EVALUATION OF PDMS-BASED UV-CROSSLINKED HYDROGELS PROPERTIES FOR TISSUE ENGINEERING APPLICATIONS

LIM KAR WAI

UNIVERSITI SAINS MALAYSIA

2018

EVALUATION OF PDMS-BASED UV-CROSSLINKED HYDROGELS PROPERTIES FOR TISSUE ENGINEERING APPLICATIONS

by

LIM KAR WAI

Thesis submitted in the fulfilment of the requirements for the degree of Master of Science

September 2018

ACKNOWLEDGEMENT

My sincerest gratitude goes to my supervisor, Dr. Zuratul Ain Abdul Hamid for her guidance and support throughout the research period. Her extensive knowledge has been a very big help for me to do my research and to write this thesis. Not only that, her kindness and understanding in which have given me courage to continue this research although there were obstacles along the way.

I would like to acknowledge to my scholarship sponsor MyBrain 15 for the financial support during my master program. This work was also supported by Grant from FRGS (National) and RUI (Internal). Million thanks to technical and administrative staffs who has kindly guided me to do this research right from the beginning until the end, especially Mr. Faizal, Mr. Sharil, Mr. Farid, Mr. Kemuridan, Mr. Abdul Rahim and Madam Lai Fan Choong. Many thanks to Universiti Sains Malaysia (USM) and SMMRE for research facilities.

I must express my very profound gratitude to my beloved parents and my family for encouraging and supporting me throughout my years of study and through the process of researching and writing this thesis.

Not forget to mention, thanks to my colleagues and fellow postgraduate students who has given me support throughout the year. Thank you for all your kindness, friendship and moral support during my research study. I will cherish every bittersweet memory during my stay here in USM.

TABLE OF CONTENTS

ACK	NOWLEI	DGEMENT			ii
TABLE OF CONTENTS ii					iii
LIST	OF TAB	LES			vii
LIST	OF FIGU	JRES			viii
LIST	OF ABB	REVIATIO	NS		xii
LIST	OF SYM	BOLS			XV
ABST	FRAK				xvi
ABST	FRACT				xviii
СНА	PTER ON	JE: INTRO	DUCTION		
11	Research	h backgroun	d		1
1.2	Problem	statement			3
1.3	Objectiv	res of the stu	dv		5
1.4	Research	n scope	2		5
		•			
CHA	CHAPTER TWO: LITERATURE REVIEW				
2.1	Tissue engineering			7	
2.2	Scaffold	S			9
	2.2.1	Key requi	rements for des	igning scaffolds	9
	2.2.2	Classifica	tions of scaffold	ds	11
2.3	Hydroge	els			13
	2.3.1	Definition	n of hydrogels		13
	2.3.2	Classifica	tions of hydrog	els	14
		2.3.2 (a)	Classification	based on their sources	14
		2.3.2 (b)	Classification	based on polymeric composition	20
		2.3.2 (c)	Classification	based on physical appearances	20
	2.3.3	Crosslink	ing method of h	ydrogels	24
		2.3.3 (a)	Physical cross	linking methods	24
		2.3.3 (b)	Chemical cross	sslinking methods	24
			2.3.3 (b) (i)	Chain-growth	25
				polymerization/crosslinking	
			2.3.3 (b) (ii)	Step-growth	26
				polymerization/crosslinking	
2.4	Photopo	lymerizatior	n/crosslinking re	eaction	26

2.3 PDM	S-based hydrogels		30
2.5.1	Polydime	lydimethylsiloxane (PDMS)	
2.5.2	Fabricatio	on of PDMS-based hydrogels	32
2.5.3	PDMS-Pl	EG hydrogels	33
2.5.4	Potential	application of PDMS-based hydrogels	34
	2.5.4 (a)	Contact lenses	34
	2.5.4 (b)	Drug delivery	34
	2.5.4 (c)	Tissue engineering	35
2.6 Comp	patible PDMS/I	PEG hydrogels	35
2.7 Sumr	nary		38
CHAPTER	THREE: EXP	ERIMENTAL	
3.1 Mate	rials		40
3.2 Synth	esis of PDMS	precursors	41
3.2.1	PDMS-Si	H precursor	41
3.2.2	PDMS-M	A precursor	43
3.3 Curin	g profile of PD	rofile of PDMS-MA precursors	
3.4 Prepa	ration of PDM	on of PDMS-PEG hydrogels	
3.5 Mate	rials characteriz	erization	
3.5.1	PDMS pr	ecursors	45
	3.5.1 (a)	Fourier transform infrared spectroscopy (FTIR)	45
	3.5.1 (b)	Gel permeation chromatography (GPC)	46
	3.5.1 (c)	Nuclear magnetic resonance (NMR) spectroscopy	46
	3.5.1 (d)	X-ray fluorescence (XRF)	46
3.5.2	PDMS-Pl	EG hydrogels	47
	3.5.2 (a)	Gel content	47
	3.5.2 (b)	Swelling test	47
	3.5.2 (c)	Wetting properties	47
	3.5.2 (d)	Differential scanning calorimetry (DSC)	48
	3.5.2 (e)	Dynamic mechanical analysis (DMA)	48
	3.5.2 (f)	Texture profile analysis (TPA)	49
	3.5.2 (g)	Atomic force microscopy (AFM)	49
	3.5.2 (h)	Scanning electron microscopy (SEM)	49
	3.5.2 (i)	Optical microscopy	50
	3.5.2 (j)	Ultraviolet-visible (UV-vis) spectroscopy	50
	3.5.2 (k)	In-vitro cytotoxicity test	51

CHAPTER FOUR: RESULTS AND DISCUSSION

REFE	RENCES				100
5.2	Recomme	endation			99
5.1	Conclusio	on			98
CHAI	PTER FIV	'E: CONCI	LUSION AND	RECOMMENDATION	
	4.3.7	In-vitro cy	ytotoxicity test		94
	4.3.6	Protein ad	lsorption		93
		4.3.5 (b)	DMA		91
		4.3.5 (a)	Texture profil	e analysis (TPA)	87
	4.3.5	Mechanic	Mechanical testing		87
	4.3.4	Wettability			85
	4.3.3	3 Swelling properties		83	
		4.3.2 (d)	SEM		81
				inclusion of AMA	
			4.3.2 (c) (ii)	Degree of compatibility after	78
				inclusion of AMA	
			4.3.2 (c) (i)	Degree of compatibility before	74
		4.3.2 (c)	AFM analysis		74
		4.3.2 (b)	DSC analysis		71
		4.3.2 (a)	Visual inspect	ion	69
	4.3.2	Compatib	ility of PDMS-	PEG hydrogels	69
	4.3.1	Chemical	structures of Pl	DMS-PEG hydrogels	68
4.3	Character	rization of H	PDMS-PEG hyd	lrogels	68
	4.2.3	Gel conte	nts		66
	4.2.2	Degree of	conversion		64
	4.2.1	Photocros	slinking mecha	nism of PDMS-MA	61
4.2	Curing st	udy of PDN	IS-MA precurs	ors	61
	4.1.3	Molecular	weight		59
	4.1.2	Purity			58
		4.1.1 (b)	PDMS-MA		55
		4.1.1 (a)	PDMS-SiH		53
	4.1.1	Chemical	structures		53
4.1	Character	rization of H	PDMS precurso	rs	53

APPENDICES

Appendix A: Synthesis: stochiometric calculation Appendix B: ¹H-NMR data of all PDMS precursors (in detail) Appendix C: ¹H-NMR spectrum of PDMS-MA (H) precursor (in detail) Appendix D: ¹H-NMR spectrum of PDMS-MA (M) precursor (in detail) Appendix E: ¹H-NMR spectrum of PDMS-MA (L) precursor (in detail) Appendix F: New r.i of the MA signals in the PDMS-MA polymers Appendix G: ¹H-NMR of allyl methacrylate (AMA) Appendix H: Determination of M_n by ¹H-NMR end group analysis Appendix I: The topographical image of PDMS(1K)-PEG hydrogels Appendix J: UV-Vis absorption spectra of BSA

Appendix K: BSA calibration curve

LIST OF PUBLICATIONS

LIST OF TABLES

Table 2.1	Physical properties of PEG (Olabisi & Adewale, 2016)	17
Table 2.2	Major differences between the heterogeneous polymerization techniques (Arshady, 1992)	23
Table 2.3	Physical properties of PDMS (Kuo, 1999)	31
Table 3.1	Materials used in this project	40
Table 3.2	The formulation for the synthesis of PDMS-SiH precursors with different Mn for the CROP process	42
Table 3.3	The formulation for the synthesis of PDMSMA precursors with different Mn from hydrosilylation process	43
Table 3.4	Parameters used for this project	44
Table 4.1	Chemical compositions of the activated charcoals	59
Table 4.2	Comparison of Mn, Mw, and PDI	60
Table 4.3	Effects of Mn and AMA reactive diluent on the gel content	67
Table 4.4	Tg of PDMS (1K)-PEG hydrogels	72
Table 4.5	Tg of PDMS (1K)-PEG hydrogels	73
Table 4.6	Hansen solubility parameter	81
Table 4.7	Elemental analysis of the selected hydrogels	81

LIST OF FIGURES

		Page
Figure 2.1	Tissue engineering triad	8
Figure 2.2	Chemical structure of PHEMA	16
Figure 2.3	Chemical structure of PEG	18
Figure 2.4	The differences between type I and type II photoinitiators (Adapted from Nguyen & West, 2002, in Figure 1 of pp. 4308)	28
Figure 2.5	Mechanism of radical dissociation from Irgacure® 2959, Irgacure® 651 and Irgacure® 184 (Adapted from Mironi-Harpaz et. al., 2012, in Figure 1 of pp. 1840)	30
Figure 2.6	Chemical structures of PDMS	32
Figure 2.7	The PDMS and PEG used in the work of Cui et. al. (2012)	34
Figure 3.1	A simplified flow chart of the project	39
Figure 3.2	General schemes for the synthesis of PDMS-MA precursors	42
Figure 4.1	D ₄ and PDMS-SiH spectra	53
Figure 4.2	¹ H-NMR spectra of the PDMS-SiH precursors. Note: the siloxyl methyl groups on the terminal ends would experience a slightly different magnetic environment from the rest (siloxyl methyl groups in n repeating unit), due to neighboring H atoms (hydride) would split the signal into doublet (n+1 rule), as labelled	54
Figure 4.3	IR spectra of the PDMS before and after hydrosilylation process	55
Figure 4.4	¹ H-NMR spectra of the PDMS-MA precursors. The signal labelled as X is the AMA residual	56
Figure 4.5	General schemes of the hydrosilylation reaction	57

Figure 4.6	Colour changes of PDMS-MA (H) precursor after extraction	58
Figure 4.7	The photocrosslinking mechanism of PDMS-MA using Irgacure 651 (DMPA) as photoinitiator	62
Figure 4.8	Proposed 3-D crosslinked network structure of PDMS-MA gels	63
Figure 4.9	Effects of (a) Mn and (b) AMA on degree of conversion. 6.25 AMA and 12.5 AMA in (b) refers to the amount of AMA loading in wt.%. The onset time at which the DC reached plateau was taken as the optimized curing time	64
Figure 4.10	(a) Photograph and (b) optical micrograph of the selected PDMS-MA gels	66
Figure 4.11	IR spectra of PDMS-MA (1K), PEG-DA (0.7K) precursors and their respective hydro(gels) before and after photocrosslinking reaction ($t_{crosslinking}$ = 30 minutes), respectively.	68
Figure 4.12	Length scales covered by different analytical technique for the assessment of the phase miscibility (Hillerström, 2009)	69
Figure 4.13	(a) Physical appearances of the PDMS (1K)-PEG mixtures and their respective PDMS (1K)-PEG hydrogels at different PEG wt.%, and (b) effects of AMA on 70:30 PDMS (6K)-PEG gels homogeneity	70
Figure 4.14	DSC thermogram of PDMS (1K)-PEG hydrogels	71
Figure 4.15	DSC thermogram of PDMS (6K)-PEG hydrogels	73
Figure 4.16	AFM phase diagram (top view) of the PDMS-PEG gels. 0, 5, 15 and 30 in the right hand side indicates the amount of PEG (wt.%) used in the preparation of PDMS-PEG hydrogel	75
Figure 4.17	Topographic images of the PDMS (6K)-PEG blends before and after incubated in water. Colour contrast from bright to dark represents a total range of 400 nm	77
Figure 4.18	(a) Phase image (top), (b) 3-D topographical images and (c) height images of the 95-5 PDMS (6K)-PEG hydrogels as the function of AMA loading. Colour contrast from bright to dark represents a total range of 300 nm	78

Figure 4.19	(a) Phase image (top view), (b) 3-D topographical images and (c) height images of the 70-30 PDMS (6K)-PEG hydrogels as the function of AMA loading. Colour contrast from bright to dark represents a total range of 300 nm	79
Figure 4.20	SEM micrograph of 70-30 of the PDMS (6K)-PEG with 12.5 wt.% AMA hydrogels	82
Figure 4.21	SEM micrograph of (a) 70-30 PDMS (6K)-PEG and (b) 70-30 PDMS (6K)-PEG with 6.25 wt.% AMA hydrogels	82
Figure 4.22	ESR % of the PDMS-PEG hydrogels. * Statistically significant from each other (effect of PEG) at $p \le 0.05$. # Statistically significant from each other (effect of the matrix's Mn) at $p \le 0.05$	83
Figure 4.23	ESR % of the PDMS (6K)-PEG hydrogels as the function of AMA wt.%. * Statistically significant from each other at $p \le 0.05$	84
Figure 4.24	Contact angle of the PDMS-PEG hydrogels. * Statistically significant from each other (effect of PEG) at $p \le 0.05$. # Statistically different from each other (effect of the matrix's Mn) at $p \le 0.05$	86
Figure 4.25	Contact angle of the PDMS (6K)-PEG hydrogels as the function of AMA wt.%. * Statistically significant from each other at $p \le 0.05$	87
Figure 4.26	Effects of AMA loading on gels hardness. * Statistically significant from each other at $p \leq 0.05$	89
Figure 4.27	Effects of AMA loading on gels cohesiveness. * Statistically significant from each other at $p \le 0.05$	90
Figure 4.28	Effects of AMA loading on gels resilience. * Statistically significant from each other at $p \le 0.05$	91
Figure 4.29	Storage modulus of the selected samples	92
Figure 4.30	Loss modulus of the selected samples	93
Figure 4.31	Amount of protein adsorbed to the selected hydrogels. * Statistically significant from the control (100-0 gels with 6.25 wt.% AMA) at $p \le 0.05$	94

- Figure 4.32 Cell viability of L929 cell lines (P18) as the function of matrix's 96 Mn , PEG and AMA. Error bar: \pm SD. * Statistically significant from the medium control at $p \le 0.05$. # statistically different from each other at $p \le 0.05$
- Figure 4.33 Cell morphology of the selected samples under an inverted 97 microscope at 10X magnification

LIST OF ABBREVIATIONS

μm	micrometer
¹ H-NMR	Proton nuclear magnetic resonance
2-D	Two dimensional
3-D	Three dimensional
AFM	Atomic force microscopy
AMA	Allyl methacrylate
ASTM	American Society of Testing and Materials
CROP	Cationic ring opening polymerization
DMA	Dynamic mechanical Analysis
DMAP	2, 2-dimethyl-2-phenyl-acetophenone
DSC	Differential scanning Calorimetry
ECM	Extracellular matrix
EDX	Electron-dispersive X-ray spectroscopy
ESR	Equilibrium swelling ratio
FDA	Food and drug administration
FESEM	Field-emission scanning electron microscopy
FTIR	Fourier transform infrared
GPC	Gel permeation chromatography
H ₂ 0	Water
НА	Hydroxyapatite

IPN	Interpenetrating hydrogels networks
IR	Infrared
mg	milligram
min	minutes (time)
ml	mililiter
nm	nanometer
HDPE	High density polyethylene
MA	Methacrylate
NMR	Nuclear magnetic resonance
NVP	1-vinyl-2-pyrrodinone
PAA	Poly(acrylic acid)
PCL	Poly(ε- caprolactone)
PDMS	Polydimethylsiloxane
PDMS-MA	Polydimethylsiloxane dimethacrylate
PDMS-SiH	Polydimethylsiloxane dihydrido
PEG	Polyethylene glycol
PEGDA	Polyethylene glycol diacrylate
PEO	Poly(ethylene oxide)
PGA	Poly(glycolic) acid
PHEMA	Polyhydroxyethylmethacrylate
PLA	Poly(lactic) acid

PLGA	Poly(lactic-co-glycolic) acid
PNIPAM	Poly(N-isopropylacrylamide)
PS	Polystrene
PVA	Polyvinyl alcohol
SEM	Scanning electron microscopy
ТСР	tri-calcium phosphate
TE	Tissue engineering
TPA	Texture profile analysis
UV	ultra-violet
UV-vis	UV-visible spectroscopy
XRF	X-ray fluorescence
β-TCP	β-tricalcium phosphate

LIST OF SYMBOLS

%	Percentage
M _d	Weight of dried samples
M _s	Weight of swollen samples
M _n	Number-average molecular weight
M _w	Weight-average molecular weight
T _c	Crystallization temperature
Tg	Glass transition temperature
T _m	Melting temperature
°C	Degree celcius
Е	Molar extinction coefficient
E'	Storage (elastic) modulus
E	Loss modulus
ESR	Equilibrium swelling ratio
Hz	Hertz
Ν	Newton
PDI	Polydispersity index
Rz	Surface roughness
SCA	Static contact angle
wt.%	Weight percentage

PENGKAJIAN SIFAT-SIFAT HIDROGEL UV-SAMBUNG SILANG BERDASARKAN PDMS UNTUK APLIKASI KEJURUTERAAN TISU

ABSTRAK

Kajian ini menunjukkan penghasilan hidrogel berdasarkan PDMS yang mempunyai sifat-sifat yang boleh disesuaikan. Dua UV-sambung silang PDMS yang mempunyai berat molekul (M_n=1k & 6k g/mol) disintesis dahulu dan kemudian disambung-silang dengan PEGDA (M_n=0.7k g/mol) pada pelbagai nisbah berat (wt.%), dengan irgacure sebagai pemula UV. Bagi PDMS yang mempunyai M_n yang sederhana (6k), alil metakrilat (AMA) digunakan sebagai pengubah reaktif untuk meningkatkan keserasian dua polimer yang sangat tidak saling melaruti. Campuran polimer akan menjadi hidrogel selepas terdedah kepada penyinaran UV yang mempunyai rantau panjang gelombang 315-400 nm pada intensiti purata ~ 8-10 mW / cm² selama 30 minit. Sifat-sifat keserasian, termal, pembengkakkan, pembasahan, mekanikal, penjerapan protein dan sitotoksisiti hidrogel akan dinilaikan. Daripada kajian kalorimetri pengimbasan berbeza (DSC), walaupun dua Tg didapati bagi hidrogel yang dihasilkan dari PDMS (1k) yang mempunyai Mn rendah, hidrogel tersebut keserasian yang baik disebabkan oleh permukaan adalah homogen pada setiap PEG wt.% seperti yang ditunjukkan oleh keputusan daya atomic mikroskop (AFM). Hidrogel yang dihasilkan dari PDMS (6k) sangat tidak serasi terutamanya berlaku kepada 30 wt.% PEG dengan pemisahan fasa-makro berlaku. Permasalahan ini telah diselesaikan selepas AMA dimasukkan. Pemisahan fasa bagi hidrogel mempengaruhi sifat-sifat lain di mana permukaan hidrogel yang lebih hidrofobik telah menurunkan sifat pembengkakkan dan pembasahan disebabkan oleh jumlah domain PEG yang sedikit, lalu menjadikannya kurang hidrofilik, selepas kemasukan AMA. Penjerapan protein di permukaan hidrogel ini adalah lebih tinggi jika permukaannya dikuasai oleh permukaan PDMS, namun penjerapan masih rendah jika berbanding dengan PDMS murni. Kekakuan hidrogel bertepatan dengan rangkaian tisu lembut yang boleh diterima pada ~ 0.5-1 MPa, dan kekakuan hidrogel tersebut meningkat dengan peningkatan PEG wt.%, dan penurunan AMA wt.%. Digabungkan dengan sitotoksisitas yang rendah, hidrogel yang dihasilkan berpotensi digunakan sebagai perancah dalam bidang kejuruteraan tisu.

EVALUATION OF PDMS-BASED UV-CROSSLINKED HYDROGELS PROPERTIES FOR TISSUE ENGINEERING APPLICATIONS

ABSTRACT

This work presents the fabrication of PDMS-based hydrogels with tunable properties via direct blending. Two UV-crosslinkable PDMS with different molecular weights (Mn=1k & 6k g/mol) were first synthesized and then UV-cured with PEGDA (M_n=0.7k g/mol) at various wt.% ratio, in the presence of Irgacure as photoinitiator. For the medium Mn PDMS (6k), allyl methacrylate (AMA) was used as reactive modifier to enhance compatibility of the two highly immiscible polymers. The liquid mixtures were converted into hydrogels after exposed to UV irradiation at a wavelength region of 315-400 nm at the average intensity of 10 mW/cm² for 30 minutes. Compatibility, thermal, swelling, wetting, mechanical, protein adsorption and cytotoxicity properties of these PDMS hydrogels were evaluated. From differential scanning calorimetry (DSC) study, although two T_g were observed in the hydrogels fabricated from the low Mn PDMS (1k), they were all compatible since the hydrogel surface was homogeneous at any PEG wt.% ratio, as supported by AFM result. The hydrogels fabricated from the PDMS (6k) were highly incompatible and this was especially the case for the 30 wt.% PEG with the occurrence of macrophase separation. This problem was solved with addition of AMA. The phase separation of these PDMS (6K) hydrogels affected other properties in which the more hydrophobic gel surface, after the addition of AMA, had lowered their swelling and wetting properties since there was a fewer amount of PEG domains to render the hydrophilic surface. Protein adsorption to these hydrogel was higher if the surface was dominated by the PDMS surfaces, yet the adsorption was still lower than the bare PDMS. Stiffness of the hydrogel was fall within an acceptable range of soft tissue at ~ 0.5-1 MPa, with the stiffness increased with the increased of PEG loading, and/or the decreased of AMA loading. Coupled with their non-cytotoxic property, the fabricated PDMS-based hydrogels could potentially be used as scaffolds for tissue engineering applications.

CHAPTER ONE

INTRODUCTION

1.1 Research background

Hydrogels are polymeric materials that have three-dimensional network structure with the ability of absorbing abundant water while maintain their integrity due to chemical and/or physical crosslinking (Xie et. al., 2017). Hydrogels have been widely investigated in the past few decades, due to their exceptional potential in wide range of applications, ranging from food industry (Xie et. al., 2017), agricultural (Vundavalli et. al., 2015), pharmaceutical (Peppas et. al., 1999) and tissue engineering (Munoz-Pinto et. al., 2012; Hou et. al, 2010). Among these applications, hydrogels for tissue engineering applications have become a major area of interest with several commercial products already developed, mostly in skin reconstruction (Chu et. al., 2002; Falanga & Sabolinski, 1999; Fitton et. al., 2001). Some unique properties that make hydrogels applicable in the field of tissue engineering include their excellent water-absorbing capabilities, a degree of softness that highly resemble to the natural tissues, biocompatibility and other attractive characteristic. During the last two decades, natural hydrogels were gradually replaced by synthetic hydrogels which has long service life, high capacity of water absorption, and high gel strength (Ahmed, 2015). In addition, synthetic hydrogels usually have well-defined structure that can be modified to yield tailorable functionality and degradability. Many synthetic hydrophilic polymers such as PEG, PVA, PAA, PNIPAAM and other synthetic polymers, well fits the definition of hydrogels. Among them, PEG which possess many unique properties likes hydrophilicity, flexibility, non-toxicity, non-immunogenicity and low non-specific proteins adsorption, has been widely employed as hydrogels in the field of tissue engineering (Varghese et. al., 2009).

Compared with the single-component hydrogels, researchers nowadays prefer to use multi-component hydrogels, since single-component hydrogels cannot fulfil all the criteria required for an ideal scaffold, such as they are mechanically fragile and non-degradable. Micro- or nano-composite hydrogels, copolymeric hydrogels, multipolymer interpenetrating polymeric hydrogels (IPN), semi-IPN hydrogels and polymer blends are some examples of promising multi-component hydrogels that are widely investigated due to their sustainability in the field of tissue engineering. In these hydrogels, new properties that are different from the intrinsic properties of the original materials can be easily endowed by combining two or more components together. For instance, organic-inorganic PEG/PDMS hydrogels has been fabricated by Hou et. al. (2010) and they found that these hydrogel scaffolds demonstrated the ability to guide mesenchymal stem cells (MSCs) towards osteogenic differentiation with increased levels of PDMS microparticles. Besides, a nanocomposite made up of PEG and clay has been developed by Varghese et. al. (2009) and they elucidated that the overall mechanical properties of PEG hydrogels were improved by adding up to 10 wt.% of clay. It is worth to mention that one of the similarity between both studies is that the hydrogels were prepared by photocrosslinking technique in their respective projects. As compared to thermal or redox initiated crosslink mechanisms, photo-induced free radical hydrogen crosslinking produces less heat while allowing for improved spatial and temporal control (Hou et. al, 2010), due to short-term UV-exposure, i.e. within a minutes. Hence, photopolymerization is generally considered as a safe method to encapsulate cells. Another advantage of in situ polymerization is that specific shapes can be tailored made to fit exactly the tissue defects need to be repaired.

UV-curable PDMS-based hydrogels is another class of hydrogels, which are widely used as contact lenses (Lin et. al., 2014). Compared to the hydrogels which made up of only hydrophilic chains, these hydrogels is mainly consisted of hydrophobic PDMS. They also possess the unique properties of PDMS, such as biocompatible, high gas permeability, low T_g and unique viscoelastic properties when lightly crosslinked. PDMS have been used in various biomedical applications, such as transdermal (Mikolaszek et. al., 2016), antifouling coating (Gu et. al., 2016), ultrafiltration, drug delivery system (Racles, 2013) and tissue engineering applications (Munoz-Pinto et. al., 2012; Sung et. al., 1999; Pedraza et. al., 2013;

Si et. al., 2016). Although the potential uses of PDMS as scaffolds have been widely studied, their hydrophobicity always hinders their applicability in biomedical applications, due to non-specific protein adsorption following implantation (Wong & Ho, 2009; Zhang & Chiao, 2015). This phenomenon is highly unfavourable since protein adsorption to the hydrophobic surface is often irreversible and proteins will denature once they absorb to the surface. Surface modification of PDMS is a facile method to endow the surface hydrophilic, but it is often involved a complex process which consumes time and the uses of solvents which is toxic. Compared to surface modification, blending of PDMS with PEG is a simple and time saving method to impart hydrophilicity not only in the surface, but also within the bulk. Regard to this, this project focuses on the fabrication of two-component hydrogels that are comprised of two different acrylate-functionalized polymers, which is PDMSMA as the major phase and PEGDA as the minor phase, by means of photocrosslinking reaction. By the end, it should be able to translate the PDMS-PEG products into hydrogels by varying the number-average molecular weight (M_n) of PDMS and the composition of PDMS and PEG.

1.2 Problem statement

Controlled synthesis of PDMS with a well defined M_n is a prerequisite for the success of this project since physical and chemical properties of a crosslinked polymer is mainly governed by the M_n of matrix. Cationic ring opening polymerization (CROP) is a facile method to obtain the predesigned molecular weight and the molecular architecture, as described elsewhere (Toskas et. al., 2006). D₄ monomers is widely used to afford the linear homopolymer PDMS chains and the SiH containing endcappers is used to terminate the growing chains at the end of reaction (Bi et. al., 2007). Many groups have used this chemical scheme to synthesis a myriad of functionalized PDMS. For example, Hou et. al. (2010) had further functionalized the SiH-terminated PDMS chains with allyl methacrylate (AMA) via hydrosilylation reaction to endow them the photocrosslinking moieties.

The fabrication of PDMS-based hydrogels via solution polymerization technique is highly unfavourable since aqueous solvent is not miscible with PDMS (Lee et. al., 2003). On

the other hand, organic solvent is not often used due to its cytotoxicity. Therefore, the crosslinking reaction of PDMS is usually done in bulk polymerization technique. Conversely, one issue dealing with the bulk technique is the effects of molecular weight and/or viscosity on the curing efficiency of a polymer, particularly referred to the high M_n homologs. Regard to this matter, reactive diluents has been widely used to induce dilution effect during the crosslinking reaction of a polymer, mostly in epoxy coating (Li et. al., 2014). However, only few research papers have been reported on the use of reactive diluents in PDMS, due to the fact that PDMS has limited miscibility with most reactive diluents, such as acrylate- and methacrylate based reactive diluents. Therefore, microphase separation was identified by the observation of two T_g due to the separated reactive diluent phases (Yu et. al., 1985; Pouget et. al., 2009). In this project, AMA is proposed to be incorporated into the PDMS curing formulation to facilitate the crosslinking reaction since it fulfils the basic definition of reactive diluent (Ash, 2007). AMA should be incorporated in the right amount to avoid the phase separation problem, even though the functionalized methacrylate (MA) moieties in the terminal ends of PDMS chains made it more miscible with the reactive diluent.

PDMS is inherently hydrophobic, making it difficult for water to penetrate into its crosslinked structure. Therefore, PDMS should be blended with PEG in order to transform them into hydrogels. One of the issues dealing with this technique is the degree of miscibility or phase separation when blending the two polymers together. Macro-phase separation will occur if the PDMS-PEG blends suffer a change of composition, which force them into non-stable region. To avoid the phase separation problem, the M_n of PDMS and the amount of PEG loading should be carefully designed to get a desirable compromise between swelling properties and material's compatibility. Besides, PEG was used to switch the PDMS surface from hydrophobic to hydrophilic one. Nevertheless, micro-phase segregation of PEG to the surface remains a challenge since PEG tends to buried within the PDMS matrix for their low surface energy (Gu et. al., 2016; Wang et. al., 2011). Therefore, surface properties of these

hydrogels were carefully controlled by varying the PDMS-PEG composition, until PEG is micro-phase segregated to render the surface hydrophilic.

1.3 Objectives of the study

The objectives of this study are simplified as follows.

- 1. To synthesize PDMSMA precursors with a well-defined molecular weight (M_n)
- 2. To study the effects of different M_n of PDMS and different reactive diluent loading on the curing characteristics of the PDMSMA precursors
- To investigate the effects of different ratio of PDMS to PEG on the compatibility, swelling, surface, mechanical, protein adsorption and toxicity properties of the PDMS-PEG hydrogels

1.4 Research scope

The acid-catalyzed CROP process was used to synthesize PDMS precursors with low PDI value. The equilibrium chain redistribution during the reaction was likely to impart variations on the PDI value of the PDMS precursors, especially for the high M_n homologs (12000 g/mol). Therefore, the PDI value of PDMS precursor was first determined by GPC and ¹H-NMR end group analysis, respectively. UV-crosslinking of the PDMS precursor was only further proceeded if the PDI value obtained was less than or equal to 1.5.

Stickiness of the pristine PDMS gel can cause handling problem during sample characterization, especially referred to the PDMS with M_n of 6000 g/mol and 12000 g/mol. Besides, this issue might cause cytotoxicity in *in-vitro* testing as the stickiness of the gels also reflected to a considerable amount of leftover unreacted oligomer. The issue should be first addressed before it is further blended with PEG. Therefore, curing profile was first developed to determine the optimized curing time (t_c) as the function of different M_n of PDMS and different reactive diluent loading, in order to fabricate the PDMS gels with the least unreacted oligomers in the shortest possible time. However, PDMS precursors with M_n