal replacement therapy
IQR	Interquartile range
IV	Intravenous
kDa	Kilo dalton
KDIGO	Kidney Disease Improving Global Outcomes
KIM-1	Kidney injury molecule-1
LC	Liquid chromatography
LCECA	Liquid chromatography electrochemical array
LC-MS	Liquid chromatography- mass spectrometer
LDH	Lactate dehydrogenase
LDL	Low density lipoprotein
MC	Mean centring

MCD	Minimal change disease
MDRD	Modification of Diet in Renal Disease
MIC	Minimum inhibitory concentration
MMP-9	Matrix metalloproteinase-9
МОН	Ministry of Health
MREC	Medical Research and Ethics committee
MS	Mass spectrometer
NADH	Nicotinamide adenine dinucleotide
NAG	N-acetyl-β- <i>v</i> -glucosamidase
NGAL	Neutrophil gelatinase-associated lipocalin
NHE3	Sodium-hydrogen exchanger isoform-3
NMR	Nuclear magnetic resonance
NOESY	Nuclear Overhauser effect spectroscopy
NS	Number of scan
NSAIDs	Nonsteroidal anti-inflammatory drugs
O1p	Centre of spectra
OD	Once daily

OPLS-DA	Orthogonal Partial Least Squares Discriminant Analysis
p1	Pulse length
PAR	Pareto
PAS	Periodic-acid Schiff
PCA	Principal Component analysis
PLS	Partial Least Squares
PLS-DA	Partial Least Squares Discriminant Analysis
Pro-ANP	Prohormone of atrial natriuretic peptide
Q^2	Goodness of prediction
QSAR	Quantitative Structure Activity relationship
R ²	Goodness of fit
RBP	Retinol-binding protein
RCF	Relative centrifugal force
RG	Receiver gain
RIFLE	Risk, injury, failure, loss and end stage kidney disease
ROA	Route of administration
ROS	Reactive oxygen species

RRT	Renal replacement therapy
SC	Subcutaneous
SCr	Serum creatinine
SD	Sprague Dawley
SD	Standard deviation
SIMCA	Soft Independent Modelling of Class Analogy software
SPSS	Statistical Package for the Social Sciences
SWH	Sweep width in hertz
TD	Time domain
TMNO	Trimethylamine-N-Oxide.
ТМОА	Trimethylamine-N-oxide
TMS	Tetramethylsilane
TSP	Sodium salt of 2,2,2,2,-tetradeutero-4,4- dimethyl-4- silapentanoic acid
UO	urine output
UV	Unit variance
Vd	Volume distribution
VIP	Variable Importance in Projection

α-GST α-	gluthathione-S-transferase
----------	----------------------------

π -GST π -gluthathione-S-transferase

LIST OF SYMBOLS

1D	One dimensional
2D	Two dimensional
Cl-	Chloride
dl	deciliter
D_2O	Deuterated water
H^+	Hydrogen
H ₂ O	Water
Hz	Hertz
K^+	Potassium
K ₂ HPO ₄	Dipotassium phosphate
m/z	mass over charge
mg	miligram
MHz	mega hertz
Na H ₂ PO ₄	Monosodium phoshate
Na ⁺	sodium

Na₂ HPO₄ Disodium phosphate

NaF	Sodium flouride
NaN ₃	Sodium azide
ppm	part per million
δ	Chemical shift

MERAMAL KENEFROTOKSIKAN ARUHAN GENTAMICIN MENGGUNAKAN PENDEKATAN PHARMACOMETABONOMIC

ABSTRAK

Gentamicin merupakan antibiotik yang digunakan secara meluas namun kesan kenefrotoksikan menjadi kebimbangan utama kerana ia disaring melalui buah pinggang untuk disingkirkan ke dalam urin. Walaubagaimanapun, diagnosis pada peringkat awal adalah sukar dan tiada metabolit yang boleh digunapakai sebagai penanda bio yang tersedia. Maka, dalam kajian ini, penyelidikan terhadap perubahan pola metabolit di dalam sampel serum dan urin sebelum administrasi gentamicin dijalankan melalui pendekatan 'Pharmacometabonomic' menggunakan NMR dan model untuk meramal kenefrotoksikan aruhan gentamicin dibangunkan seterusnya mengenalpasti metabolit berkaitan sebagai penanda bio. Matlamat kajian ini adalah menentukan potensi metabolit di dalam urin dan serum untuk meramal kenefrotoksikan aruhan gentamicin. Seterusnya, model pre-dos dibangunkan menggunakan sampel serum dan urin dalam tikus 'Sprague Dawley' (SD) yang di selidik menggunakan spektroskopi NMR bersama analisis statistik multivariate seperti analisis komponen utama (PCA) dan analisis orthogonal separa persegi terdiskriminasi (OPLS-DA); kemudian, satu set metabolit sebagai penanda bio dikenalpasti. Seterusnya, model yang telah dibangunkan disahkan di dalam sekumpulan tikus SD dan manusia. Gentamicin menghasilkan kesan toksik pada

sebahagian tikus SD, tetapi memberi kesan yang sedikit pada yang lain (kumpulan tidak toksik), seperti yang dinilai melalui keputusan kimia klinikal untuk kenefrotoksikan dan histologi. Selanjutnya, analisis statistik multivariat menunjukkan pemisahan yang signifikan di antara tikus SD yang toksik dengan tidak toksik dalam model yang telah dibangunkan. Model pre-dos serum dicirikan oleh satu set metabolit yang terdiri daripada laktat, trimethylemin-N-oksida, xilosa, deoxyinosine, 2-fosfogliserat and glukosa-6-fosfat. Manakala model pre-dos urin, dicirikan oleh satu set metabolit seperti asid 4-pyridoxic, 2-oxoglutarate, sitrat, betaina, asid hipurik, alantoin and urea. Metabolit ini dikesan melalui serum dan urin di dalam tikus SD yang mengalami nefrotoksik sebelum administrasi gentamicin dan berhubungkait dengan glikolisis anaerobik, kegagalan metabolism purina dan hiperuremia, kitaran asid trikarbosilik dan penekanan urin asid 4-piridoksik. Selain itu, model pre-dos serum dan urin juga telah divalidasi dalam tikus SD dan manusia namun keputusan kesensitifan, kejituan dan ketepatan model adalah rendah disebabkan saiz sampel yang kecil. Dapatan ini membuktikan model yang dihasilkan melalui pendekatan 'pharmacometabonomic' berpotensi untuk meramal kenefrotoksikan aruhan gentamicin sebelum pemberian dos gentamicin dan untuk pemahaman lanjut tentang ciri setiap metabolit yang dikenalpasti.

PREDICTING GENTAMICIN INDUCED NEPHROTOXICITY USING PHARMACOMETABONOMIC APPROACH

ABSTRACT

Gentamicin has been one of the most widely used antibiotics. However, its nephrotoxicity is a major concern as it is filtered through the kidneys for excretion in the form of urine. However, early diagnosis is difficult and no reliable metabolites as biomarkers are currently available. Thus, in this study, a pharmacometabonomic approach using nuclear magnetic resonance (NMR) spectroscopy to investigate the altered metabolic pattern in serum and urine samples prior to administration of gentamicin has been employed to develop a model to predict nephrotoxicity induced by gentamicin and to identify the metabolic biomarkers associated. The aim of this study is therefore to determine the potential of urine and serum metabolites in predicting gentamicin induced nephrotoxicity. Subsequent to this, a pre-dose model was developed using urine and serum samples in a number of Sprague Dawley (SD) rats under investigation using NMR spectroscopy. The finding were subjected, first to a multivariate statistical analysis, such as a principal components analysis and orthogonal partial least-squares discriminant analysis; then, a set of metabolites biomarkers was identified. Next, the developed models were validated in groups of SD rats and human subjects. Gentamicin produced toxic responses in some SD rats (toxic group), but had little effect in others (nontoxic group), as judged by their clinical chemistry for nephrotoxicity and histological results. The multivariate statistical analysis following this showed a significant separation between toxic and

non-toxic SD rats, from which the model was developed. The pre-dose serum was characterised by a set of metabolites of lactate, trimethylamine-N-oxide, xylose, deoxyinosine, 2-phosphoglycerate and glucose-6-phosphate. Meanwhile, in the predose urine of the SD rats, a set of metabolites was characterised by 4-pyridoxic acid, 2-oxoglutarate, citrate, betaine, hippuric acid, allantoin and urea of citrate, hippurate and glycine. These metabolites were detected from the serum and urine of SD rats with nephrotoxicity prior to gentamicin administration and associated with anaerobic glycolysis, impaired purine metabolism and hyperuricemia, tricarboxylic acid cycle, and depressed urinary 4-pyridoxic acid. Following this, the pre-serum and urine models were validated in SD rats and human subjects with poor sensitivity, specificity and accuracy due to a limitation in sample size. These findings demonstrate that the models produced by the pharmacometabonomic approach may be useful in predicting gentamicin induced nephrotoxicity prior doses and in arriving at a deeper understanding of the characteristics of each metabolite identified.

CHAPTER 1 – INTRODUCTION

Gentamicin is a well known antibiotic causing kidney injury in proximal tubules is a very crucial matter. Kidney is a vital organ in human body. These pair of bean shaped organ important function is in execratory of waste produced by human body. Other than that, kidney also important in regulation of blood pressure, minerals, red blood cells production and blood acid-base balance (Rylander et al., 2006). Importantly, drug induced nephrotoxicity contribute about 19-25% of kidney injury (Avent et al., 2011; Begg & Barclay, 1995). Nephrotoxicity is a term given for any poisonous or insult to the kidney. Many studies have been conducted worldwide to overcome and improve drug induced nephrotoxicity incidence in human. The techniques used for diagnosing and monitoring nephrotoxicity are evolving continually with new methods emerging including the discovery of biomarkers. This includes the new omics technology such as metabolomic, metabonomics and pharmacometabonomic. This new method had a promising hope in drug treatment which tailored to patients characteristics, thus it will improve efficacy and reduce the number and severity of nephrotoxicity. The objective of current study is to experiment, determine and evaluate the potential of urine and blood metabolite in predicting gentamicin induced nephrotoxicity. The role of this study to help the clinician personalized drug treatment according to patients need which it can maximize the benefits of the treatment and at the same time minimize the side effects.

1.1 Limitations of Current Practice

No proven methods are available to predict gentamicin induced nephrotoxicity prior to administration. Until the development of new biomarkers, detection of nephrotoxicity will only be discovered after the administration of gentamicin and/or until an overt change in renal function.

1.2 Statement of the Problem

To-date, no studies/methods have been proven to predict gentamicin induced nephrotoxicity in individuals prior to administration of the drug. In general, the detection of nephrotoxicity will only be perceived after administration of antibiotics/drugs and/or until an overt change in renal function is evident. In the current practice of hospitals, gentamicin is monitored using drug concentration in the blood after initiation of therapy. The present study, hopefully, can address current limitations. The objective of the current study is therefore to experiment, determine and evaluate the potential of human urine and serum metabolites in predicting antibiotic induced nephrotoxicity. It aims to help the clinician personalise drug treatment according to the patient's need in order to maximise the benefits of the treatment and at the same time minimise its side effects.

1.3 Study Hypothesis

This study hypothesises that gentamicin induced nephrotoxicity can be predicted using a pre-dose metabolic profile of urine and serum via pharmacometabonomic method using NMR.

1.4 Study Objectives

1.4.1 General Objective

The objectives of the current study are to determine the potential of urine and serum metabolites in predicting gentamicin induced nephrotoxicity.

1.4.2 Specific Objective(s)

The specific objectives of the study were to:

- Explore the potential of pre-dose urine and serum metabolites to discriminate between subjects with and without gentamicin toxicity
- 2. Observe changes in metabolites of the urine and serum of subjectsbefore and after treatment with gentamicin
- Associate the identified metabolites with the relevant clinical chemistry and histopathology of the kidney
- 4. Determine the putative metabolites that could predict gentamicin induced nephrotoxicity
- 5. Evaluate the possibility of using the same metabolites in predicting gentamicin induced nephrotoxicity in rats and humans.

CHAPTER 2 – LITERATURE REVIEW

2 The Kidney

The kidneys, also known as renals, are a vital organ in the human body. This pair of bean-shaped organs, each about the size of a clenched fist, and the adrenal glands sitting on top of them, release renin which affects blood pressure as well as sodium and water retention. The primary function of the kidneys is in the excretion of waste produced by the body and the removal of excessive fluid. In addition to that, the kidneys are also important in the regulation of blood pressure, minerals, production of red blood cells and maintaining the blood acid-base balance (Lattanzio & Kopyt, 2009).

The kidneys are found along the posterior muscular wall of the abdominal cavity. They lie behind the peritoneum that lines the abdominal cavity and are thus considered to be retroperitoneal organs. They are protected from external damage by the ribs and muscles of the back. Additionally, the adipose tissue known as perirenal fat which surrounds the kidneys acts as protective padding. If each kidney were bisected from top to bottom, the two major regions of the granular cortex and medulla could be visualised. Nephron is the functional unit of the kidneys which makes up one million functional units in the kidneys. Nephron has two main parts: one is the glomerulus. The other is Bowman's Capsule found in the renal corpuscle and renal tubule, consisting of a proximal convoluted tubule, loop of Henle, a distal convoluted tubule, a collecting tubule and a collecting duct (Figure 2.1) (Deshmukh & Wong, 2009).



Figure 2.1: Human Kidney Anatomy. Adapted from (Deshmukh & Wong, 2009)

Nephron is responsible for filtering the blood in the kidneys according to three main processes known as glomerular filtration, tubule secretion and tubule reabsorption. Blood enters the glomerulus via afferent arterioles under high pressure in the renal corpuscles, forcing substances to be sieved across the leaky endothelial-capsular membrane and podocyte into the nephron, passing Bowman's capsule into proximal convoluted tubules. Podocytes, which are special epithelial cells formed from the layer of the Bowman's capsule surrounding the capillaries of the glomerulus, work with the endothelium of the capillaries to separate the urine from blood passing through the glomerulus into the kidney tubules. An ultra-filtration process transports to the kidney tubules substances consisting of water, small proteins, salts (Na⁺, Cl⁻, K⁺, H⁺), glucose, nitrogenous waste products, such as urea, and other metabolic waste together with drug metabolite residues in the plasma and protein in the blood. Tubular reabsorption begins as soon as the filtrate enters the proximal convoluted tubule as the most active but selective reabsorption agent. This process involves near total reabsorption of organic nutrients, and the hormonally regulated reabsorption of water and ions. Sodium, bicarbonate, glucose, amino acids, vitamins, and most cations enable ion reabsorption via active transportation, whilst other substances such as water, lipid soluble solutes and urea reabsorbed via passive diffusion driven by chemical or concentration gradient. The proximal tubule is also an important site for the secretion of organic acids and bases such as bile salts, oxalate, urate and catecholamines (Christensen & Birn, 2001).

The descending limb of the loop of Henle is permeable to water and reabsorbs via osmosis. The ascending limb, however, though impermeable to water is permeable to electrolytes, which cause reabsorption of sodium, chloride, potassium through active transportation and passive diffusion of magnesium and calcium ions driven by electrochemical gradient and hydrogen ions secreted into the lumen. The first part of the distal tubule forms part of the juxtaglomerular complex that provides feedback control of the glomerular filtration rate and blood flow, and the continuing distal tubule segment absorbs most electrolytes in the same way as the ascending loop of Henle. On the other hand, the late distal tubule and cortical collecting tubule reabsorb sodium, potassium ions and water, while secreting hydrogen ions into the tubular lumen. The final urine output depends on the absorption of water and sodium in the collecting duct which depends on its antidiuresis hormoneal level (Deshmukh & Wong, 2009). Figure 2.2 summarize activities involve in kidney.



Figure 2.2: Secretion and reabsorption of various substances throughout the nephron in kidney. Adapted from (Deshmukh & Wong, 2009)

2.1 Acute Kidney Injury

2.1.1 Epidemiology

The true overall epidemiology of AKI is limited because of the different definitions of AKI and the variation in populations around the world. AKI is known in layman's terms as sudden impairment of kidney function. Although many definitions of AKI have being published, the Clinical Practice Guidelines produced from Kidney Disease Improving Global Outcome (KDIGO) and the UK Renal Association have agreed to define AKI as shown in Table 1.1 (Association, 2007; Khwaja, 2012b).

Table 2.1: Definition of AKI

AKI is defined by the criteria below:

- Increase in serum creatinine (SCr) by x0.3 mg/dl (x 26.5 µmol/l) within 48 hours; or
- Increase in SCr to x1.5 times baseline, which is known or presumed to have occurred within the prior 7 days; or
- Urine volume 0.5 ml/kg/h over 6 hours

The cause of AKI can be divided into three categories as follows (Choudhury & Ahmed, 2006); prerenal, intrinsic and postrenal AKI. Prerenal AKI resulting from decreased renal perfusion, often as a consequence of cardiac failure, volume depletion and low blood pressure, such as after severe hemorrhage sepsis burns. Intrinsic AKI resulting from abnormalities within the kidney itself, which can be further divided into sub categories (a) AKI caused by glomerularnephritis, (b) AKI caused by tubular necrosis, which might be contributed by severe renal ischemia or toxins and medication. Meanwhile, postrenal AKI, meaning the obstruction of the

urinary system any where from the calices to bladder outflow. Most of the obstructions are caused by abnormalities of the lower urinary tract outside the kidney such as kidney bladder outlet obstruction and kidney stones in both ureters. Nevertheless, some studies have documented the frequent occerrence of AKI in intensive care unit (ICU) cases which possibly affect about two-thirds of such patients (Dennen et al., 2010; Lameire et al., 2008; Parazella, 2010; Uchino, 2006). It is therefore crucial to know the incidence, etiology and clinical features of AKI to promote preventive strategies and to implement adequate resources for the management of AKI. The mortality rate of patients having AKI is approximately 50% and the figure rises to 70% of patients treated in the ICU (Liano et al., 1996; Lins et al., 2000; Parazella, 2010).

2.1.2 Drug Induced Nephrotoxicity

The incidence of drug induced nephrotoxicity is difficult to determine but about 18.3% cases of acute tubular necrosis (ATN) are caused by medication (Choudhury & Ahmed, 1997). It is notworthy that the reported incidence of nephrotoxicity due to administration of antibiotics is 36% (Choudhury & Ahmed, 2006). Different drugs may cause injury at different sites in the kidneys and such injury caused by drugs can be categorised as tubular damage, glomerular damage, interstitial damage and vascular damage (Choudhury & Ahmed, 2006). The tubules are more susceptible to toxic injury for the following reasons. First, high concentrations of filtered toxicants are present in the tubular fluid due to their function in the reabsorption of solutes and

water. Second, tubular epithelial cells have a large number of transporters, resulting in high intracellular concentrations of toxicants (Choudhury & Ahmed, 1997, 2006; Naughton, 2008). Third, tubular epithelial cells have high energy requirements for supporting metabolism as well as for transporting solutes. Tubular injury can be the result of direct toxicity in the tubular epithelium, or obstructive insult or inflammation. The most frequent drugs leading to tubular injury are aminoglycosides, radiocontrast media, cisplatin, amphothericin B, pentamidine, tacrolimus, cephaloridine, and many more. In glomerular damage, the drugs involved include lithium, pamidronate, sirolimus, NSAIDs and interferons, which lead to conditions known as minimal change disease (MCD) and Focal segmental glomerulosclerosis (FSGS). Most common forms of interstitial damage are acute and happen as a result of allergic reactions that elicit a strong inflammatory response. Antibiotics such as penicillins, cephalosporins, sulfanamides, rifampicin, nonsteroidal anti-inflammatory drugs (NSAIDs), phenytoin and thiazide diuretic have been associated with interstitial damage. The less common form of renal injury caused by drugs are vascular damage, associated with drugs such as propylthiouracil, hydralazine, methimazole, sulfasalazine, phenythoin and minocycline (Begg & Barclay, 1995; Choudhury & Ahmed, 1997, 2006; Parazella, 2010). Other than that, acute and chronic risk factors as listed in Table 2.1 might increase the risk of drug induced nephrotoxicity.

Chronic risk factors	Acute risk factors	
Age > 65 years	Sepsis/infection	
Chronic kidney disease	Volume Depletion	
Diabetes mellitus	Acute decompensated heart failure	
Malignancy	Hypotension	
Cardiovascular disease	Complex & major surgery	
Peripheral vascular disease	Trauma	
-	Mechanical ventilation	

Table 2.2: Risk factors for drug induced nephrotoxicity (Parazella, 2010)

2.2 Aminoglycosides

Aminoglycoside antibiotics, which are derived from Streptomyces and Micromonospora, have very potent bactericidal activity (Avent et al., 2011). The basic chemical structure required for both potency and the spectrum of antimicrobial activity of aminoglycosides is that of one or several aminated sugars joined in glycosidic linkages to a dibasic cyclitol Figure 2.3 (Mingeot-leclercq & Tulkens, 1999). Currently, there are nine (9) aminoglycosides available in the market: gentamicin, tobramycin, amikacin, streptomycin, neomycin, kanamycin, paromomycin, netilmicin and spectinomycin as listed in Table 2.3 and the chemical structure as shown in Figure 2.3.

Gentamicin, tobramycin and amikacin are the most widely prescribed by physicians (Radigan et al., 2010). Most aminoglycosides are admistered via the parentral route as they are not absorbed through the stomach (Radigan et al., 2010; Tulkens et al., 1999). A significant clinical toxicities, such as nephrotoxicity and ototoxicity, can result from aminoglycosides. Clinical studies have shown that about 5-25% of adult patients receiving aminoglycosides experienced drug induced toxicity (Avent et al., 2011; Choudhury & Ahmed, 1997; Guo & Nzerue, 2002; Hock & Anderson, 1995; Meyer, 1986) (Pannu & Nadim, 2008) (Radigan et al., 2010).

2.2.1 Gentamicin

Gentamicin is one of the most widely prescribed drugs in the aminoglycoside group. Gentamicin is effective for serious infections caused by Gram-negative organisms (Avent et al., 2011; Leong et al., 2006; Selby et al., 2009). Rapid bactericidal activity and comparatively low incidence of resistance in most community- and hospitalassociated gram-negative pathogens make it beneficial for empirical management when rapid control of a serious infection is required (Jana & Deb, 2006; Shakil et al., 2008).

Types of	Year of	Origin
Aminoglycosides	discovery	
Streptomycin	1944	Streptomyces griseus
Neomycin	1949	Streptomyces fradiae
Kanamycin	1957	Streptomyces kanamyceticus
Gentamicin	1963	Actinomycete Micromonospora
		purpurea
Tobramycin	1967	Streptomyces tenebrarius
Amikacin	1972	Semisynthetic derivative of
		kanamycin
Netilmicin	1976	Semisynthetic derivative of sisomisin
		from micromonospora sp.

Table 2.3: Types of aminoglycosides (Begg & Barclay, 1995)



Figure 2.3: Chemical structure of Aminoglycosides antibiotics. A streptomycin, B neomycin, C kanamycin D, Gentamicin, E Tobramycin, F Amikacin, G Netilmicin

2.2.1.1 Mechanism of Action

The basic mechanism of gentamicin in the destruction of bacteria is by impairing bacterial protein synthesis through binding to prokaryotic ribosomes. Gentamicin is diffused through the outer membrane of gram-negative bacteria through disruptions of Mg²⁺ bridges between adjacent lipopolysaccharide molecules (Barclay et al., 1999; Jana & Deb, 2006). Gentamicin is transported into cytoplasmic by active transportation since the size of the gentamicin particle is too large for channel transportation. However, this step is rate-limiting and can be blocked by calcium and magnesium ions, hyperosmolarity, low pH and anaerobic conditions (Barclay et al., 1999; Begg & Barclay, 1995). Through energy dependent transportation, gentamicin binds to the 30S subunit of ribosomes in cytosol and disturbs the elongation of the chain by impairing the reading process which controls translational accuracy (Tulkens et al., 1999). Moreover, gentamicin causes a decrease in translational accuracy and inhibits translocation of the ribosome of a bacterial cell (Yoshizawa et al., 1998).

2.2.1.2 Clinical Use

As one of the most commonly prescribed aminoglycosides, gentamicin can be used alone or in combination with other antibiotics for preventive, empirical or directed therapy. In preventive therapy, only a single dose is sufficient generally before surgical procedures. For empirical therapy, the regime should not exceed 48 hours, and the monitoring of plasma concentration is not required. However, in the case of directed therapy with gentamicin, monitoring of the plasma concentration is necessary if the treatment is prolonged for more than 48 hours (Avent et al., 2011; Moulds & Jeyasingham, 2010). Directed therapy is limited for specific indications as listed in Table 2.4 (*National Antibiotic Guidelines 2014*, 2014). The extended interval gentamicin doses or once daily dose have shown more benefits and are preferable to traditional multiple daily dosages (Abdel-Bari et al., 2011; Barclay et al., 1999; Lacy et al., 1998). A once daily dosage of gentamicin provides a higher concentration in the blood and reduces the toxicity effects due to a 'gentamicin free' period created during the interval time of administration (Abdel-Bari et al., 2011; Barclay et al., 1999; Lacy et al., 1998; Nicolau et al., 1995).

Preventive therapy	Empirical therapy	Directed therapy
Single dose only	Duration < 48 hours	Duration > 48 hours
Alone or combination	combination	combination
 Surgical Procedure Hysterectomy/ laparoscopy Orthopaedic surgery Urology surgery Cardiac surgery Appendicitis surgery 	Infective endocarditisChorioammionitis	 Soft tissues infection Muscular, skeletal & soft tissue trauma Compound trauma Urology infection Kidney abscess Acute prostatitis Acute uncomplicated pyelonephritis

Table 2.4: Indications for parenteral gentamicin

2.2.1.3 Pharmacokinetics and Pharmacodynamics

Gentamicin is poorly absorbed through oral adminstration and it has a half life of 30-60 minutes post administration via intramuscular and intravenous means. It is a nonprotein bound drug with a volume distribution (Vd) approximating the volume of extracellular fluid of 20-35% of body weight (Niemiec et al., 1987; Triginer et al., 1990). However, Vd will expand when there is an increase in volume of extracellular fluid as in patients with ascites, edema, or other enlarged third space (Robertson, 2005). Once in the systemic circulation, their poor lipid solubility prevent the gentamicin being absorbed into most tissues and the majority of the drug pass through glomerular filtration and is renally excreated unchanged form. However a significant propotion typically 15% accumulate within the proximal tubule epitelial cells. A careful dose adjustment of gentamicin is therefore essential for patients with existing renal problems as it is excreted completely by glomerular filtration (Roberts et al., 2012). Gentamicin has concentration-dependent antibacterial activity which means, that the rate of killing the bacteria increases with the increased of gentamicin concentration (Lacy et al., 1998). Studies have shown that, with a higher ratio of gentamicin concentration to minimal inhibitory concentration (MIC) of 8-10:1, the bactericidal effects of gentamicin will be maximised (Moore et al., 1984a, 1984b; Zelenitsky et al., 2003). On the downside, gentamicin has a narrow therapeutic index, which can lead to toxicity if the drug is prescribed without proper monitoring.

2.2.1.4 Nephrotoxicity

Even though gentamicin is well known for its very effective antibiotic properties and low resistance, its usage is limited because of the risk of nephrotoxicity. The clinical manifestation of gentamicin induced nephrotoxicity is nonoliguric renal failure, with a slow rise in serum creatinine and hypoosmolar urinary output, which develop after several days of treatment (Tulkens et al., 1999). It is well known that aminoglycosides are excreted unchanged by glomerular filtration and reabsorbed into the proximal tubule via endocytosis mechanism mediated by megalin, a 600 kDa multiligand receptor located on apical brush border of the proximal tubule (Choudhury & Ahmed, 2006; Tulkens et al., 1999; Zhai et al., 2000). Megalin, previously known as the Heymann nephritis autoantigen, is a large glycoprotein 330 receptor and member of the low density lipoprotein (LDL) receptor gene family. It is is located mainly in the renal proximal tubule epithelium and labyrinth epithelium of the ears (Farquhar et al., 1995; Zhai et al., 2000) as well as on the retinal epithelium and yolk sac (Zhai et al., 2000). There, gentamicin accumulates within the lysosomes compartment to inhibit lysosomal enzymes, resulting in impaired function and acute tubular necrosis (Radigan et al., 2010). The histopathology of the kidneys has shown severe nephropathy mainly in proximal convoluted tubules nine (9) days after the patients were put on gentamicin treatment (Lenz et al., 2005).

2.3 Current Practice in Diagnosis and Management of Nephrotoxicity caused by Gentamicin

2.3.1 Current Practice to Detect Nephrotoxicity caused by Gentamicin

In ancient Greek medical history, AKI was diagnosed, once there was evidence of a reduction in urine volume (Waikar et al., 2008). As time passed, numerous studies have been conducted worldwide to improve the tools used in the diagnosis and management of AKI. The current indicator used in clinical settings to diagnose AKI is the glomerular filtration rate (GFR), supported by a comprehensive patient history, and physical examination. Patient data, such as vital signs, input/output chart, daily weight, past and current laboratory data (example, serum creatinine (SCr) and blood urea nitrogen), fluid balance and medications are a crucial and classic approach in

helping the diagnosis of AKI and its causes. Data gathered from the patient will be used to diagnose AKI based on the presence of increased serum creatinine and/or blood urea nitrogen (BUN) levels and/or a decreased urine output (Akcay et al., 2010). SCr and BUN have been widely used as biomarkers to diagnose AKI because both are the major by-products of protein metabolism and are normally filtered by the glomerulus. Therefore, any increase in serum concentrations of creatinine and BUN is an indicator of decreased GFR. The baseline renal function can also be estimated using the Modification of Diet in Renal Disease (MDRD) formula or the Cockcroft-Gault formula in adults and the Schwartz formula for children (Table 2.5) (Naughton, 2008; Palevsky et al., 2013). These formulae are useful in categorising the type of AKI for dosage adjustment of drugs when the creatinine clearance (CrCl) falls below 50 ml/min (Naughton, 2008; Palevsky et al., 2013). Hence, the most upto-date development of tools to diagnose and classify AKI are based on creatinine increase and decrease in the estimated glomerular filtration rate (GFR) or urine output. These are used as biomarkers and are being utilised in the RIFLE criteria (Rrenal risk, I-injury, F-failure, L-loss of kidney function, E-end stage kidney disease [ESKD]) and AKIN (Acute Kidney Injury Network) criteria to differentiate the AKI. On the other hand, the detection of nephrotoxicity induced by gentamicin, apart from the above method, comes from therapeutic drug monitoring, linear regression analysis, using population methods or Bayesian estimation procedures (Avent et al., 2011). All the procedures listed can detect nephrotoxicity induced by gentamicin only when there is clear evidence of renal damage. Furthermore, a case reported by Bennett (Bennett et al., 1979) and Edwards (Edwards et al., 1976) have shown that therapeutic drug monitoring of aminoglycosides was not accurate in estimating the amount of aminoglycosides in the body as a certain level of gentamicin is still detected in the kidney and urine but not in the blood serum despite the discontinuation of gentamicin (Edwards et al., 1976; Gilbert et al., 1979). This is because, half –life of gentamicinin in kidney tissue is 109 hours as compared with serum half-life of 35 minutes (Gilbert et al., 1979). The prolonged accumulation of gentamicin in the kidney might increased the risk for nephrotoxicity.

2.3.2 Current Management of Gentamicin Induced Nephrotoxicity

The management and treatment plans of patients with AKI are varied and depend on etiologic factors. Generally, patients at high risk of developing AKI, such as elderly patients with concomitant chronic diseases, should be monitored closely for AKI. Specifically, guarding against gentamicin induced nephrotoxicity would require the discontinuation of gentamicin after the assessment of benefit-risk of the drug. In addition, laboratory parameters should be obtained vigilantly together with maintenance of optimum hemodynamics, and close surveillance for complications of renal dysfunction (eg: acidosis, electrolyte abnormalities) and controlling sepsis (Akcay et al., 2010; Guo & Nzerue, 2002; Kellum & Lameire, 2013; Lattanzio & Kopyt, 2009).

Table 2.5: Overview table of equation to estimate kidney function

Formula	Estimation Formula	Rationale
CKD-epi eGFR	141 x min (SCr/κ,1) ^α x max (SCr/κ,1) ^{-1.209} x 0.993 ^{Age} x 1.018 [if	
	female] x 1.159 [if black]	
	$\kappa = 0.7$ (females) & 0.9 (males)	
	α = -0.329 (females) & -0.411 [males]	
	min indicates the minimum of SCr/ κ or 1, and max indicates the	
	maximum of SCr/k or 1.	
MDRD eGFR(Stevens,	eGFR = 175x standardised Scr (mg/dl) ^{-1.154} x age (years) ^{-0.203}	To assess renal function stage chronic
Coresh, Greene, &	x (0.742 if patient is female) x (1.210 if patient is black)	kidney disease (Levey et al., 1999; Levey et
Levey, 2006)	Where GFR is expressed as ml/min/1.73m ² of body surface area.	al., 2003)
Cockroft and Gault	eCrCl = ([140- age (years)] x body weight [kg]) / Scr [µmol/L] x	To adjust drug dosage for renal function in
Creatinine	constant	adults
Clearance(Cockcroft &	Where the constant is 1.23 in male and 1.04 in females. Expressed in	
Gault, 1976)	ml/min.	
Schwartz (Schwartz,	eCrCl = (length [cm] x k) / serum creatinine (mg/dL)	To adjust drug dosage for renal function in
Haycock, Edelmann, &	k =	children
Spitzer, 1976)	0.45: infants 1 to 52 weeks of age	
	0.55: children one to 13 years of age	
	0.70: males 14 to 17 years of age	
	0.55: females 14 to 17 years of age	

eCrCl = estimated creatinine clearance; eGFR = estimated glomerular filtration rate; MDRD = Modification of diet in Renal Disease

2.3.3 Update on Kidney Injury Biomarkers

New markers can now detect kidney injury prior to the rise of SCr and clear evidence of kidney damage. SCr and BUN are well known markers used in the detection of AKI but these biomarkers are considered poor markers of renal dysfunction because SCr and BUN are insensitive and not specific. Scr concentration is greatly influenced by abundant non-renal features (such as body weight, race, age, gender, total body volume, drugs, muscle metabolism, and protein intake) (Coca et al., 2008; Group., 2012; Khwaja, 2012a). Furthermore, acute changes in SCr behind the stage of kidney damage may be delayed; hence using Scr alone to estimate kidney dysfunction may lead to undermining the early detection of kidney injury, thus fail in early diagnosis of sepsis-induced AKI, all of which could also result in further incorrect assessment of treatment efficacy (Coca et al., 2008; Han et al., 2008; Khwaja, 2012b). Therefore, the need for more sensitive and specific biomarkers arises with more research to explore new biomarkers to diagnose AKI earlier. A good diagnostic and monitoring marker should be affordable, accurate, stable, specific and sensitive to the disease/condition under investigation. Other than that, the characteristics needed to formulate biomarkers for acute kidney injury (AKI) are listed in Table 2.6. Several new biomarkers have been proposed and are in various stages of development and validation. Predictor of AKI as listed in Table 2.7 consist of 21 unique biomarkers in both serum and urine from 31 studies (Coca et al., 2008). The biomarkers might be characterized according to their specific performance, for instance: differential diagnosis in established AKI, early detection of AKI and prognosis of AKI.

Table 2.6: Characteristics of ideal biomarker for acute kidney injury

- Non-invasive
- Easily detectable in accessible samples like serum or urine
- Highly sensitive and specific for AKI, also in the presence of concomitant injury involving other organs
- Rapidly and reliably measurable
- Capable of early detection of AKI
- Able to give insight into etiology, nature and duration of insult
- A marker of injury in addition to marker of function
- Predictor of AKI severity and reversibility
- Helpful in monitoring course of and response to interventions
- Useful as surrogate end-point for clinical interventional studies
- Unaffected by other biological variables
- Inexpensive

Table 2.7: New biomarkers to predict AKI

Differential diagnosis in established AKI	Early detection	Prognosis
established AKI • Serum \checkmark Cystatin C \checkmark Carb Hb \checkmark NGAL • Urine \checkmark NGAL • GST \checkmark NAG \checkmark IL-18 \checkmark α -1 microglubolin \checkmark NHE3 \checkmark KIM-1 \checkmark MMD 0	• Serum \checkmark Cystatin C \checkmark Pro-ANP \checkmark NGAL \checkmark Neutrophil-CD11b • Urine \checkmark NGAL \checkmark IL-18 \checkmark KIM-1 \checkmark GST \checkmark π -GST \checkmark π -GST \checkmark $($ CST)	 Serum <i>RRT</i>: Cystatin C, NGAL <i>Death</i>: IL-6, IL-8, IL-10 Urine <i>RRT</i>: NGAL, Cystatin C, α-1 microglubolin, RBP, β-1 microglubolin, NAG, α-GST, CCT L DU KIM
• MMP-9	$\begin{array}{c} \checkmark & \text{AP} \\ \checkmark & \text{AP} \\ \checkmark & \text{NAG} \\ \checkmark & \text{LDH} \\ \checkmark & \text{MMP-9} \end{array}$	1 ✓ <i>Death</i> : NGAL, IL- 18, NAG, KIM-1

2.3.3.1 Serum Biomarkers

The most widely discussed biomarkers are cystatin C, which has proven to be a good marker in the detection of AKI at 24 and 48 hours. It has also performed extremely well in identifying established AKI as a marker of glomerular filtration and for predicting the prognosis for RRT. On the other hand, neutrophil gelatinase-associated lipocalin (NGAL) is a good marker for renal tubular inflammation but its sensitivity towards early diagnosis of AKI is low; neither is it able to discriminate between subjects who do not require RRT or who subsequently require RRT (Coca et al., 2008).

2.3.3.2 Urine Biomarkers

The elevation of urinary IL-18, kidney injury molecule-1 (KIM-1) and N-acetyl-β-Dglucosaminidase (NAG) are also predictors of AKI caused by acute tubular necrosis. The same study revealed that, the combinination of KIM-1, NAG and metalloproteinase-9 (MMP-9) yielded perfect results for diagnosing AKI. Sodiumhydrogen exchanger form-3 (NHE-3) is also a good marker in detecting proximal tubule injury. Urinary IL-18 and urinary NGAL are good metabolites as an early predictive biomarker of AKI; even though urinary IL-18 has low sensitivity, it has a high specificity and rarely shows false positive elevation in AKI. In the case of riskstratified patients who are undergoing critical dialysis, urinary KIM-1, NAG and IL-18 are the most beneficial markers compared with the others. There are also studies showing metabolite changes in the urine when induced by gentamicin in rats. The metabolites identified are serine, threeonine, glutamine, agmatine and phosphate while 5-methyltetrahydrofolate is a putative biomarker in the kidney tissue for nephrotoxicity induced by gentamicin (Boudonck, Mitchell, et al., 2009; Boudonck, Rose, et al., 2009).

2.4 The Potential Role of Pharmacometabonomics in Nephrotoxicity

Pharmacometabonomics is a new method arising from -omics and has a very promising potential in detecting toxicity. As defined by Clayton et el, pharmacometabonomics is the prediction of the outcome of a drug or xenobiotic intervention in an individual based on mathematical models with a pre-intervention metabolites signature (Clayton et al., 2006). The advantages of these new methods are the ability to personalise drug treatments according to each patient's needs; this can therefore maximise treatment benefits and minimise side effects. Ideally, each patient will have the right medicine at the right dosage at the right time. With this new technology, urine was used to predict the effectiveness of the treatment, which is more convenient and non-invasive for the monitoring of drugs. Previous studies have shown the application of pharmacometabonomics as a predictor of toxicity in drug management (Table 1.8).

Therefore. aimed of this study, to investigate the ability of the pharmacometabonomics technique to predict aminoglycoside induced nephrotoxicity in human. Gentamicin will be used because it is commonly prescribed and the most toxic of aminoglycosides compared with others (Paul, 1986). As more people are now looking more into personalised medicine or treatment, scientists globally are endeavouring to identify the complexities of personalised medicine.