

**EFFECTS OF TUALANG HONEY ON PARAQUAT
INTOXICATION IN RATS**

TANG SUK PENG

UNIVERSITI SAINS MALAYSIA

2016

**EFFECTS OF TUALANG HONEY ON
PARAQUAT INTOXICATION IN RATS**

by

TANG SUK PENG

**Thesis submitted in fulfilment of the requirements
for the Degree of
Doctor of Philosophy**

December 2016

ACKNOWLEDGEMENTS

First and foremost, I would like to express my deepest gratitude to my supervisors, Prof. Dr. Siti Amrah Sulaiman, Prof. Dr. Gan Siew Hua and Assoc. Prof. Dr. K.N.S. Sirajudeen for their invaluable guidances, advice and continual supports. Thank you for being so patient with me and for never failing to offer help whenever I encountered any difficulty in my research. I wish to express my deepest appreciation to Prof. Dr. Hasnan Jaafar for his professional guidance and his forever willingness to help and to give his best suggestions. My special thanks to Dr. Muzaimi Mustapha for his willingness to be part of my research team and for his invaluable suggestions. Overall, without the knowledge and assistance from my supervisors this study would not have been successful.

I would like to thank Prof. Dr. Aida Hanum Ghulam Rasool, Head of Department of Pharmacology, Prof. Dr. Mohd Suhaimi Abdul Wahab, former Head Department of Pharmacology and all staff from Pharmacology Department for their support and kind assistances during the study period. Special thanks to Pn. Norzihana.

I would like to acknowledge all staff and former staff from Animal Research and Service Centre (ARASC), Pathology Lab and Central Research Lab (CRL) who gave the permission to use all required equipment and the necessary materials to complete the different experiments. Not forgetting the lecturers from Biostatistic Unit, School of Medical Sciences, for their patient guidances in statistical analysis. My special thanks to Prof. Dr. Tan Soo Choon for his tremendous support in GC/MS and LC-MS/MS analysis in his lab as well as the entire members of USAINS Biomics Sdn.

Bhd for their help in chromatographic analysis in USM Main Campus, Penang. I am also grateful to Pn. Che Nin Man from the National Poison Centre, USM for her kind advice in GC/MS analysis.

I am also grateful to Dr. Mahaneem Mohamed, Dr. Erejuwa Omotayo, Dr. Fadhli, Dr. Wan Nazirah, Era, Kak Sarah, Kak Wardah, Chua Yung An, Ka Liong, Safiah, Kah Haw, Sam, Siew Wai, Ching Ching, Foo, Yi Yi, Farah, Kak Suzana, Kak Aida, Seetha, Lavaniya, Kak Azeera, Shafika, Cik Haswati, Cik Syazwani and many others. I sincerely appreciate all the help, directly or indirectly rendered during my study.

I would like to acknowledge Universiti Sains Malaysia (USM) for financially support the project under Research University (RU) grant (Grant No.: 1001/PPSP/813055) and RU-Postgraduate Research Grant Scheme (RU-PRGS, Grant No.: 1001/PPSP/8142002). I would like to express my sincere appreciation to USM and Ministry of Higher Education, Malaysia for the scholarship award under *Rancangan Latihan Kakitangan Akademik/ Skim Latihan Akademik IPTA (RLKA/SLAI)*. I would also like to acknowledge Federal Agricultural Marketing Authority, Malaysia for supplying Tualang honey in this study and Kaneka Corporation, Japan for their kind supply of ubiquinol (Kaneka QH™).

Last but not least, I would like to express my heartiest appreciation to my family and friends from all over the places for their endless care and understandings over the years. Thank you so much for everything.

TABLE OF CONTENTS

	Page
ACKNOWLEDGEMENTS	ii
TABLE OF CONTENTS	iv
LIST OF TABLES	xi
LIST OF FIGURES	xii
LIST OF PLATES	xiv
LIST OF SYMBOLS AND ABBREVIATIONS	xv
ABSTRAK	xviii
ABSTRACT	xx
CHAPTER 1 INTRODUCTION	1
1.1 Research Background.....	1
1.2 Objectives.....	5
1.2.1 Study 1: The effects of Tualang honey on single-high dose exposure of paraquat in rats (acute toxicity study).....	5
1.2.2 Study 2: The effects of Tualang honey on repeated-low dose exposures of paraquat in rats (subacute toxicity study)	6
CHAPTER 2 LITERATURE REVIEW	7
2.1 Paraquat.....	7
2.1.1 History and uses	7
2.1.2 Mode of action as herbicide	8
2.1.3 Paraquat toxicity.....	8
2.2 Toxicokinetics of Paraquat.....	9
2.3 Mechanism of Paraquat Toxicity	10

2.4	Acute Health Effects of Paraquat	15
	2.4.1 Clinical classification of acute paraquat poisoning	17
	2.4.2 Clinical management of paraquat poisoning	18
2.5	Chronic Health Effects of Paraquat.....	21
	2.5.1 Paraquat neurotoxicity.....	21
	2.5.2 Paraquat-induced oxidative stress-related neuronal damage.....	23
2.6	Honey	25
	2.6.1 Antioxidant properties of honey	25
	2.6.1.1 Phenolic compounds.....	28
	2.6.1.2 Organic acids and amino acids	29
	2.6.1.3 Maillard reaction products	29
	2.6.1.4 Enzymes.....	30
	2.6.2 Tualang honey	30
CHAPTER 3 MATERIALS AND METHODS.....		33
3.1	Materials.....	33
	3.1.1 Tualang honey	33
	3.1.2 Ethics statement.....	33
	3.1.3 Animals	33
	3.1.4 Chemicals and reagents	34
	3.1.5 Commercial kits and consumables	34
	3.1.6 Instrument.....	34
3.2	Study 1: The effects of Tualang Honey on Single-High Dose Exposure of Paraquat in Rats	39
	3.2.1 Preparation of paraquat and honey working solutions	39
	3.2.2 Experimental design	39

3.2.2.1	Determination of paraquat and honey doses for study 1.....	39
3.2.2.1 (a)	Determination of paraquat dose	39
3.2.2.1 (b)	Determination of Tualang honey dose	42
3.2.2.2	Evaluation of different honey dosing regimens in paraquat intoxicated rats	44
3.2.3	Histological analysis.....	47
3.2.3.1	Preparation of working solutions.....	47
3.2.3.2	Tissue processing.....	48
3.2.3.3	H & E staining of paraffin-embedded tissue sections ..	49
3.2.3.4	Histological assessment	49
3.3	Study 2: The Effects of Tualang Honey on Repeated-Low Dose Exposures of Paraquat in Rats.....	50
3.3.1	Experimental design	50
3.3.2	Sample collection for biochemical analysis	51
3.3.2.1	Blood collection and serum preparation.....	51
3.3.2.2	Brain and lung tissue collection and preparation of tissue homogenate.....	53
3.3.2.2 (a)	Preparation of working solutions	53
3.3.2.2 (b)	Whole brain and lung tissue collection	53
3.3.2.2 (c)	Identification and dissection of midbrain region.....	54
3.3.2.2 (d)	Preparation of tissue homogenate	58
3.3.3	Sample collection for histological and immunohistochemical analysis	59
3.3.3.1	Preparation of working solutions.....	59
3.3.3.2	Tissue collection, fixation and processing.....	60
3.3.3.2 (a)	Whole animal perfusion-fixation.....	60
3.3.3.2 (b)	Tissue collection and post-fixation	60
3.3.3.2 (c)	Midbrain tissue sectioning and processing for immunohistochemical analysis	63

3.3.3.2	(d) Lung tissue sectioning for H & E staining	63
3.3.4	Measurement of oxidative stress markers in the midbrain	65
3.3.4.1	Total protein content	65
3.3.4.2	Superoxide dismutase assay	66
3.3.4.3	Catalase assay	71
3.3.4.4	Glutathione peroxidase assay	75
3.3.4.5	Glutathione reductase assay	78
3.3.4.6	Glutathione-S-transferase assay	80
3.3.4.7	Total glutathione assay	83
3.3.4.8	Total antioxidant capacity	89
3.3.4.9	Malondialdehyde concentrations	90
3.3.5	Measurement of tyrosine hydroxylase concentrations in the midbrain	94
3.3.6	Immunohistochemical assessment of tyrosine hydroxylase in the substantia nigra pars compacta	97
3.3.7	Measurement of oxidative stress markers in the lung	100
3.3.7.1	Total protein content	100
3.3.7.2	Superoxide dismutase assay	100
3.3.7.3	Catalase assay	100
3.3.7.4	Glutathione peroxidase assay	100
3.3.7.5	Glutathione reductase assay	100
3.3.7.6	Glutathione-S-transferase assay	100
3.3.7.7	Total glutathione assay	101
3.3.7.8	Total antioxidant capacity	101
3.3.7.9	Malondialdehyde concentrations	101
3.3.8	Histological assessment of the lung	101
3.4	Statistical Analysis	102
CHAPTER 4 RESULTS		103

4.1	Study 1: The Effects of Tualang Honey on Single-High Dose Exposure of Paraquat in Rats	103
4.1.1	Determination of the dose of paraquat and Tualang honey.....	103
4.1.1.1	Determination of the paraquat dose	103
4.1.1.2	Determination of Tualang honey dose.....	106
4.1.2	Outcome of several different honey dosing regimens in paraquat-intoxicated rats	108
4.1.2.1	Survival analysis	108
4.1.2.2	Histological assessment	110
4.1.2.2 (a)	Lung	110
4.1.2.2 (b)	Kidney	116
4.1.2.2 (c)	Liver	121
4.2	Study 2: The Effects of Tualang honey on Repeated-Low Dose Exposures of Paraquat in Rats.....	126
4.2.1	Body weight	126
4.2.2	Serum biochemical parameters	129
4.2.2.1	Lipid profiles	129
4.2.2.2	Kidney function tests	129
4.2.2.3	Liver function tests	129
4.2.3	The effects of Tualang honey administration on repeated paraquat exposures in the oxidative stress markers of the midbrain region of rats	133
4.2.3.1	Superoxide dismutase activity	133
4.2.3.2	Catalase activity.....	133
4.2.3.3	Glutathione peroxidase activity	136
4.2.3.4	Glutathione reductase activity	136
4.2.3.5	Glutathione-S-transferase activity	136
4.2.3.6	Total glutathione level	136

4.2.3.7	Total antioxidant capacity.....	142
4.2.3.8	Malondialdehyde level.....	142
4.2.4	The effects of Tualang honey administration on repeated paraquat exposures in the midbrain dopaminergic neurons of rats	145
4.2.4.1	Measurement of tyrosine hydroxylase concentrations in the midbrain.....	145
4.2.4.2	Immunohistochemical assessment of tyrosine hydroxylase in the substantia nigra pars compacta region	147
4.2.5	The effects of Tualang honey administration on repeated paraquat exposures in the oxidative stress markers of the rats' lung.....	150
4.2.5.1	Superoxide dismutase activity	150
4.2.5.2	Catalase activity.....	150
4.2.5.3	Glutathione peroxidase activity	153
4.2.5.4	Glutathione reductase activity	153
4.2.5.5	Glutathione-S-transferase activity	153
4.2.5.6	Total glutathione level	153
4.2.5.7	Total antioxidant capacity.....	158
4.2.5.8	Malondialdehyde level.....	158
4.2.6	Histological assessment of lung	161
CHAPTER 5 DISCUSSION.....		163
5.1	Study 1: The Effects of Tualang Honey on Single-High Dose Exposure of Paraquat in Rats	163
5.1.1	Determination of the the dose of paraquat and honey.....	163
5.1.2	Evaluation of the outcome of different honey dosing regimens in paraquat-intoxicated rats.....	165

5.2	Study 2: The Effects of Tualang honey on Repeated-Low Dose Exposures of Paraquat in Rats.....	169
5.2.1	Body weight changes.....	169
5.2.2	Serum biochemical parameters	170
5.2.2.1	Lipid profiles	170
5.2.2.2	Renal and liver function tests	172
5.2.3	The effects of Tualang honey administration on repeated paraquat exposures in the midbrain region of rats	174
5.2.4	The effects of Tualang honey administration on repeated paraquat exposures in the rats' lung.....	184
5.2.5	The plausible protective mechanism of Tualang honey.....	185
CHAPTER 6 CONCLUSION		188
6.1	Summary and Conclusion	188
6.2	Study Limitations and Recommendations for Future Research.....	190
REFERENCES.....		192
APPENDICES		
LIST OF PUBLICATIONS AND PRESENTATIONS		

LIST OF TABLES

	Page
Table 2.1	Effects of honey on oxidative stress parameters in several toxin-induced disease models. 26
Table 2.2	Phenolic compounds, organic acids and water-soluble vitamins detected in Tualang honey..... 32
Table 3.1	List of chemicals and reagents. 35
Table 3.2	List of commercial kits..... 37
Table 3.3	List of instruments..... 38
Table 3.4	Rats grouping and type of treatment in all experimental groups. 45
Table 3.5	Preparation of SOD working standard solutions..... 69
Table 3.6	Preparation of H ₂ O ₂ working standard solutions..... 73
Table 3.7	Preparation of GSSG working standard solutions..... 86
Table 3.8	Preparation of MDA working standard solutions..... 92
Table 4.1	Effects of different honey dosing regimen on median survival time of paraquat-intoxicated rats..... 109
Table 4.2	Percentage of body weight changes relative to week 0 in all experimental groups..... 128
Table 4.3	Comparison of serum lipid profiles between the different experimental groups..... 130
Table 4.4	Comparison of kidney function tests between the different experimental groups..... 131
Table 4.5	Comparison of liver function tests between the different experimental groups..... 132

LIST OF FIGURES

	Page
Figure 2.1 Structures of paraquat and putrescine	11
Figure 2.2 Schematic representation of the mechanism of paraquat toxicity.....	12
Figure 2.3 Pathogenesis of paraquat-induced lung toxicity.....	14
Figure 2.4 Structures of paraquat, MPTP and its active metabolite MPP ⁺	22
Figure 3.1 Experimental procedures for determination of paraquat dose.....	41
Figure 3.2 Experimental procedures for determination of TH dose	43
Figure 3.3 Experimental procedure to evaluate the outcome of different honey dosing regimen in PQ-intoxicated rats.	46
Figure 3.4 Experimental procedure to evaluate the protective effects of Tualang honey on repeated exposure of paraquat in rats' midbrain and lung.	52
Figure 3.5 Perfusion-fixation of rats through heart.	62
Figure 3.6 Principle of SOD assay.....	67
Figure 3.7 Principle of GPx assay	75
Figure 3.8 Principle of GST assay	81
Figure 3.9 Principle of total GSH assay	84
Figure 4.1 Percentage of rats showing severe signs of toxicity within one week after paraquat intoxication grouped by paraquat dose investigated.....	105
Figure 4.2 Mortality rate and mean duration to death of paraquat-intoxicated rats grouped by Tualang honey dose tested.....	107
Figure 4.3 Mean body weight (g) of rats throughout the experimental periods ..	127
Figure 4.4 SOD activity in the midbrain region.	134

Figure 4.5	CAT activity in the midbrain region.	135
Figure 4.6	GPx activity in the midbrain region.	138
Figure 4.7	GR activity in the midbrain region.	139
Figure 4.8	GST activity in the midbrain region.	140
Figure 4.9	Total GSH level in the midbrain region.	141
Figure 4.10	TAC in the midbrain region.	143
Figure 4.11	MDA level in the midbrain region.	144
Figure 4.12	TyrH concentration in the midbrain region.	146
Figure 4.13	The number of TyrH-positive neurons in the midbrain region.	149
Figure 4.14	SOD activity in the lung.	151
Figure 4.15	CAT activity in the lung.	152
Figure 4.16	GPx activity in the lung.	154
Figure 4.17	GR activity in the lung.	155
Figure 4.18	GST activity in the lung.	156
Figure 4.19	Total GSH level in the lung.	157
Figure 4.20	TAC in the lung.	159
Figure 4.21	MDA level in the lung.	160

LIST OF PLATES

	Page
Plate 3.1	Structural landmarks on the adult rat brain used for dissection of midbrain region. 55
Plate 3.2	Positioning of rat brain onto brain blocker and diagrammatic figure illustrating placement of razor blades 56
Plate 3.3	Dissection of the midbrain region of rat..... 57
Plate 3.4	Indicator for proper perfusion. 61
Plate 3.5	Dissection of coronal striatum and midbrain sections..... 64
Plate 4.1	Representative photomicrographs of H & E stained lung section..... 112
Plate 4.2	Representative photomicrographs of H & E stained kidney section.. 117
Plate 4.3	Representative photomicrographs of H & E stained liver section 122
Plate 4.4	TyrH-IHC staining of the coronal midbrain section 148
Plate 4.5	Representative photomicrographs of H & E stained lung section..... 162

LIST OF SYMBOLS AND ABBREVIATIONS

%	percentage
$\cdot\text{OH}$	hydroxyl radical
μ	micro
A/G	albumin/globulin
AEAC	ascorbic acid equivalent antioxidant capacity
ALP	alkaline phosphatase
ALT	alanine transaminase
ANOVA	analysis of variance
A-P	anterior-posterior
APAP	acetaminophen
AST	aspartate transaminase
BG	blood glucose
BSA	bovine serum albumin
BUN	blood urea nitrogen
CAT	catalase
Cb	cerebellum
CC	cerebral cortex
CCl_4	carbon tetrachloride
CDC	Centers for Disease Control and Prevention
CDNB	1-chloro-2,4-dinitrobenzene
CoQ10	coenzyme Q10
CP	cerebral peduncle
DAB	3,3'-diaminobenzidine
ddH ₂ O	double distilled water
df	degrees of freedom
DOA	Department of Agriculture, Malaysia
DPPH	2,2,-diphenyl-1-picrylhydrazyl
DTNB	5,5'-dithio- <i>bis</i> -2-nitrobenzoic acid
DTPA	diethylenetriaminepentaacetic acid
EDTA	ethylenediaminetetraacetic acid
ELISA	enzyme-linked immunosorbent assay
FAMA	Federal Agricultural Marketing Authority
FR	Fenton reaction
FRAP	ferric reducing antioxidant power
g	gram
<i>g</i>	gravitational force
GGT	gamma-glutamyl transferase
GPx	glutathione peroxidase
GR	glutathione reductase
GSDNB	1-S-glutathionyl-2,4-dinitrobenzene
GSH	reduced glutathione
GSSG	glutathione disulfide (oxidized GSH)
GST	glutathione-S-transferase
H & E	haematoxylin and eosin
H ₂ O ₂	hydrogen peroxide
HCl	hydrochloric acid

HDL	high density lipoprotein
HMF	hydroxymethylfurfural
Hpc	hippocampus
HRP	horseradish peroxidase
Hth	hypothalamus
HWR	Haber-Weiss reaction
i.m.	intramuscular
i.p.	intraperitoneal
i.v.	intravenous
IC	inferior colliculus
IL-1 β	interleukin-1 beta
IPCS	International Program on Chemical Safety
IQR	interquartile range
K ₂ HPO ₄	potassium phosphate, dibasic
KCl	potassium chloride
KH ₂ PO ₄	potassium dihydrogen phosphate
L	litre
LDL	low density lipoprotein
LPO	lipid peroxides
LV	left ventricle
m	milli
MDA	malondialdehyde
MES	2-(N-morpholino)ethanesulfonic acid
MO	medulla oblongata
MOH	Ministry of Health, Malaysia
MPA	metaphosphoric acid
MPP ⁺	1-methyl-4-phenylpyridinium ion
MPTP	1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine
Na ₂ HPO ₄	sodium phosphate, dibasic
NaCl	sodium chloride
NADP ⁺	nicotinamide adenine dinucleotide phosphate
NADPH	reduced nicotinamide adenine dinucleotide phosphate
NaH ₂ PO ₄	sodium dihydrogen phosphate
NaOH	sodium hydroxide
nm	nanometer
O ₂	oxygen
O ₂ ⁻	superoxide anion
OC	optic chiasm
OECD	Organization for Economic Co-operation and Development
ORAC	oxygen radicals absorbance capacity
p.o.	per os
Pb	lead
PBS	phosphate buffer saline
PFA	paraformaldehyde
PG	pituitary gland
PoA	preoptic area
PQ	paraquat
PQ ^{•+}	paraquat mono-cation radical
PQ ²⁺	paraquat dication
QH	ubiquinol

RA	right atrium
ROH	alcohols
ROOH	hydroperoxides
ROS	reactive oxygen species
s.c.	subcutaneous
SC	superior colliculus
SD	standard deviation
SEM	standard error of mean
SN	substantia nigra
SNpc	substantia nigra pars compacta
SOD	superoxide dismutase
SPSS	Statistical Package for the Social Sciences
TAC	total antioxidant capacity
TB	total bilirubin
TBA	thiobarbituric acid
TBS	Tris-buffered saline
TEAC	Trolox equivalent antioxidant capacity
TH	Tualang honey
Thal	thalamus
TMB	3,3',5,5'-tetramethylbenzidine
TNB	5-thio-2-nitrobenzoic acid
TP	total protein
Tris-HCl	tris(hydroxymethyl)aminomethane-hydrochloride
Trolox	6-hydroxy-2,5,7,8-tetramethylchromane-2-carboxylic acid
TyrH	tyrosine hydroxylase
v/v	volume per volume
w/v	weight per volume
WBC	white blood cells
WHO	World Health Organization
χ^2	chi-squared

KESAN MADU LEBAH TUALANG TERHADAP INTOKSIKASI PARAKUAT PADA TIKUS

ABSTRAK

Parakuat (PQ) adalah sejenis racun herba yang digunakan dengan meluas di seluruh dunia dan telah dipostulasi menunjukkan kesan toksik melalui penghasilan pelbagai spesies oksigen reaktif berikutan dengan kerosakan oksidatif pada komponen utama sel. Madu lebah Tualang (TH) dilaporkan mempunyai sifat antioksidan dan berkemungkinan membantu memperbaiki kerosakan oksidatif dalam kes keracunan PQ. Oleh itu, objektif utama kajian ini adalah untuk menilai kesan perlindungan madu TH pada ketoksikan akut (kajian 1) dan subakut PQ (kajian 2) di kalangan tikus. Tikus jantan Sprague-Dawley berusia lapan minggu digunakan. Pada kajian 1, dos oral PQ dan TH ditentukan pada 225 mg/kg dan 0.2 g/kg, masing-masing. Kesan rawatan TH tunggal dan berulang seterusnya dikaji: kumpulan rawatan tunggal menerima TH pada 0.5 (PQ + TH0.5h), 2 (PQ + TH2h) atau 6 (PQ + TH6h) jam selepas pemberian PQ; kumpulan rawatan berulang menerima TH pada 0.5, 2 dan 6 jam (PQ + THtrp) atau rawatan diteruskan sebanyak sekali sehari untuk enam hari berterusan (PQ + TH7d), masing-masing. Masa mandiri untuk setiap tikus dicatat sehingga hari ke-28 sebelum ia dikorbankan. Rawatan dengan TH tidak dapat menambah baik kadar kemandirian tikus yang diracunkan oleh PQ. Namun demikian, kumpulan tikus yang menerima rawatan berulang menunjukkan masa mandiri median yang lebih panjang secara signifikan berbanding dengan kumpulan PQ + TH6h. Rawatan dengan TH juga menambah baik hasil histologi pada tikus yang diracunkan dengan PQ terutamanya pada peparu yang mencadangkan bahawa kemungkinan kegunaan TH dapat melambatkan kesan toksik PQ. Pada kajian 2, kesan perlindungan TH pada tekanan oksidatif aruhan PQ pada otak tengah dan

peparau dikaji. Tikus dirawat dengan air suling (kumpulan N dan PQ; 2 mL/kg/hari), TH (kumpulan TH dan PQ + TH; 1.0 g/kg/hari) atau ubiquinol (PQ + QH; 0.2 g/kg/hari) setiap hari sepanjang masa kajian. Dua minggu selepas rawatan masing-masing, tikus diberikan larutan salin (N dan TH; 1 mL/kg/minggu, i.p) atau PQ (PQ, PQ + TH dan PQ + QH; 10 mg/kg/minggu, i.p) seminggu sekali selama empat minggu berterusan. Tikus dikorbankan seminggu selepas suntikan terakhir salin atau PQ. Hasil kajian menunjukkan bahawa kumpulan yang menerima TH (TH dan PQ + TH) atau ubiquinol (PQ + QH) mengandungi kandungan urea serum yang lebih rendah ($p < 0.05$). Kreatinina serum juga menurun secara signifikan di dalam kumpulan PQ + TH berbanding dengan kumpulan kawalan N dan TH. Kandungan ALT yang lebih rendah ($p < 0.05$) juga dilihat pada kumpulan TH dan PQ + TH apabila dibandingkan dengan kumpulan PQ. Perubahan ini mencadangkan bahawa rawatan TH mungkin memberikan kesan bermanfaat terhadap fungsi ginjal dan hati tikus. Berikutan pemberian PQ empat mingguan, aktiviti GPx otak tengah menurun secara signifikan. Kerosakan neuron dopaminergik akibat aruhan PQ dilihat apabila terdapat pengurangan signifikan bilangan neuron tirosina hidroksilase-immunopositif di kawasan *substantia nigra pars compacta* otak tengah. Penurunan signifikan aktiviti-aktiviti SOD dan GST di peparu juga diperhatikan pada kumpulan PQ berbanding dengan kumpulan N. Secara keseluruhan, rawatan TH dapat memperbaiki kesan-kesan toksik yang dilihat di otak tengah dan peparu, di mana kesan perlindungan TH adalah setanding dengan ubiquinol, ubat kawalan yang digunakan dalam kajian 2. Hasil kajian mencadangkan bahawa rawatan TH dapat memberi kesan perlindungan terhadap ketoksikan subakut PQ. Kesan ini mungkin berlaku akibat sifat antioksidan yang dimiliki TH.

EFFECTS OF TUALANG HONEY ON PARAQUAT INTOXICATION IN RATS

ABSTRACT

Paraquat (PQ) is an herbicide widely used in the world and has been postulated to exert its toxic effects via the production of various reactive oxygen species causing subsequent oxidative damage at key cellular components. Tualang honey (TH) has been reported to possess good antioxidant properties and may help ameliorate the oxidative damages in case of PQ poisoning. Therefore, the main objectives of this study were to evaluate the possible protective effect of TH in acute (study 1) and subacute (study 2) PQ toxicities in rats. Male Sprague-Dawley rats aged eight weeks old were used. In study 1, selected oral doses of PQ and TH were 225 mg/kg and 0.2 g/kg, respectively. The effects of single and multiple TH treatments on PQ-intoxicated rats were then investigated: single TH treatment groups received TH at 0.5 (PQ + TH0.5h), 2 (PQ + TH2h) or 6 (PQ + TH6h) hours following PQ administration; multiple TH treatment groups received TH at 0.5, 2 and 6 hours (PQ + THtrp) or further daily treatment for next six days (PQ + TH7d) following PQ administration, respectively (n = 6 per group). The survival time of each rat was recorded until day 28 before sacrifice. Although treatment with TH did not improve the survival rate of PQ-intoxicated rats, the median survival time of rats which received multiple TH treatments was significantly longer when compared to group PQ + TH6h. Furthermore, TH treatment improved the histological outcome of PQ-intoxicated rats particularly in the lungs, thus suggesting the potential role of honey in delaying the toxic effects of PQ. In study 2, the protective effect of TH on PQ-induced oxidative stress in rats' midbrain and lung regions were investigated (n= 15 per group). The rats were orally treated with distilled water (Groups N & PQ, 2

mL/kg/day), TH (Groups TH & PQ + TH, 1.0 g/kg/day) or ubiquinol (Group PQ + QH, 0.2 g/kg/day) throughout the experimental period. Two weeks after the respective treatments, the rats were administered with saline (Groups N & TH; 1 mL/kg/week, i.p) or PQ (10 mg/kg/week, i.p.; Groups PQ, PQ + TH and PQ + QH) once a week for four consecutive weeks. The animals were then sacrificed a week following the final injection of saline or PQ. Serum urea was significantly lower in groups which received TH (TH and PQ + TH) or ubiquinol (PQ + QH). Serum creatinine was markedly reduced in group PQ + TH as well, when compared to controls (N and TH). Significantly lower levels of ALT were observed in groups TH and PQ + TH when compared to group PQ. These findings suggest that TH treatment may have some beneficial effects on the kidney and liver's function. Following four-weekly PQ-administration, the midbrain GPx activity was significantly reduced when compared to healthy control (N). PQ-induced dopaminergic neuronal damage was demonstrated by a significant reduction in the number of tyrosine hydroxylase-immunopositive neurons in the midbrain substantia nigra pars compacta while in the lungs, marked reduction in the activities of SOD and GST were observed in group PQ when compared to the control group. Treatment with TH ameliorates the toxic effects seen in the midbrain and lungs with comparable effects to ubiquinol, the control drug used in study 2. These findings suggest that pre-treatment with TH showed some protective effects against subacute PQ toxicities. It is plausible that the protective effects of TH are conferred by its antioxidant properties.

CHAPTER 1

INTRODUCTION

1.1 RESEARCH BACKGROUND

Problems resulting from paraquat (PQ) exposure are reported all over the world and are mainly caused by suicidal intent, accidental poisoning or occupational exposures (Eddleston and Bateman, 2007; IPCS, 1991). PQ is known to be a producer of various reactive oxygen species (ROS) via single electron redox cycle *in vivo* (Abdollahi *et al.*, 2004; Autor *et al.*, 1977; Bus and Gibson, 1984). ROS readily attacks key cellular structures and molecules (lipids, carbohydrates, proteins and nucleotides) thus causing cellular deleterious effects which formed the basis of various disease conditions (Halliwell, 2009; Slater, 1984; Valko *et al.*, 2007).

In acute toxicity, high concentrations of PQ often lead to death due to multi-organ failures with major pneumotoxic effects seen (Bismuth *et al.*, 1990; Dinis-Oliveira *et al.*, 2007a; Fung *et al.*, 1999; Ghazi-Khansari *et al.*, 2005; Krieger, 2001). PQ is selectively concentrated in the lungs where the high alveolar oxygen content further increase the degree of lung injury by oxidative damage and exerts its major pneumotoxic effects in acute poisoning, thus producing diffuse alveolitis and loss of surfactant followed by progressive pulmonary fibrosis (Bismuth *et al.*, 1990; Yeh *et al.*, 2006). Similar effects may also occur in other major organ systems that eventually lead to death in the animals from multiple organ failure. In fact, ingestion of high doses of PQ usually leads to death from multi-organ failures. Smaller ingestion doses may also lead to delayed death as a result of extensive pulmonary

fibrosis and respiratory failure (Dinis-Oliveira *et al.*, 2006b; Suntres, 2002). Conventional approach in treatment of PQ poisoning focused on three main areas which include prevention of absorption from the gastrointestinal tract, enhancement of elimination of PQ from the body and administration of therapies directed against toxicity. Nevertheless, these treatment methods have been disappointing and the mortality rate remained very high (Eddleston and Bateman, 2007; Franzen *et al.*, 1991; Gawarammana and Buckley, 2011; Lin *et al.*, 1996).

On the other hand, unlike the major pneumotoxic effects occurring during acute poisoning, repeated low dose exposures to PQ is believed to be neurotoxic (Cicchetti *et al.*, 2005; Dinis-Oliveira *et al.*, 2006a; Franco *et al.*, 2010; Hatcher *et al.*, 2008). The effects of chronic exposure to PQ have gained considerable attention due to its wide usage around the world particularly among agricultural workers and PQ formulation workers (Cannon and Greenamyre, 2011; Drechsel and Patel, 2008; Tanner *et al.*, 2011). More alarmingly, there are growing evidences from epidemiological studies which implicated the possible involvement of environmental toxin such as pesticides as an aetiological factor for Parkinson's disease (Franco *et al.*, 2010; Hatcher *et al.*, 2008; McCormack *et al.*, 2002; Semchuk *et al.*, 1993).

Studies conducted by Hertzman *et al.* (1990) and Liou *et al.* (1997) indicated that exposure to PQ are associated with higher incidence of Parkinson's Disease. PQ-induced oxidative stress has been demonstrated in the substantia nigra region of Parkinson's disease brain (Franco *et al.*, 2010). A study by Ranjbar *et al.* (2002) reported that workers from PQ formulating factory are also at higher risk of having an oxidative stress as evidenced by elevated lipid peroxidation and decreased of

antioxidant power. In experimental models, rats or mice exposed to PQ demonstrate a selective degeneration of nigrostriatal dopaminergic neurons, one of the neuropathological hallmark of Parkinson's disease (Brooks *et al.*, 1999; Kang *et al.*, 2010; Kuter *et al.*, 2007; LeDoux, 2005; McCormack *et al.*, 2006).

Since the main suggested mechanism for PQ toxicity, either acute or chronic, involves the production of ROS, it may be hypothesized that an antidote against PQ poisoning should be a substance with good antioxidant properties. Honey is known to be rich in both enzymatic and non-enzymatic antioxidants. It contains both aqueous and lipophilic antioxidants with the interaction between these antioxidants suggesting its potential as an ideal natural antioxidant that can act at different cellular sites in the case of PQ poisoning (Angela, 2003; Küçük *et al.*, 2007; Nagai *et al.*, 2006).

Tualang honey is a wild honey harvested from the Tualang trees found in the Malaysia rain forest. Various studies have been conducted in an effort to evaluate the possible medicinal uses of Tualang honey including investigating its anti-cancer properties in cell culture and animal model (Fauzi *et al.*, 2011; Ghashm *et al.*, 2010), the protective effects of honey from cigarette smokes' damage (Mohamed *et al.*, 2011b), animal menopausal model (Zaid *et al.*, 2010), anti-diabetic properties (Erejuwa *et al.*, 2009; Erejuwa *et al.*, 2010a; Erejuwa *et al.*, 2010b) as well as in wound management and as an antimicrobial (Khoo *et al.*, 2010; Nasir *et al.*, 2010; Tan *et al.*, 2009). In addition, the neuroprotective effects of Tualang honey have also been previously reported in chronic cerebral hypoperfusion-induced neurodegeneration in the hippocampus of rats (Saxena *et al.*, 2014). Supplementation of Tualang honey improved the hippocampal and medial prefrontal cortex

morphology, memory performances and cholinergic system in stressed ovariectomised rats (Al-Rahbi *et al.*, 2014a; Al-Rahbi *et al.*, 2014b). One of the purported mechanisms of action is mainly contributed by its antioxidant properties.

A study by Mohamed *et al.* (2010) showed that Tualang honey contains phenolic compounds with good antioxidant activities. However, to date the protective effects of honey on PQ toxicity have not been investigated. Therefore, the main purpose of this study is to evaluate the possible protective effects of Tualang honey on PQ-induced acute toxicity (study 1) and subacute toxicity (study 2) in Sprague-Dawley rats.

1.2 OBJECTIVES

1.2.1 Study 1: The effects of Tualang honey on single-high dose exposure of paraquat in rats (acute toxicity study)

The general objective for study 1 is to evaluate the possible protective effect of Tualang honey in acute PQ poisoning. The specific objectives for study 1 include:

- (i) To determine the dose of PQ that contributes to severe signs of toxicity following a week of its administration to rats.
- (ii) To determine the dose of Tualang honey that can ameliorate PQ-intoxication in rats.
- (iii) To determine the effects of single and multiple doses of Tualang honey treatments based on the survival rate at 28 days following PQ-intoxication in rats.
- (iv) To determine the effects of single and multiple doses of Tualang honey treatments on the lung, kidney and liver's histological changes of PQ-intoxicated rats.

1.2.2 Study 2: The effects of Tualang honey on repeated-low dose exposures of paraquat in rats (subacute toxicity study)

The general objective for study 2 is to evaluate the possible protective effects of Tualang honey in the midbrain region and lungs of rats exposed to four weekly administration of PQ. The specific objectives for study 2 include:

- (i) To determine the effects of Tualang honey on serum biochemical profiles of rats exposed to four weekly administration of PQ.
- (ii) To determine the effects of Tualang honey on the oxidative stress parameters and tyrosine hydroxylase levels in the midbrain of rats exposed to four weekly administration of PQ.
- (iii) To determine the effects of Tualang honey on immunohistochemical detection of tyrosine hydroxylase in the substantia nigra pars compacta region of rats exposed to four weekly administration of PQ.
- (iv) To determine the effects of Tualang honey on oxidative stress parameters and histological changes in the lung of rats exposed to four weekly administration of PQ.

CHAPTER 2

LITERATURE REVIEW

2.1 PARAQUAT

2.1.1 History and uses

Paraquat (PQ) or 1,1'-dimethyl-4,4'-bipyridinium, is a synthetic quaternary ammonium compound first described by Weidel and Rosso in 1882. The redox properties of PQ was discovered by Michaelis and Hill (1933) in 1933. In the early years PQ was commonly called methyl viologen and was used as a redox indicator. This is because PQ dication (PQ^{2+}) readily undergoes single electron reduction to form a stable free radical monocation ($PQ^{\cdot+}$) with blue or violet colours (Krieger, 2001). However, its herbicidal properties were only discovered in 1955 and finally it was first introduced to the market as a herbicide in 1962 by Imperial Chemical Industries Limited (ICI, now Syngenta). PQ is usually formulated as the dichloride salt. Although the patent protection on PQ has expired, Syngenta, one of the world's largest agrochemical companies, remains the major manufacturer for PQ under the trade name Gramoxone®, accounting for at least 50% of its total world market (Dinham, 2003; Wesseling *et al.*, 2001).

PQ is a non-selective, fast-acting contact herbicides applied for wide spectrum control of broad-leaved and grassy weeds, but have no effects on matured bark. It is used on over 100 crops in approximately 100 countries, some of which includes United States of America, China, Mexico, Thailand, Malaysia and Japan (Syngenta, 2015; Wesseling *et al.*, 2001). Its frequent application in a wide variety of plantation

crops have been reported to increase agricultural productivity in both developed and developing countries (Carlile, 2006; Centers for Disease Control and Prevention, 2003; IPCS, 1984).

2.1.2 Mode of action as herbicide

PQ exerts its herbicidal activity through the disruption of the normal electron flow in photosystem I which in turn inhibits the light reaction of photosynthesis. Once in the chloroplast where photosynthesis occurs, the positively charged PQ ion reacts with the free electron from photosystem I to produce PQ radicals. The radicals then react with oxygen to form various reactive oxygen species (ROS) which attack the unsaturated fatty acids of membranes, resulting in the destruction of membrane integrity and eventually cell death. Therefore, PQ is also known as cell membrane disruptors (Ashton and Crafts, 1981).

2.1.3 Paraquat toxicity

Despite its beneficial effects in agriculture, PQ is toxic to human and animals. Since its introduction in the early 1960's, fatalities due to intentional, accidental or occupational exposure to PQ have been frequently reported (IPCS, 1984; WHO, 2010). PQ is classified by the World Health Organization (WHO, 2010) as class II or moderately hazardous pesticide for acute toxicity based on its oral median lethal dose (LD₅₀) in rat (150 mg/kg). It is of relatively low hazard when used with adequate personal protective measures. However, serious delayed side effects may be developed which may even be fatal when the concentrated product is orally ingested or become in direct contact with the skin (WHO, 2010). The US Environmental Protection Agency (EPA) classified PQ dichloride into different categories of acute

toxicity based on the type of administration route i.e. highly toxic (category I) by inhalational route, moderately toxic (category II) by oral route, slightly toxic (category III) by dermal route; moderate to severe eye irritation (category II), and minimal dermal irritation (category IV) (Environmental Protection Agency, 1997).

2.2 TOXICOKINETICS OF PARAQUAT

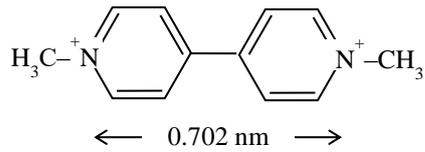
The intact skin is relatively impermeable to PQ but dermal absorption is enhanced when the skin is damaged (Wester *et al.*, 1984). Most cases of PQ poisonings however, results from oral ingestion. A rapid but incomplete absorption of ingested PQ occurs mainly from the small intestine. In human, it is estimated that only less than five per cent of the ingested amount reach the bloodstream over one to six hours following its ingestion (Houze *et al.*, 1990).

Regardless of its route of administration into the mammalian systems, PQ is rapidly distributed via the blood circulation to all organs and tissues of the body although its storage is not prolonged in any tissues, since it is being eliminated by the kidneys between days and weeks. The absorbed PQ is largely (> 90%) excreted as a parent compound through the kidneys within 24 hours if normal kidney function is retained. Disturbance of normal renal function following high ingestion doses may cause renal tubular necrosis which in turn affects the elimination-distribution and accumulation in other organs (Hawksworth *et al.*, 1981). Houze *et al.* (1990) described a bi-exponential plasma concentration-time curve, with the initial and late phases of elimination half-lives being five and 84 hours, respectively. Plasma PQ is then selectively concentrated in the lungs via an energy-dependent process due to its structural similarity with naturally-occurring polyamines (such as putrescine,

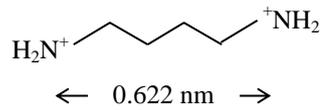
cadaverine, spermidine and spermine) following which it is taken up by the alveolar cells (Fung *et al.*, 1999) and exerts its major pneumotoxic effects. The structures of PQ and diamine putrescine are shown in Figure 2.1.

2.3 MECHANISM OF PARAQUAT TOXICITY

The main suggested mechanism for PQ toxicity is its ability to undergo cellular redox cycling with the subsequent production of various ROS (Bus and Gibson, 1984). Similar to its herbicidal mode of action, PQ^{2+} is readily reduced to mono-cation radical ($PQ^{\cdot+}$) by several enzymes including NADPH-cytochrome P450 reductase, xanthine oxidase, NADH-ubiquinone oxidoreductase (complex I of mitochondrial respiratory chain) and nitric oxide synthase (Day *et al.*, 1999; Fukushima *et al.*, 1993; Han *et al.*, 2006; Kitazawa *et al.*, 1991; Tawara *et al.*, 1996; Winterbourn and Button, 1984). $PQ^{\cdot+}$ is then rapidly re-oxidized to its original dicationic form in the presence of oxygen (electron acceptor). During the process, superoxide radical is generated at the expense of NADPH (electron donor). Further redox cycling of PQ leads to generation of other ROS including hydrogen peroxide and hydroxyl radical. Consequently, depletion of cellular NADPH may result in the disturbance of NADPH-requiring biochemical processes. Furthermore, ROS can induce oxidative damage to lipids, proteins and nucleic acids (Bus and Gibson, 1984; Dinis-Oliveira *et al.*, 2008; Lascano *et al.*, 2012). Schematic representation of mechanism of PQ toxicity is illustrated in Figure 2.2.



paraquat



putrescine

Figure 2.1 Structures of paraquat and putrescine showing the geometric standards of the distance between nitrogen (N) atoms. The substrate selectivity for polyamine uptake system includes two (or more) charged nitrogens, an ideal distance between the nitrogen and a nonpolar spacer to separate these charges (Boelsterli, 2007; Dinis-Oliveira *et al.*, 2008)

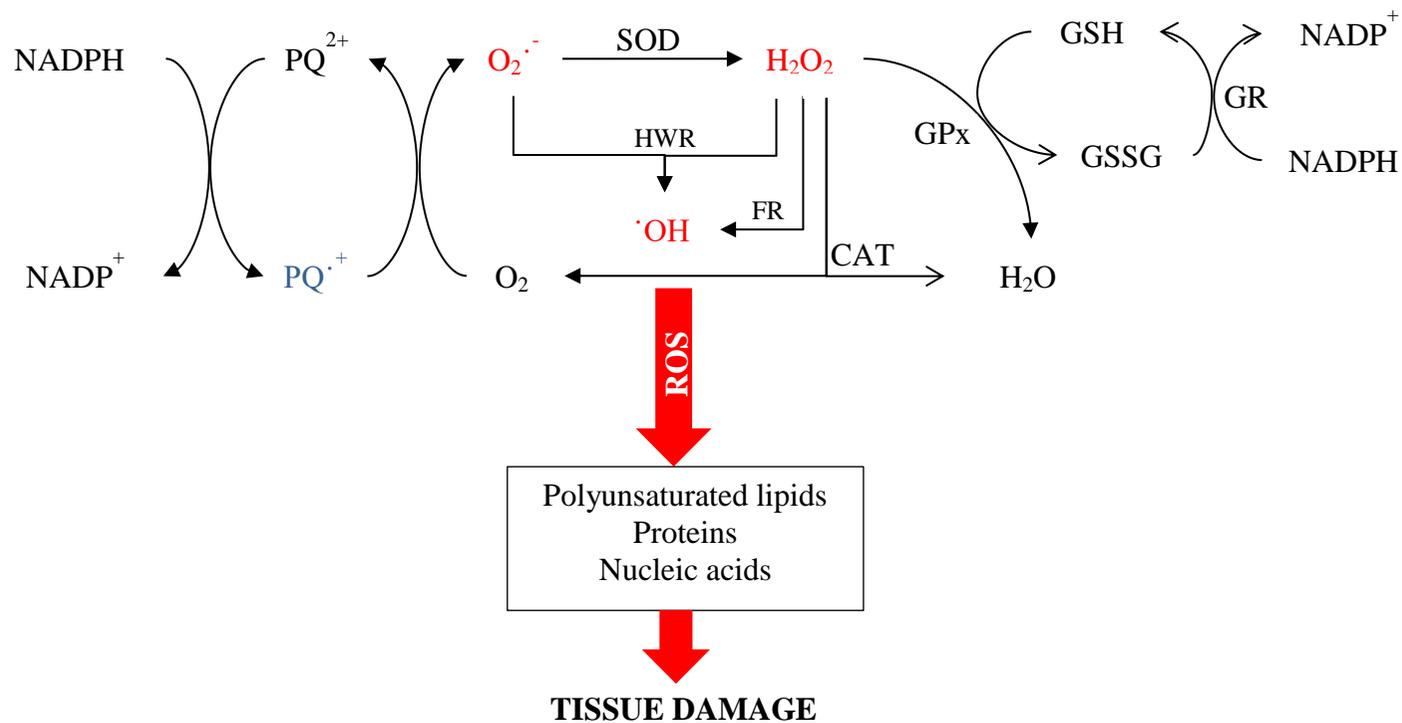
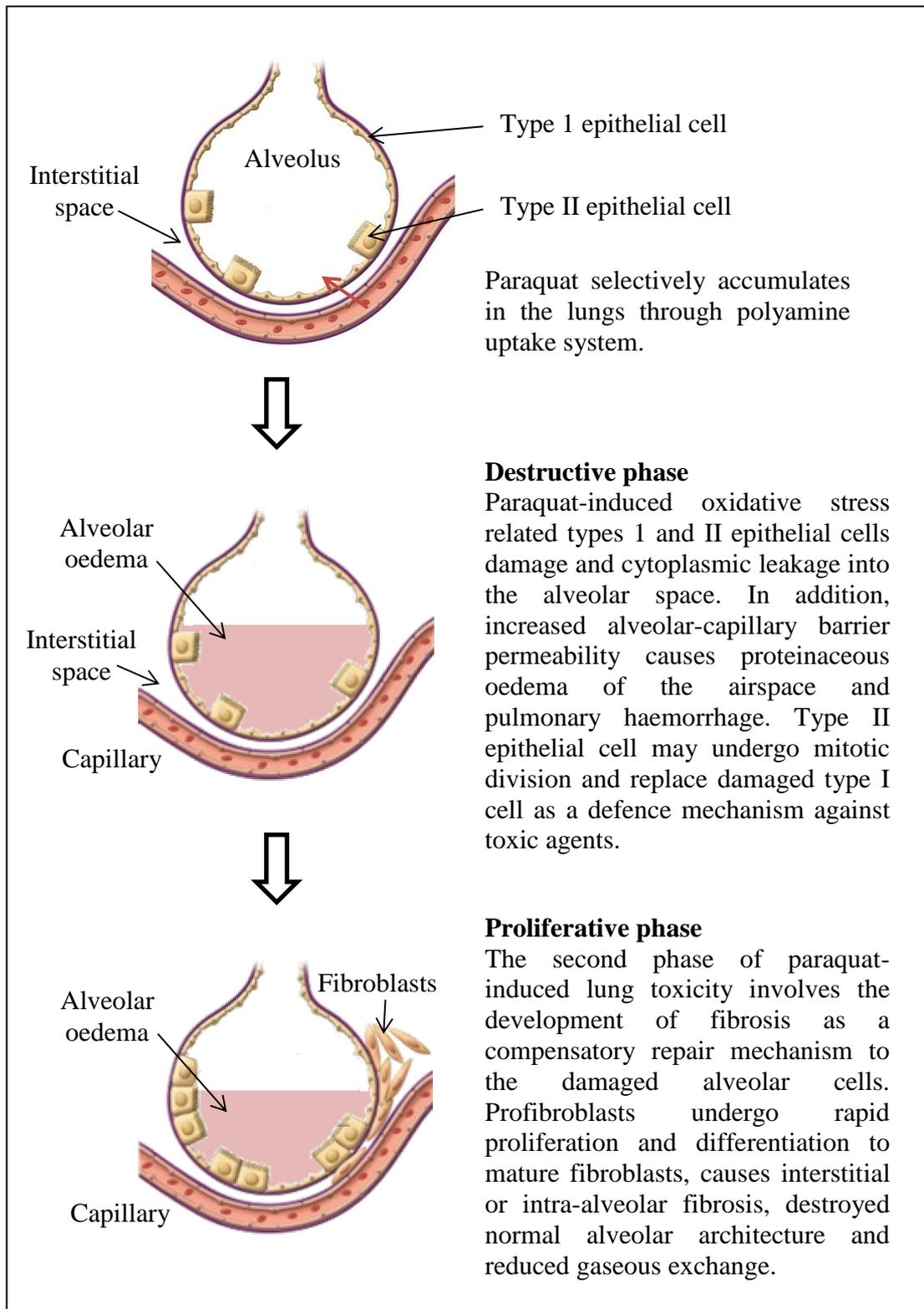


Figure 2.2 Schematic representation of the mechanism of paraquat toxicity. (Abbreviations: CAT, catalase; GPx, glutathione peroxidase; GR, glutathione reductase; GSH, reduced glutathione; GSSG, glutathione disulfide; H₂O₂, hydrogen peroxide; HWR, Haber-Weiss reaction; FR, Fenton reaction; NADP⁺, nicotinamide adenine dinucleotide phosphate; NADPH, reduced nicotinamide adenine dinucleotide phosphate; O₂, oxygen; O₂^{·-}, superoxide anion; ·OH, hydroxyl radical; PQ²⁺, paraquat; PQ^{·+}, paraquat mono-cation radical; ROS, reactive oxygen species; SOD, superoxide dismutase) [Adapted from Dinis-Oliveira *et al.* (2008)].

PQ-induced generation of ROS and the subsequent oxidative stress reactions occur in most organs. However, the toxic effects of PQ are particularly severe in the lungs due to its selective accumulation through the polyamine uptake system. There are two distinct phases in the development of pulmonary fibrosis by PQ. In the early destructive phase, the alveolar types I and II epithelial cells are damaged, resulting in a second proliferative phase defined by alveolitis and finally leading to an extensive lung fibrosis (Bus and Gibson, 1984). Pulmonary damage is manifested by oedema, haemorrhage, infiltration of inflammatory cells into the interstitial and alveolar spaces and proliferation of bronchial epithelium. Eventually, individuals die of respiratory failure (Bus and Gibson, 1984; Dinis-Oliveira *et al.*, 2008; Ghazi-Khansari *et al.*, 2005; Smith *et al.*, 1990; Suntres, 2002) (Figure 2.3).



Paraquat selectively accumulates in the lungs through polyamine uptake system.

Destructive phase

Paraquat-induced oxidative stress related types 1 and II epithelial cells damage and cytoplasmic leakage into the alveolar space. In addition, increased alveolar-capillary barrier permeability causes proteinaceous oedema of the airspace and pulmonary haemorrhage. Type II epithelial cell may undergo mitotic division and replace damaged type I cell as a defence mechanism against toxic agents.

Proliferative phase

The second phase of paraquat-induced lung toxicity involves the development of fibrosis as a compensatory repair mechanism to the damaged alveolar cells. Profibroblasts undergo rapid proliferation and differentiation to mature fibroblasts, causes interstitial or intra-alveolar fibrosis, destroyed normal alveolar architecture and reduced gaseous exchange.

Figure 2.3 Pathogenesis of paraquat-induced lung toxicity.[Adapted from Dinis-Oliveira *et al.* (2008) and Matthay *et al.* (2012)]

2.4 ACUTE HEALTH EFFECTS OF PARAQUAT

Acute pesticide poisoning, often due to single-high level exposure, is one of the major health concerns of pesticide usage. Globally, suicides or intentional self-poisoning is considered as a major contributing factor for acute pesticide poisoning (Jeyaratnam, 1990). The estimated number of suicides in the year 2012 were 804 000 worldwide with 75.5% of the global suicides occurring in the low and middle income countries including Africa, Central America, South-East Asia and the Western Pacific regions (WHO, 2014). Pesticide self-poisoning is one of the three most common methods for suicide, accounting for approximately 30% of the global suicides, especially in the rural agricultural areas of low and middle income countries (WHO, 2014). Besides intentional poisonings, it has also been reported that an estimated 355,000 fatalities occurred due to unintentional poisonings worldwide (WHO, 2003).

Due to its popularity and ease of access, PQ is extensively used as a suicide agent particularly among developing countries (Gunnell and Eddleston, 2003; Jeyaratnam, 1990; Wesseling *et al.*, 2001). Nevertheless, a study by Seok *et al.* (2009) showed that only 38% of parasuicidal subjects intentionally select PQ suggesting that almost two-thirds of the subjects who ingested PQ occurred simply due to its wide availability during suicide attempts. On the other hand, accidental and occupational exposures to PQ were often associated with inappropriate use of pesticides, the lack of personal protective equipment or safety instructions on containers as well as accidental contamination with herbicide (Eddleston and Bateman, 2007; Eddleston *et al.*, 2002; Van Wendel de Joode *et al.*, 1996).

In Malaysia, almost three quarters (73.4%) of poisoning involving PQ were due to suicides, followed by accidental poisoning (13.8%) and occupational exposure (1.07%) (Jeyaratnam, 1990). Similar findings were reported between the year 2000 and 2006, where most chemical poisonings in Malaysia were parasuicidal, followed by accidental, homicidal and occupational-related poisonings where PQ is one of the main pesticides involved (MOH, 2006; Sirajuddin *et al.*, 2001). Poisoning (together with injury and other consequences of external causes) has been reported to be among the top ten reasons of hospitalisation and mortality in government hospitals in Malaysia (MOH, 2010; MOH, 2012). For instance, the review on the medical records of 79 PQ poisoning cases admitted to Hospital Taiping, Perak between January 2008 and October 2011 showed that 69.6% were of intentional exposure, followed by 26.6% accidental and 3.8% occupational exposures (Tan *et al.*, 2013).

Since pesticide poisoning accounts for nearly one-third of global suicides, it is suggested that death can be prevented if the use of toxic pesticides was restricted, or the access reduced through proper storage or disposal by individuals or communities. In addition, optimizing the medical management and the quality of care for poisoning cases are necessary (Gunnell *et al.*, 2007).

In an effort to reduce the accessibility of this highly toxic herbicide, beginning from August 2002, the Malaysian government has decided to ban PQ's use while all other previously registered PQ products would be phased out in stages by November 2007. However, the ban was lifted in November 2006 and registration of PQ was allowed for all crops (Krishnamoorthy, 2006). To date, 59 pesticides which contain PQ dichloride as an active ingredient are registered by the Pesticides Board, Department of Agriculture (DOA), Malaysia from February 2010 to January 2015 (DOA, 2015a).

Nevertheless, as of 2014, the use of herbicides containing PQ was restricted only to weed control in palm oil, rubber or dry paddy plantations as well as for killing of pineapple stumps only with its use forecasted to be banned again by the year 2020 (DOA, 2015b).

2.4.1 Clinical classification of acute paraquat poisoning

Clinical experiences of treating PQ-intoxicated patients help to predict the prognosis of PQ poisoning. Generally, the severity of PQ poisoning can be classified into three categories: mild, severe or acute fulminant poisoning. Patients who experienced mild poisoning, with an estimated ingestion of less than 20 mg PQ ion per kg body weight (or < 7.5 mL of 20% formulation for a 70 kg person), is often asymptomatic or may develop minor gastrointestinal symptoms such as vomiting and diarrhoea. However, in these cases, full recovery usually occurs (Vale *et al.*, 1987).

In cases of moderate to severe poisonings, with estimated ingestion of 20 to 40 mg PQ ion per kg body weight (or 7.5 – 15 mL of 20% formulation for a 70 kg person), patients may initially suffer from gastrointestinal symptoms, followed by multiple organs injuries. Proximal tubular dysfunction usually evolved and with renal insufficiency often noted, which may further affect the main route of PQ elimination and contribute to poor prognosis (Gil *et al.*, 2005). In addition, liver toxicity as revealed by associated biochemical abnormalities may also occur. Centrilobular hepatic necrosis and cholestasis are usually seen during postmortem examination of patients having impaired liver function (Dinis-Oliveira *et al.*, 2008). Death usually occurs in most cases. Nevertheless, patients who survived acute poisoning often developed pulmonary fibrosis, resulting in delayed death which usually occurs within

two to four weeks as a result of respiratory failure and is often fatal (Lock and Wilks, 2010; Vale *et al.*, 1987).

In cases of fulminant poisoning, with an estimated ingestion of more than 40 mg paraquat ion per kg body weight (or > 15 mL of 20% formulation for a 70 kg person), the mortality is almost 100%. Death usually occurs within the same day or no longer than a few days due to multiple organ failure where patients usually succumb before the development of pulmonary fibrosis (Reigart, 2009; Sabzghabae *et al.*, 2010; Vale *et al.*, 1987).

2.4.2 Clinical management of paraquat poisoning

Clinically, there is no specific antidote for PQ poisoning. Although a wide variety of therapies have been extensively studied, the mortality rate remains high with indeterminate treatment efficacies (Hong *et al.*, 2014; Nagami *et al.*, 2013). Therefore, the goal is to relieve symptoms and treat complications (i.e. to provide a supportive care). Conventional approaches in treating PQ poisoning include prevention of absorption from the gastrointestinal tract, enhanced elimination of PQ from the body and instituting therapy directed against toxicity symptomatically (Gawarammana and Buckley, 2011; Gil *et al.*, 2014).

The initial treatment of poisoning usually consists of administrations of oral absorbents such as Fuller's earth, activated charcoal or bentonite, all of which are aimed at neutralizing PQ in the gastrointestinal tract and enhancing its excretion in the faeces by using purgatives, mannitol or gastric lavage. For instance, routine treatment with multiple doses of activated charcoal also yielded no clinical benefit

with similar mortality rate compared to the non-charcoal treated groups (Eddleston *et al.*, 2008). Gastric lavage may be considered for patients who presented within an hour of ingestion of potentially life-threatening amount of PQ (Vale, 1997). However, serious complications which include hypoxia, gastrointestinal tract or pharynx perforation, disturbances of fluid and electrolytes balances may also occur (Benson *et al.*, 2013; Vale and Kulig, 2004).

Haemoperfusion, haemodialysis, haemofiltration and forced diuresis may be used to enhance excretion of systematically absorbed PQ or allow stabilization of haemodynamic status of patients with multiorgan failures (Gao *et al.*, 2015; Gawarammana and Buckley, 2011). A combined therapy with haemoperfusion and continuous venovenous haemofiltration may prolong the survival time and reduce the number of early deaths due to multiorgan failures, thus providing an opportunity for further treatment. However, the mortality rate remains high and patients eventually die from respiratory failure (Gao *et al.*, 2015; Koo *et al.*, 2002).

Other supportive therapies include surgical approaches (lung transplantation) and radiotherapy (to prevent fibroblast proliferation and ultimate interstitial fibrosis) to the diseased lungs although they have not been proven to be effective (Franzen *et al.*, 1991). There are some reports that pulse immunosuppression with the use of methylprednisolone and cyclophosphamide can help prevent lung fibrosis in PQ-intoxicated patients although further confirmation with clinical trials are still required (Koh *et al.*, 2014; Lin *et al.*, 2006; Lin *et al.*, 1996).

Current researches on PQ poisoning have also been directed towards the use of antioxidants, since PQ induces its toxic effect through oxidative stress-mediated mechanisms. For example, the use of superoxide dismutase (SOD) in PQ intoxication allows the conversion of highly toxic superoxide anions to potentially less toxic hydrogen peroxide and water. However, the finding from a study has demonstrated that administration of SOD by continuous intravenous infusion failed to improve PQ's toxic effects (Block, 1979). SOD also failed to protect against PQ poisoning in vitamin E-deficient animals, possibly because in the absence of vitamin E, peroxidative chain reactions are triggered and sustained by small amount of superoxide anion escaping detoxification by SOD (Block, 1979). Moreover, SOD cannot enter the target membrane or can hardly adhere to the targets due to its high molecular mass and charge (Ilizarov *et al.*, 2001; Suntres, 2002).

Other antioxidants investigated in search of the treatment for PQ poisoning include vitamins C and E, melatonin, iron chelators, low molecular weight thiol-containing antioxidants [e.g. glutathione (GSH), N-acetylcysteine, metallothionein] and mono-unsaturated fatty acids. However, most of the antioxidants failed to modify PQ's toxicity which is attributed by their inability to cross cell membrane barriers and/or due to their rapid clearance from cells (Suntres, 2002). More recently, the use of liposomal antioxidants (e.g. liposome-entrapped GSH and α -tocopherol liposomes) or low molecular weight SOD mimetics leads to increased therapeutic potentials against PQ pulmonary toxicity since they presumably facilitate the intracellular delivery. Therefore, much remains to be known about the use of intracellular and extracellular antioxidants in PQ toxicity with no single strategy appearing to show

improvement in the outcome for PQ poisoning (Suntres, 2002) indicating that there is still a need to search for new treatment modalities.

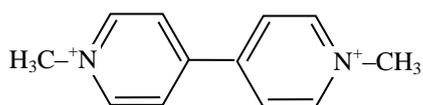
2.5 CHRONIC HEALTH EFFECTS OF PARAQUAT

In addition to the above, the possible delayed health effects from long term-low dose exposure is another major health concern of pesticide use particularly due to occupational exposure by the workers who were exposed to PQ for a long period of time. Additionally, chronic low dose exposures to PQ may also cause adverse respiratory effects among the workers. A study conducted among PQ workers in the Western Cape, South Africa indicated that under the usual field conditions, working with PQ over a long period of time is associated with desaturation of arterial oxygen, especially during exercise (Dalvie *et al.*, 1999). Castro-Gutierrez *et al.* (1997) reported an increase in the prevalence of respiratory symptoms among the workers. Analysis of blood samples from PQ-formulating factory workers have been reported to show increased in lipid peroxidation and decreased in antioxidant power (Ranjbar *et al.*, 2002).

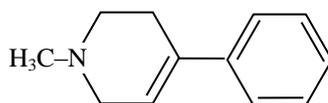
2.5.1 Paraquat neurotoxicity

In addition to its well-known pneumotoxic effects, the possible chronic neurotoxic effects of PQ have gained a wide interest over the past two decades due to its structural similarity with 1-methyl-4-phenylpyridinium ion (MPP⁺) (Tieu, 2011; Wu *et al.*, 2012) (Figure 2.4). MPP⁺ is an active metabolite of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) with known dopaminergic neurotoxic effect contributing to acute Parkinsonism in human (Langston *et al.*, 1983; Shimizu *et al.*, 2003b). The by-product from synthetic heroin is not naturally found in the environment as

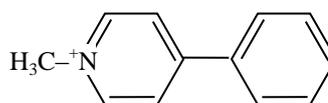
compared to the widely used herbicide PQ, further supported by human epidemiological studies which found that chronic exposure to pesticides including PQ, particularly among agricultural workers are associated with significantly higher incidence of Parkinson's disease (Hertzman *et al.*, 1990; Liou *et al.*, 1997; Petrovitch *et al.*, 2002).



Paraquat



MPTP



MPP⁺

Figure 2.4 Structures of paraquat, MPTP and its active metabolite MPP⁺. (Abbreviations: MPP⁺, 1-methyl-4-phenylpyridinium ion; MPTP, 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine).

2.5.2 Paraquat-induced oxidative stress-related neuronal damage

In an animal experimental model, a single PQ exposure did not induce any neurodegeneration in mice but can lead to microglial activation, making neurons more susceptible to damage following subsequent exposures (Purisai *et al.*, 2007). In an effort to understand the toxicokinetics of PQ in the brain, a study by Prasad *et al.* (2009) reported that PQ persists in the ventral midbrain with an estimated half-life of approximately 28 days, irrespective of its route of administration. The prolonged persistence suggests that accumulation of PQ following its repeated exposure is expected (Prasad *et al.*, 2009). Furthermore, systemic exposures of PQ in the rats or mice can induce selective dopaminergic neurodegeneration in the midbrain substantia nigra (SN) i.e. one of the neuropathological hallmarks of Parkinson's disease. In addition, dopamine depletion and increased levels of α -synuclein protein aggregation were also reported (Fernagut *et al.*, 2007; Manning-Boğ *et al.*, 2003; Wills *et al.*, 2012).

The brain naturally contains relatively low levels of antioxidants with high amounts of polyunsaturated fatty acids making it more susceptible to oxidative injuries due to redox imbalance. Of the brain's neuronal cell types, the dopaminergic neurons in the nigrostriatal system are selectively vulnerable to oxidative injury because dopamine metabolism itself can generate high levels of ROS (Chinta and Andersen, 2008). The role of oxidative stress in PQ-induced dopaminergic neurodegeneration in the SN region have previously been reported (Kang *et al.*, 2010; McCarthy *et al.*, 2004; Peng *et al.*, 2005; Somayajulu-Nitu *et al.*, 2009), thus suggesting the use of antioxidants as one of the possible therapeutic approaches for neurodegenerative disorders. For example, the study by Peng *et al.* (2005) demonstrated a protective role of synthetic

SOD/catalase mimetics in PQ-induced neurotoxicity in both rat dopaminergic cell lines and in adult mice. In addition, administrations of the antioxidant coenzyme Q₁₀ (as known as ubiquinone) have also shown some neuroprotective effects in animal models of neurodegenerative disease (Cleren *et al.*, 2008; Matthews *et al.*, 1998; Somayajulu-Nitu *et al.*, 2009). Human preliminary data suggested daily supplementation of coenzyme Q₁₀ appears to slow the disease progression in early Parkinson's disease (Shults *et al.*, 2002). Nevertheless, no evidence of clinical benefit was observed in a phase III clinical trial in patients diagnosed with Parkinson's disease within five years who received the coenzyme Q₁₀ up to 16 months intervention and observation (Beal *et al.*, 2014), indicating its questionable effects.

Meanwhile, a recent published report on Phase II clinical trial using ubiquinol showed significant improvement in Parkinson's patient with wearing off (Yoritaka *et al.*, 2015). Ubiquinol, the reduced form of coenzyme Q₁₀, was previously showed to be more effective in an MPTP-induced mouse model of Parkinson's disease when compared to the same dose of coenzyme Q₁₀ (oxidized form) tested (Cleren *et al.*, 2008). It is the most common form of coenzyme Q₁₀ in vivo accounts for more than 80% of the total coenzyme Q₁₀ (ubiquinol + ubiquinone) pool in human plasma, intestine and liver (Hosoe *et al.*, 2007). When administered orally, ubiquinol showed a higher bioavailability than the oxidized form of coenzyme Q₁₀ (Mae *et al.*, 2001). However, the clinical efficacy for ubiquinol in Parkinson's patients was yet to be confirmed in a larger clinical study.