ADAPTIVE AND COOPERATIVE HARMONY SEARCH MODELS FOR RNA SECONDARY STRUCTURE PREDICTION

ABDULQADER MOHAMMED A. MOHSEN

UNIVERSITI SAINS MALAYSIA 2011

ADAPTIVE AND COOPERATIVE HARMONY SEARCH MODELS FOR RNA SECONDARY STRUCTURE PREDICTION

by

ABDULQADER MOHAMMED A. MOHSEN

Thesis submitted in fulfilment of the requirements for the degree of Doctor of Philosphy

February 2011

ACKNOWLEDGEMENTS

This thesis would not have been possible without help of others who I would to thank. First, and for most, I thank Allah for all his blessings and guidance.

I would like to thank my supervisor Associate Prof. Dr. Ahamad Tajudin bin Khader for his supervision, encouragements, guidance, constructive, and for all of his help in different aspects of the research, giving valuable suggestions and providing constructive criticism throughout my research work and preparation of this thesis. Much of appreciation to my co-supervisor Dr. Dhanesh Ramachandram for his help and support during my Ph.D. study.

I would like to acknowledge School of Computer Sciences for all the facilities and support during this research.

Thanks to members of the Computational Intelligence Research Group who were always there to discuss any research-related and unrelated issues. Abdullatif Ghallab and Farhad Nadi who brought fond memories and friendships throughout the years. I also thank the other friends in School of Computer Science, in particular Waleed Shaher.

In addition, special thanks and profound gratitude should go to Waleed Ahmed, Abdul-Hameed Al-Namshah and Yahya Al-Marrani for helping me in proofreading the thesis.

Last but not least, many thanks also go out to my parents, and my wife and children. They have always shown their faithful support during the long hours I have spent in studying and writing. I appreciate their patience during the trying time of doctoral research. Finally, I am thankful for the prayer and encouragement of my family and friends.

TABLE OF CONTENTS

Ackr	owledge	ements		ii
Table	e of Cont	tents		iii
List	of Tables			ix
List	of Figure	es		xi
List	of Abbre	viations		xvi
List	of Symbo	ols		xix
Abst	rak			XX
Abst	ract			xxi
СНА	PTER 1	- INTRO	DDUCTION	
1.1	Proble	m statemer	at and Motivations	2
1.2	Harmo	ony Search	Algorithm	5
1.3	Object	ives		6
1.4	Scope	and Limita	tions	7
1.5	Resear	ch Approa	ch	8
1.6	List of	Contributi	ons	10
1.7	Thesis	Outline an	d Organization	11
СНА	PTER 2	– LITER	ATURE REVIEW	
2.1	Harmo	ony Search	Algorithm	12
	2.1.1	Fundame	ntal Procedures of HS	13
	2.1.2	HS Optin	nization Steps	14
		2.1.2(a)	Initialize the Problem and Algorithm Parameters	14
		2.1.2(b)	Initialize the Harmony Memory	16
		2.1.2(c)	Improvise a New Harmony	16
		2.1.2(d)	Harmony Memory Update	17

		2.1.2(e) Termination Criterion Check	18
	2.1.3	Adaptive Parameters	18
	2.1.4	Multiple Harmony Memories Models	21
2.2	RNA S	Secondary Structure	22
2.3	RNA S	Secondary Structure Determination	24
	2.3.1	X-ray Crystallography	25
	2.3.2	Nuclear Magnetic Resonance	25
	2.3.3	Secondary Structure Determination Methods Limitations	26
2.4	RNA S	Secondary Structure Prediction Methods	26
	2.4.1	Multiple Sequences Methods	27
	2.4.2	Single Sequence Methods	29
		2.4.2(a) Dynamic Programming Algorithms	29
		2.4.2(b) Metaheuristic Methods	32
2.5	Summ	ary	35
СНА	PTER 3	 HARMONY SEARCH ALGORITHM FOR RNA SECONDARY STRUCTURE PREDICTION 	
3.1	Adapta	ation of HS for RNA Secondary Structure Prediction	38
	3.1.1	RNA Secondary Structure Modeling and Representation	40
	3.1.2	Helix Generation Algorithm	41
	3.1.3	Harmony Decoding	43
	3.1.4	Objective Function	45
		3.1.4(a) efn2 Model	46
		3.1.4(b) RNAeval Model	46
3.2	Param	eters Adaptation Model	46
	3.2.1	Introduction	46
	3.2.2	AHSRNAFold	48
3.3	Coope	rative HS Model	50

	3.3.1	Number of HMs	52
	3.3.2	The Communication Interval.	54
	3.3.3	The Communication Rate	54
	3.3.4	The Communication Topology	55
	3.3.5	The Communication Policy	56
	3.3.6	CHSRNAFold	56
3.4	Summ	ary 5	58
СНА	PTER 4	- EXPERIMENTAL STUDY	
4.1	Config	uration of Experiments and Setup6	50
	4.1.1	Data Set of RNA Sequences	60
	4.1.2	Evaluation Measurement	52
	4.1.3	Statistical Analysis 6	53
	4.1.4	Computing Environments	54
	4.1.5	Choosing the Thermodynamic Model	54
		4.1.5(a) Objectives 6	54
		4.1.5(b) Detailed Description of the Results	54
		4.1.5(c) Discussion	67
	4.1.6	Modified Helix Generation Algorithm	67
		4.1.6(a) Objective	58
		4.1.6(b) Detailed Description of the Results	58
4.2	Experi	mental Study of HSs' Variants Parameters	70
	4.2.1	Experiment I: Parameter Settings of HSRNAFold	70
		4.2.1(a) Objective of the Experiment	70
		4.2.1(b) Experimental Setup	71
		4.2.1(c) Detailed Description of the Results	72
		4.2.1(d) Discussion	77
	4.2.2	Experiment II: Adaptation of the HS Parameters for RNA Folding 8	80

		4.2.2(a)	Objectives of the Experiment	80
		4.2.2(b)	The Experimental Setup	80
		4.2.2(c)	Detailed Description of the Results	81
		4.2.2(d)	Discussion	82
	4.2.3	Experime	ent III: Parameters Tuning of CHSRNAFold	83
		4.2.3(a)	Objective of the Experiment	84
		4.2.3(b)	The Experimental Setup	84
		4.2.3(c)	Detailed Description of the Results	85
		4.2.3(d)	Discussion	90
4.3	Experi	ment IV Co	omparison between the Three HS Variants	91
	4.3.1	Objective	e of the Experiment	92
	4.3.2	Detailed 1	Description of the Results	92
	4.3.3	Discussio	on	95
4.4	Summa	ary		95
			UATION AND DISCUSSION	
5.1	RNA S	Sequences I	Results and Discussion	96
	5.1.1	Saccharo	myces cerevisiae 118 nt	96
	5.1.2	Escherich	nia coli 120 nt	98
	5.1.3	Deinococ	ecus radiodurans 124 nt	101
	5.1.4	Metarhizi	ium anisopliae var. anisopliae(3) 394 nt	103
	5.1.5	Metarhizi	ium anisopliae var. anisopliae(2) 456 nt	105
	5.1.6	Acanthan	noeba grifini 556 nt	106
	5.1.7	Drosophi	la virilis 784 nt	108
	5.1.8	Xenopus	laevis 945 nt	111
	5.1.9	Ailurus fi		113
		7 maras n	ulgens 964 nt	113
5.2	Compa		her Algorithms	114

	5.2.2	Comparison to RNAFold	118
	5.2.3	Comparison to P-RnaPredict	120
	5.2.4	Comparison to HelixPSO	122
	5.2.5	Comparison to SARNA-Predict	125
	5.2.6	Receiver Operating Characteristic	129
	5.2.7	Discussion	131
	5.2.8	Summary	132
СНА	PTER 6	- CONCLUSION	
6.1	Summa	ary of Contributions	134
6.2	Future	Work	135
Refe	rences		137
APPI	ENDICE	S	144
APPI	ENDIX A	A – OPTIMIZATION	145
	A.0.1	Global Optimization	146
	A.0.2	Continuous Optimization	148
	A.0.3	Desecrate Optimization	148
	A.0.4	Combinatorial Optimization	149
A.1	Metahe	euristics	151
	A.1.1	Trajectory Methods	152
		A.1.1(a) Simulated Annealing	153
	A.1.2	Population-based Methods	154
		A.1.2(a) Genetic Algorithm	155
		A.1.2(b) Particle Swarm Optimization	156
APPl	ENDIX I	B - RIBONUCLEIC ACID (RNA)	158
	B 0 3	Deoxyribonucleic Acid (DNA)	158

	B.0.4	Ribonucleic Acid (RNA)	159
	B.0.5	RNA Types	161
	B.0.6	RNA and Protein Synthesis	162
B.1	Bioinfo	ormatics	163
APPE	NDIX C	C – STATISTICS	166
C.1	Definiti	on	166
C.2	GLM-U	Univariate Repated Measures	168
APPE	NDIX D	O – STATISTICAL ANALYSIS FOR HMS	169
APPE	NDIX E	Z – STATISTICAL ANALYSIS FOR HMCR	173
APPE	NDIX F	- STATISTICAL ANALYSIS FOR PAR	175
APPE	NDIX C	G – STATISTICAL ANALYSIS FOR BW	177
APPE	NDIX H	I – STATISTICAL ANALYSIS FOR AHSRNAFOLD	180
APPE	NDIX I	– STATISTICAL ANALYSIS OF NUMBER OF HM PARAMETER	182
APPE	NDIX J	- STATISTICAL ANALYSIS OF INTERVAL	185
APPE	NDIX K	X – STATISTICAL ANALYSIS OF COMMUNICATION RATE	187
APPE	NDIX L	– STATISTICAL ANALYSIS OF THERE HS VARIANTS	190
List o	f Publica	ntions	192

LIST OF TABLES

		Page
Table 3.1	An example for the permutation representation of the RNA secondary structure prediction in HS algorithm with harmony memory size=5 harmonies.	39
Table 4.1	Tested RNA organisms with their accession numbers, classes, lengths and number of base pairs in their known structures.	61
Table 4.2	Comparison of best results of HSRNAFold with <i>efn2</i> thermodynamic and with RNAeval (eval) in terms of correctly predicted base pairs (TP), incorrectly predicted base pairs (FP) and base pairs in native structure not predicted (FN).	65
Table 4.3	Comparison of best results of HSRNAFOLD with <i>efn2</i> thermodynamic and with RNAeval (eval) in terms of sensitivity (SE), specificity (SP), and F-measure (FM).	66
Table 4.4	The runtime of HSRNAFold in seconds for both efn2 and RNAeval (eval).	67
Table 4.5	RNA organisms with their known helices, all generated helices and helices after reduction.	68
Table 4.6	The nine testing RNA sequences with their accession numbers, classes and lengths.	71
Table 4.7	HSRNAFold parameters variations.	71
Table 4.8	The results of HMS variants with the corresponding ρ -value.	73
Table 4.9	The results of HMCR variants with the corresponding ρ -value.	74
Table 4.10	The results of PAR variants with the corresponding ρ -value.	76
Table 4.11	The results of BW variants with the corresponding ρ -value; the best results are shown in bold.	78
Table 4.12	Default values of AHSRNAFold.	81
Table 4.13	AHSRNAFold parameters variations.	81
Table 4.14	HSRNAFold parameters adaptation.	82
Table 4.15	CHSRNAFold parameters variations.	84
Table 4.16	The results of the number of HMs variants with the corresponding ρ -value.	86

Table 4.17	The results of communication interval variants with the corresponding ρ -value.	88
Table 4.18	The results of the communication rate variants with the corresponding ρ -value.	89
Table 4.19	A comparison between the HS variants with the corresponding ρ -value.	92
Table 5.1	Comparison of the MFE structures of CHSRNAFold(HS) and MFE structures of mfold (MF) in terms of TP, FP, FN, SE, SP and FM.	116
Table 5.2	Comparison of the highest sensitivity structures of CHSRNAFold(HS) with those of mfold (MF) in terms of TP, FP, FN, SE, SP and FM.	118
Table 5.3	Comparison of MFE structures of CHSRNAFold(HS) and MFE structures of RNAFold (RF) in terms of TP, FP, FN, SE, SP and FM.	120
Table 5.4	Comparison of the highest sensitivity structures of CHSRNAFold(HS) with the highest sensitivity structures of P-RnaPredict (GA) in terms of TP, FP, FN, SE, SP and FM.	122
Table 5.5	Comparison of the highest sensitivity structures of CHSRNAFold (HS) with those of HelixPSO (PSO) in terms of TP, FP, FN, SE, SP and FM.	124
Table 5.6	Comparison of MFE structures of CHSRNAFold (HS) and MFE of SARNA-Predict (SA) in terms of TP, FP, FN, SE, SP and FM.	126
Table 5.7	Comparison of the highest sensitivity structures of CHSRNAFold(HS) with the highest sensitivity structures of SARNA-Predict (SA) in terms of TP, FP, FN, SE, SP, FM.	128

LIST OF FIGURES

		Page
Figure 1.1	The scope of the research.	7
Figure 1.2	The Methodology.	9
Figure 2.1	Optimization procedure of the simple HS algorithm	15
Figure 2.2	Variation of PAR and BW versus generation number.	18
Figure 2.3	RNA secondary structure components: stems (helices), interior loops, hairpin loop, multi loops and bulges loops.	23
Figure 2.4	The classification of the RNA secondary structure prediction methods	28
Figure 3.1	The proposed methodology.	37
Figure 3.2	The decoding procedure of RNA secondary structure from the permutation helices.	41
Figure 3.3	Process of generating a feasible RNA secondary structure from the permutation helices.	44
Figure 3.4	Optimization procedure of AHSRNAFold algorithm.	48
Figure 3.5	Modified Variation of HMCR and PAR versus the generation number.	49
Figure 3.5(a)	HMCR variation	49
Figure 3.5(b)	PAR variation	49
Figure 3.6	A full connected cooperative HS model with multiple harmony memories.	53
Figure 3.7	A fully connected graph.	56
Figure 4.1	Plot showing the energy landscape of the best run of the Metarhizium anisopliae var. anisopliae(3) Group I intron, 23S rRNA (AF197120) sequence (394 nt).	69
Figure 4.2	Comparison between all possible helices, the helices with $FE \leq 2$ and the number of helices generated by SARNA-Predict.	70
Figure 4.3	The average best MFE recorded by HSRNAFold for test sequences when different HMS values were varied and all other parameter values were in the default value set.	73

Figure 4.4	The average best MFE recorded by HSRNAFold for test sequences when different HMCR values were varied and all other parameter values were as in the default value set.	75
Figure 4.5	The average best MFE attained by HSRNAFold for test sequences when different PAR values are varied and all other parameter values are as in the default value set.	76
Figure 4.6	The average best MFE attained by HSRNAFold for test sequences when different BW values are varied and all other parameter values are as in the default value set.	79
Figure 4.7	A comparison between HSRNAFold (fixed) and four combinations of HMCR and PAR variations: IHIP, DHDP,IHDP and DHIP.	83
Figure 4.8	A comparison of results on a particular data set with different numbers of HMs.	86
Figure 4.9	A comparison of results on a particular data set with different interval lengths.	88
Figure 4.10	A comparison of results on a particular data set with different communication rate.	90
Figure 4.11	A comparison of results on a particular data set with the different HS variants.	93
Figure 4.12	The comparison between the average best MFE of the three HS variants: HSRNAFold,AHSRNAFold and CHSRNAFold.	93
Figure 4.13	A comparison between the different HS variants performance and three RNA sequences representing short, medium and large sequences.	94
Figure 4.13(a)	Escherichia coli	94
Figure 4.13(b)	Acanthamoeba grifini	94
Figure 4.13(c)	Xenopus laevis	94
Figure 5.1	The energy landscape for a single run of the Saccharomyces cerevisiae, 5S rRNA (X67579) sequence (118 nt).	97
Figure 5.2	The correlation between the prediction accuracy and the MFE for Saccharomyces cerevisiae, 5S rRNA (X67579) sequence.	98
Figure 5.3	The visualization of known and predicted structures for the Saccharomyces cerevisiae 5SrRNA (X67579) sequence. Green for TP, blue for FP and red for FN.	99
Figure 5.4	The energy landscape of a single run of the Escherichia coli, 5S rRNA (X67579) sequence (120 nt).	100

Figure 5.5	The correlation between the prediction accuracy and the MFE for Escherichia coli, 5S rRNA (X67579) sequence.	100
Figure 5.6	The energy landscape of a single run of the Deinococcus radiodurans rRNA (AE002087) sequence (124 nt).	101
Figure 5.7	The correlation between the prediction accuracy and the MFE for Deinococcus radiodurans rRNA (AE002087) sequence.	102
Figure 5.8	The energy landscape of a single run of the Metarhizium anisopliae var. anisopliae(3) Group I intron, 23S rRNA (AF197120) sequence (394 nt).	103
Figure 5.9	The correlation between the prediction accuracy and the MFE for Metarhizium anisopliae var. anisopliae(3) Group I intron, 23S rRNA (AF197120) sequence.	104
Figure 5.10	The energy landscape of a single run of the Metarhizium anisopliae var. anisopliae(2) (AF197122) Group I intron, 23S rRNA sequence (456 nt).	105
Figure 5.11	The correlation between the prediction accuracy and the MFE for Metarhizium anisopliae var. anisopliae(2) (AF197122) Group I intron, 23S rRNA sequence.	106
Figure 5.12	The energy landscape of a single run of the Acanthamoeba grifini (U02540) Group I intron, 16S rRNA sequence (556 nt).	107
Figure 5.13	The correlation between the prediction accuracy and the MFE for Acanthamoeba grifini (U02540) Group I intron, 16S rRNA sequence.	108
Figure 5.14	The circular representation of known and predicted structures for the Acanthamoeba grifini (U02540) Group I intron, 16S rRNA. Green for TP, blue for FP and red for FN.	109
Figure 5.15	The energy landscape of a single run of the Drosophila virilis 16S rRNA (X05914) sequence (784 nt).	110
Figure 5.16	The correlation between the prediction accuracy and the MFE for Drosophila virilis 16S rRNA (X05914) sequence.	110
Figure 5.17	The energy landscape of a single run of the Xenopus laevis 16S rRNA (M27605) sequence (945 nt).	111
Figure 5.18	The correlation between the prediction accuracy and the MFE for Xenopus laevis 16S rRNA (M27605) sequence.	112
Figure 5.19	The circular representation of known and predicted structures for Xenopus laevis 16S rRNA (M27605) sequence. Green for TP, blue for FP and red for FN.	113
Figure 5.20	The energy landscape of a single run of the Ailurus fulgens Ailurus fulgens (Y08511) 16S rRNA sequence (964 nt).	114

Figure 5.21	The correlation between the prediction accuracy and the MFE for Ailurus fulgens Ailurus fulgens (Y08511) 16S rRNA sequence.	115
Figure 5.22	The comparison plot of the MFE structures sensitivity of CHSRNAFold and mfold.	117
Figure 5.23	The highest sensitivity comparison plot of CHSRNAFold and mfold.	119
Figure 5.24	The MFE comparison plot of CHSRNAFold and RNAFold.	121
Figure 5.25	The highest sensitivity comparison plot of CHSRNAFold and P-RnaPredict.	123
Figure 5.26	The highest sensitivity structures comparison plot of CHSRNAFold and HelixPSO.	125
Figure 5.27	The MFE structures comparison plot of CHSRNAFold and SARNA-Predict.	127
Figure 5.28	The highest sensitivity structures comparison plot of CHSRNAFold and SARNA-Predict.	129
Figure 5.29	The ROC plot displays both average sensitivity and specificity of all the sequences for CHSRNAFold, mfold, RNAFold, RnaPredict and HelixPSO.	130
Figure 5.30	The ROC plot displays both average sensitivity and specificity of all the sequences for CHSRNAFold and SARNA-Predict.	131
Figure A.1	The taxonomy of optimization techniques.	147
Figure B.1	The components of DNA molecule versus RNA molecule.	159
Figure B.2	The components of the RNA molecule.	160
Figure B.3	RNA and protein synthesis process.	164
Figure B.4	The bioinformatics.	164
Figure B.5	The bioinformatics classification.	165

LIST OF ALGORITHMS

3.1	HSRNAFold algorithm	40
	Standard helix generation algorithm	42
3.3	The modified helix generation algorithm	43
3.4	The modified decoding algorithm	44
3.5	AHSRNAFold algorithm	50
3.6	Cooperative HS model	52
	The exchange pseudocode	55
3.8	CHSRNAFold algorithm	58
A.1	Simulated Annealing	154
	Genetic algorithm	
A.3	Particle Swarm Optimization	157

LIST OF ABBREVIATIONS

Ant Colony Optimization. **ACO** AHSRNAFold Adaptive Harmony Search for RNA Folding. \mathbf{BW} Band width. **CHSRNAFold** Cooperative Harmony Search for RNA Folding. CI Computational Intelligence. Combinatorial Optimization. CO **COP** combinatorial Optimization Problem. **DHDP** Decreasing HMCR and PAR. Decreasing HMCR and Increasing PAR. **DHIP DNA** Deoxyribonucleic Acid. DP Dynamic Programming. EA Evolutionary Algorithms. EC Evolutionary Computation. **Fixed** Fixed values of HMCR and PAR. FM F-measure. False Negative base pairs. FN FP False Positive base pairs.

GA

Genetic Algorithm.

GHS Global-best Harmony Search.

GO Global Optimization.

HM Harmony Memory.

HMCR Harmony Memory Consideration Rate.

HMS Harmony Memory Size.

HS Harmony Search.

HSRNAFold Harmony Search for RNA Folding.

HSV Harmony Search Version.

IHDP Increasing HMCR and Decreasing PAR.

IHIP Increasing HMCR and PAR.

IHS Improved Harmony Search.

INN Individual Nearest Neighbour.

INN-HB Individual Nearest Neighbour Hydrogen Bonding Model.

MFE Minimum Free Energy.

mRNA Messenger Ribonucleic Acid.

NMR Nuclear Magnetic Resonance.

nt nucleotides.

PAR Pitch Adjustment Rate.

PSO Particle Swarm Optimizer.

RNA Ribonucleic Acid.

ROC Receiver Operating Characteristic.

rRNA Ribosomal Ribonucleic Acid.

RSR Random Selection Rate.

SA Simulated Annealing.

SE Sensitivity.

SetPSO Set Particle Swarm Optimizer.

SGHS A Self-Adaptive Global Best Harmony Search Algorithm for continuous optimization problems.

snRNA Small Nuclear RNA.

SP Specificity.

TP True Positive base pairs.

tRNA Transfer Ribonucleic Acid.

LIST OF SYMBOLS

$\triangle G$ Delta MFE

- μ The overall HM
- α Statistical α value
- ρ Statistical ρ value

MODEL MUDAH SUAI DAN KERJASAMA GELINTARAN HARMONI BAGI RAMALAN STRUKTUR SEKUNDER RNA

ABSTRAK

Penentuan fungsi molekul RNA amat bergantung kepada struktur sekundernya. Kaedah fizikal yang sedia ada untuk penentuan struktur sekunder adalah mahal dan memakan masa. Beberapa algoritma telah dicadangkan untuk peramalan struktur sekunder RNA, termasuk pengaturcaraan dinamik dan algoritma metaheuristik. Gelintaran harmoni (GH) merupakan suatu algoritma metaheuristik baru yang berjaya dalam penyelesaian berbagai jenis masalah pengoptimuman. Penyelidikan ini mengusulkan tiga varian baru algoritma GH untuk menyelesaikan masalah peramalan struktur sekunder RNA. Varian pertama yang dikenali sebagai HSR-NAFold adalah berdasarkan GH asas dan merupakan algoritma GH pertama untuk masalah ramalan struktur sekunder RNA. Varian kedua, AHSRNAFold, memperbaiki HSRNAFold dengan kawalan parameter mudah suai. Varian ketiga pula memperbaiki HSRNAFold dengan menggunakan model GH kerjasama dengan berbilang ingatan harmoni, dikenali sebagai CHSRNAFold. Kelakuan varian-varian GH yang baru itu dikaji dan impak penalaan parameter yang berlainan bagi varian-varian ini dinilai. Eksperimen dijalankan ke atas 20 individu dengan struktur yang diketahui dari empat kelas RNA. Kejituan peramalan ditentusahkan dengan menggunakan struktur natif dan algoritma terkini yang lain. Hasil penyelidikan ini menunjukkan bahawa CHSRNAFold memberikan keputusan yang lebih baik berbanding dengan keputusan beberapa algoritma terkini dari segi kejituan peramalan.

ADAPTIVE AND COOPERATIVE HARMONY SEARCH MODELS FOR RNA SECONDARY STRUCTURE PREDICTION

ABSTRACT

Determining the function of RNA molecules relies heavily on its secondary structure. The current physical methods for secondary structure determination are expensive and time consuming. Several algorithms have been proposed for the RNA secondary structure prediction, including dynamic programming and metaheuristic algorithms. Harmony search (HS) is a new metaheuristic algorithm which succeeded in solving many different types of optimization problems. This research proposes three new variants of HS algorithm to address the RNA secondary structure prediction problem. The first variant is called HSRNAFold as a first application of HS for RNA secondary structure prediction. The second variant, AHSRNAFold, improves HSRNAFold by using adaptive parameter control. The third variant, CHSRNAFold, improves HSRNAFold by using a cooperative multiple harmony memories model. The behavior of the new HS variants is investigated and the impact of tuning the different parameters of these variants is evaluated. The experiments were conducted on 20 individuals with known structures from four RNA classes. The prediction accuracy was verified with native structures and other state-of-the-art algorithms. The results demonstrate that CHSRNAFold outperformed several state-of-the-art algorithms in terms of prediction accuracy.

CHAPTER 1

INTRODUCTION

RNA is a nucleic acid which consists of a long linear polymer of nucleotide units found in the nucleus. RNA is similar to DNA, but usually it is single stranded instead of double-stranded, containing ribose rather than deoxyribose bases. It has uracil (U) in place of thymine (T).

The discovered biological functions of RNA have increased in recent times. The scope of understanding has expanded and RNA is no longer viewed as only a passive messenger of genetic information from DNA to proteins manufacturers as had been thought before. These new discoveries have motivated RNA research in many aspects.

RNA has been found to play important roles in all molecular biology such as carrying genetic information (messenger RNA), interpreting the code (ribosomal RNA) and transferring genetic code (transfer RNA). It also performs different functions including catalyzing chemical reactions (Doudna and Cech, 2002; Hansen et al., 2002), directing site specific modification of RNA nucleotides, controlling gene expression, modulating protein expression and serving in protein localization (Bachellerie et al., 2002; Meister and Tuschl, 2004). These functions of RNA molecules determine many diseases caused by RNA viruses. Understanding of the biological functions of an RNA molecule is fundamentally based on identifying its 3D structure (Tsang and Wiese, 2007a; Neethling and Engelbrecht, 2006). The primary structure of RNA is the easiest structure to be determined in the laboratory using gene sequencing techniques. It does not contain additional information about the functional structure. On the other hand, the tertiary structures are much more difficult to model where the secondary structure bonds

are stronger and can be formed faster than that of the tertiary structure (Zuker et al., 1999). Therefore, the computational approaches used to predict the structure of RNA have paid more attention to the secondary structure.

Since RNA structure and function are closely related, it is important to understand the common structure of homologous RNAs in order to discover their functional signatures. However, due to the exponential number of possible structures, RNA structure prediction is a complex problem. As such, it is still an open problem in bioinformatics.

1.1 Problem statement and Motivations

Physical methods used to determine the RNA secondary structure such as X-ray diffraction and nuclear magnetic resonance (NMR) spectroscopy are difficult, time consuming and expensive. Therefore, computational approaches to predict the secondary structure of RNA molecule can be considered as an appropriate alternative (Tsang, 2007).

RNA secondary structure prediction is not a trivial problem. It has been estimated that the number of secondary structures modeled from the input of n nucleotides is greater than 1.8^n (Doshi et al., 2004). For example, Saccharomyces cerevisiae (X67579) 5S rRNA with 118 nucleotides in length has an estimated 1.3×10^{30} secondary structure models whereas a larger RNA such as the Sulfolobus acidocaldarius (D14876) 16S rRNA, with 1493 nucleotides, has an estimated total of 1.3×10^{381} possible secondary structure models.

Two different computational approaches are currently in use to address the RNA secondary structure prediction problem. The first approach is called the comparative sequence analysis approach (Gardner and Giegerich, 2004; Gotoh, 1999). It is an iterative process performed on a set of homologous related RNA sequences. Briefly, sequence alignment works on the RNA

sequences similarities. This alignment is achieved by adding and removing gap characters (Deschenes, 2005). The purpose is to correlate sequence and function across genomes. The second approach is the single sequence approach, which predicts the secondary structure by searching for the minimum free energy.

Most of the methods were developed based on free energy minimization either by applying dynamic programming algorithms (DP) or metaheuristics. Based on free energy minimization of a single RNA sequence, dynamic programming algorithms have been studied since the early 1970s. Mathews (2006b) provided a review of the revolutions which occurred in the development of a number of these algorithms.

Nussinov et al. (1978) predicted the RNA secondary structure using the DP method by maximizing the number of base pairs. In 1980, they further adapted their original method to enhance the results using a simple nearest-neighbor energy model (Nussinov and Jacobson, 1980). Zuker and Stiegler (1981) proposed a slightly refined DP approach which models the nearest neighbor energy interactions and directly incorporated stacking into the prediction. Later, Zuker (2003) proposed the DP algorithm, mfold. It is still a popular algorithm used to find the minimum free energy (MFE) pseudoknot-free secondary structure of an RNA molecule. Furthermore, it has become the benchmark for predicting the RNA secondary structure. mfold uses a complex thermodynamic model to evaluate the free energy of the structures by seeking the pseudoknot-free secondary structure with MFE (Zuker, 1994, 2003). Later, RNAFold from the *ViennaRNA* (Hofacker et al., 1994) package was proposed as a dynamic programming algorithm to predict the RNA secondary structure through energy minimization.

Dynamic programming algorithm, as a mathematical technique, can hit the global optima in solving small problems. Nevertheless, in real world problems, there are some drawbacks. For example, when the number of variables increases, the number of evaluations increases

exponentially due to recursive nature of dynamic programming. For RNA secondary structure prediction, the huge number of structure alternatives makes it difficult to determine the most correct one (Tsang, 2007).

In another development, many metaheuristics algorithms were proposed such as genetic algorithms (GAs), simulated annealing (SA) and particle swarm optimization (PSO). GAs was shown to achieve higher base pairs prediction rates than DP (Gultyaev et al., 1998). The most recent GAs studies in this area are RnaPredict and its parallel version (P-RnaPredict) which were proposed by Wiese and his students (Wiese et al., 2007; Wiese and Hendriks, 2006). The results of both algorithms showed that their quality is comparable to mfold. SARNA-Predict which is an SA algorithm was introduced by Tsang and Wiese (2007a,b). It attempted to predict the RNA secondary structures with a low free energy. SARNA-Predict showed good results, with high number of correctly predicted base pairs, in comparison to known native structures and to other algorithms in the literature. Recently, two versions of PSO, setPSO and HelixPSO, were also proposed by Neethling and Engelbrecht (2006) and Geis and Middendorf (2007) respectively. Both algorithms were used to find secondary structures with low free energy.

The main drawback of local based metaheuristics approaches like SA is that they may get stuck in the local optimal solution. In addition, there is no guarantee that the value of the objective function at any local optimum is close to the optimum value (Aarts and Lenstra, 1997). On the other hand, the population-based metaheuristics approaches such as GA, ant colony and PSO have their drawbacks of premature convergence and stagnation (Qin et al., 2006).

1.2 Harmony Search Algorithm

Harmony Search (HS) algorithm is an optimization technique developed by Geem (Geem et al., 2001). HS mimics the musicians' improvisation process.

Researchers (Alatas, 2010; Lee and Geem, 2004; Geem et al., 2001; Mahdavi et al., 2007) summarized the features of HS over the other traditional optimization techniques: i) HS imposes less mathematical requirements, and as such it can be easily used for various types of engineering problems; ii) it does not require initial value settings of the decision variables, and thus, it may escape from local optima; iii) derivative information is not necessary due to stochastic random searches which HS uses; iv) HS can work with both discrete and continuous optimization problems; v) it can overcome the drawback of building block theory of GA by taking into account the relationship between the decision variables using its ensemble operation; and vi) HS algorithm generates a new vector by considering all of the existing vectors, rather than considering only two parents as in GA.

These features increase the flexibility of HS algorithm in solving a wide variety of optimization problems in several fields. Ingram and Zhang (2009) provided an overview of applications and developments using HS algorithm. These applications include continuous engineering optimization, vehicle routing, combined heat and power economic dispatch, water pump switching problem, optimal scheduling of multiple dam system and transport energy modeling (Fesanghary et al., 2009, 2008; Ayvaz, 2007; Lee and Geem, 2004, 2005; Mahdavi et al., 2007; Mohsen et al., 2008, 2009a,b; Saka, 2009; VASEBI et al., 2007; Ceylan et al., 2008; dos Santos Coelho and Mariani, 2009; Jaberipour and Khorram, 2010; Kaveh and Talatahari, 2009; Kaveh and Abadi, 2010; Mun and Geem, 2009a,b; Pan et al., 2010c; dos Santos Coelho and de Andrade Bernert, 2009; Zou et al., 2010).

Harmony search has three parameters which contribute interactively to the creation of new solution. The interaction between various components is the important factor to consider for the success of HS algorithm over other algorithms. As such, this interaction may guarantee a good balance between the intensification and diversification. Such a balance may prevent premature convergence and overcome the stagnation. For algorithm parameters, some evidences are available to suggest that HS is sensitive to chosen parameters (Mahdavi et al., 2007). This means that these parameters may need to be fine-tuned to obtain quality solutions. Furthermore, a group of multiple harmony memories can be used in parallel modeling. This parallel model may increase both the efficiency and effectiveness of the algorithm (Yang, 2009).

1.3 Objectives

The current algorithms used in RNA secondary structure prediction have some limitations and drawbacks. The aim of this thesis is not only to develop efficient HS variants for RNA secondary structure prediction, but also to show that the proposed variants are able to overcome the state-of-the-art algorithms in terms of performance.

The primary objectives of this thesis are summarized as follows:

- To adapt HS algorithm to address RNA secondary structure prediction problem.
 - To speed up the prediction process by refining the existing helix generation algorithm.
 - To evaluate the effect of using different settings for HS parameters such as harmony memory size (HMS), harmony memory consideration rate (HMCR), pitch adjustment rate (PAR) and bandwidth (BW) on the solution quality and the convergence behavior.

- To further enhance the accuracy of prediction and performance of HS for RNA secondary structure prediction problem by
 - Applying adaptive parameters for HS parameters.
 - Using a new cooperative model with multiple harmony memories.

1.4 Scope and Limitations

This thesis focuses on solving the RNA secondary structure prediction problem. Based on the assumption that the correct structure is a low energy structure, the RNA folding is subject to the laws of thermodynamics (Deschenes, 2005). The stability of the secondary structure depends on the amount of free energy released to form the base pairs. Therefore, the more negative the free energy of a structure is, the more stable a particular sequence is formed. This structure is called the MFE secondary structure (Layton and Bundschuh, 2005).

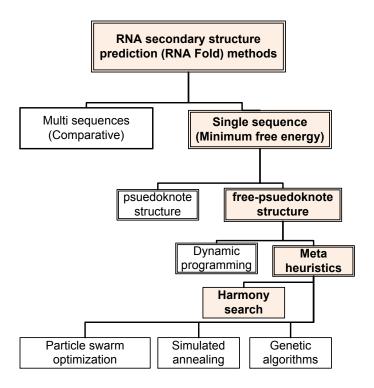


Figure 1.1: The scope of the research.

In a case where only a single sequence of a given RNA molecule is known or the number of available sequences with high similarity is low, the *ab initio* methods are used to perform RNA secondary structure prediction as an energy minimization problem. These methods are either dynamic programming or metaheuristics. This research proposes new metaheuristic methods based on HS algorithm to enhance the accuracy of the prediction. As shown in Figure 1.1, the scope of study focuses on the prediction of pseudoknots-free RNA secondary structure. The prediction of the pseudoknots RNA secondary structure does not fall within the scope of this research due to the following reasons: i) the computational complexity (Hendriks, 2005); ii) the inability of the adopted thermodynamic models to deal with pseudoknot motifs (Neethling, 2008); and iii) infrequent occurrence of pseudoknots in nature (Deschenes, 2005). In the future, if corresponding thermodynamic models support the calculation of pseudoknot energy contributions, it will be easy to extend the proposed variants to enable prediction of pseudoknots.

1.5 Research Approach

This research work is divided into three processes: preprediction, prediction and postprediction as shown in Figure 1.2.

In the preprediction process, the set of all feasible helices are generated with the calculation of the free energy for each helix using the particular thermodynamic model. In the prediction process, three variants are proposed based on HS for RNA secondary structure prediction. The first proposal is to predict the structure using basic HS without modification. The second proposal is an enhanced version of first proposal by using adaptive parameters control. Two parameters- harmony memory consideration rate (HMCR) and pitch adjustment rate (PAR)-are affected by this adaptation. The third proposal is the new cooperative model of the basic HS with multiple HMs. In all the proposals, the same mechanism is used for RNA secondary structure encoding, decoding, the thermodynamic models and the harmony representation.

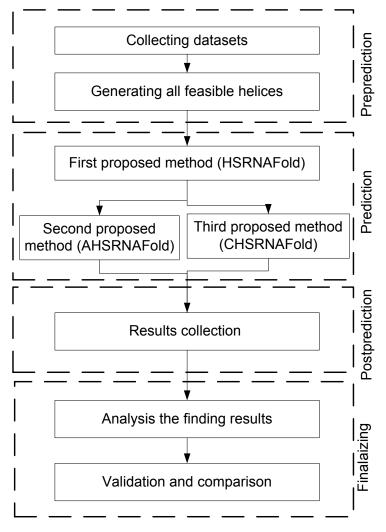


Figure 1.2: The Methodology.

The third process includes the collection of the results of each proposal for analysis and discussion. In this step, the best predicted secondary structure is also generated.

Comprehensive study and discussion are performed on a variety of RNA classes (5S rRNA, Group I intron 16S rRNA, Group I intron 23S rRNA and 16S rRNA) to study the performance of the three proposed variants of HS and the accuracy of prediction. For the performance, the convergence behavior of the three variants is examined and various parameters' setting are investigated. In terms of prediction accuracy, an evaluation of the performance of the new variants is performed via the comparison to other RNA secondary structure prediction algorithms such as mfold (Zuker, 1994, 2003), RNAFold (Hofacker et al., 1994), RnaPredict

and P-RnaPredict (Wiese et al., 2007; Wiese and Hendriks, 2006; Hendriks, 2005; Deschenes, 2005), setPSO (Neethling and Engelbrecht, 2006), HilexPSO (Geis and Middendorf, 2007) and SARNA-Predict (Tsang and Wiese, 2007b,a; Tsang, 2007)).

1.6 List of Contributions

This research investigates ideas in the direction of improving HS performance for RNA secondary structure prediction. There seem to be several exciting research issues connected with parameter control as well as the cooperative model of HS, which are investigated in this research. The main contributions include the following:

- A new variant of HS algorithm called HSRNAFold as the first application of HS for RNA secondary structure prediction (Geem, 2010).
- An improved variant of HSRNAFold based on adaptive parameters called AHSRNAFold.
- A new variant called CHSRNAFold algorithm which differed from the original HSRNAFold
 by operating on cooperative multiple HMs model to enhance both algorithm performance
 and accuracy of prediction.
- A comprehensive study of the influence of the main parameters of HSRNAFold and its subsequent variants which may affect the algorithm's performance when used in a real world optimization problem.

A comparative study of the three proposed HS variants was performed amongst themselves and against the state of the art algorithms for RNA secondary structure prediction, and then to the native structures.

1.7 Thesis Outline and Organization

The organization of the remaining chapters of this thesis is as follows:

- Chapter 2 provides an overview of the original HS algorithm.
- Chapter 3 introduces the RNA structures, RNA secondary structure prediction problem and related work, and outlines the shortcomings of existing methods.
- Chapter 4 presents the three variants of HS algorithm: HSRNAFold, AHSRNAFold and CHSRNAFold. In addition, the major modified and enhanced components of the three methods are also presented.
- Chapter 5 presents the experimental setup and a comprehensive investigation of the performance of the three variants of HS with different parameter setting.
- Chapter 6 presents the experimental results and evaluates the validity of the proposed algorithm.
- Chapter 7 presents the concluding remarks, suggestions and some possible directions for future research.

CHAPTER 2

LITERATURE REVIEW

This chapter provides a description on the HS fundamentals. Section 2.1 gives a brief overview of the HS procedures. Section 2.1.1 describes the HS procedure. Section 2.1.2 reviews the main HS optimization steps. Section 2.1.3 and 2.1.4 provide a summary of the related work that has been done in adaptive parameters and multiple harmony memories.

A review of the related literature pertaining to RNA secondary structure prediction will be provided as well. Section 2.2 gives a quick review of the RNA secondary structure. In Section 2.3, Two physical methods for determining RNA structure are presented. Section 2.4 provides information on the related work of the two major computational methods for RNA secondary structure prediction: multiple sequences and single sequence methods. Finally, Section 2.5 summarizes and concludes the chapter.

2.1 Harmony Search Algorithm

Harmony search was initiated by Geem and his colleagues in 2001 (Geem et al., 2001) as a relatively new metaheuristic for hard combinatorial optimization problems (for more details about metaheuristic and combinatorial optimization see Appendix A). Harmony search is a stochastic search technique based on the mechanism of improvisation process to find fantastic harmony. It has received a great deal of attention regarding its potential as an optimization technique for solving discrete and continuous optimization problems (for more details about discrete and continuous optimization see Appendix A).

2.1.1 Fundamental Procedures of HS

In HS, harmony parameters are usually used to create new harmony in each improvisation. The main role of these parameters is to direct the search toward the most favorable areas of the search space. These parameters are:

- Harmony memory size (HMS) representing the total number of harmonies in the HM.
- Harmony memory consideration rate (HMCR) which represents the probability of picking up values from HM to the variables in the solution vector.
- Random selection rate (RSR) representing the probability of randomly chosen feasible values from the range of all possible values to the variables in the solution vector, formally, RSR = 1 HMCR.
- Pitch adjusting rate (PAR) representing the probability of further adjusting the pitch with neighboring pitches.
- Number of improvisations (NI) representing the number of iterations to be used during the solution process, or stopping criterion.

To explain the fundamental procedures of HS, consider a harmony memory that consists of N harmonies representing potential solutions to a problem. In HS, a harmony in harmony memory is represented by a string S of length n as follows: $S = S_1, S_2, \ldots, S_j, \ldots, S_n$.

The string S is regarded as a harmony that consists of n musical instruments. The character S_j is a musical instrument at the j^{th} locus, and the different values of a musical instrument are called notes. The harmony is a potential solution to a problem corresponding to a string S called the solution vector. In minimization problems, the string with a smaller objective function value has a higher fitness.

HS starts with an initial HM of *n* harmonies generated randomly. Each harmony in the harmony memory represents a potential solution of the problem under consideration. Each harmony in the harmony memory is evaluated using an objective function. The harmonies evolve through successive iterations, called improvisations. During each improvisation, a new harmony is created through harmony operators. After that, the harmony memory is updated if the new harmony is better than its worst one. The procedure continues until the termination condition is satisfied. When the termination condition is satisfied, the best harmony obtained is regarded as an optimal or approximate optimal solution to the problem.

When applying HS to solve particular optimization problems, further detailed considerations are required: i) representation for potential solutions, ii) a way to create an initial harmony memory, iii) an evaluation process in terms of their objective function, iv) harmony parameters, v) constraint-handling techniques, vi) tuning for various parameters in HS such as HMS, HMCR and PAR and vii) termination conditions.

2.1.2 HS Optimization Steps

Figure 2.1 shows the optimization steps of HS which is presented in detail in the next subsections.

2.1.2(a) Initialize the Problem and Algorithm Parameters

Mathematically, the general form of optimization problem can be specified as follows:

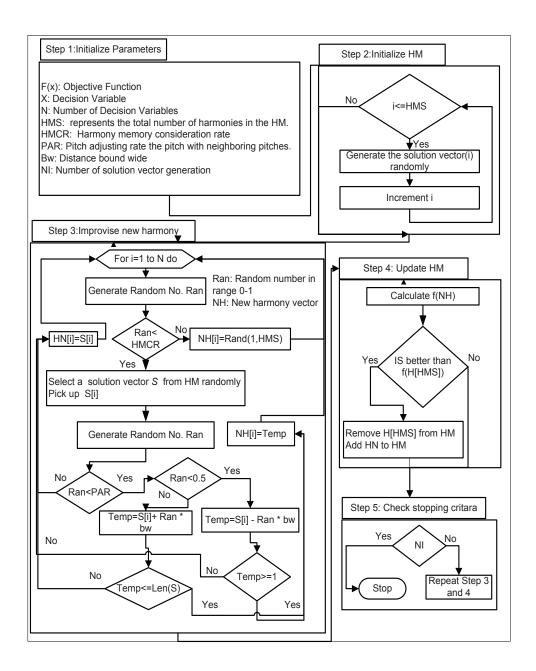


Figure 2.1: Optimization procedure of the simple HS algorithm (Mahdavi et al., 2007).

$$\begin{cases}
Minimize f(x) \\
Subject to g(x) > 0, x = \{x_1, x_2, \dots, x_n\} \\
h(x) = 0
\end{cases}$$
(2.1)

Where f(x) is the objective function; g(x) and h(x) are the inequality and equality con-

straint functions respectively; x is the set of each decision variable x_i ; and n is the number of decision variables (music instruments). HS algorithm parameters that are required to solve the optimization problem (i.e., HMS, HMCR, PAR, BW and NI) are also specified in this step. These parameters are used to improve the solution vector.

2.1.2(b) Initialize the Harmony Memory

Initialize the HM matrix($N \times HMS$) where N is the number of decision variables and M is HMS. Then fill the HM randomly by generating the feasible solution vectors. Formally, HM and the corresponding fitness function values are shown as follows:

$$HM = \begin{bmatrix} x_1^1 & x_2^1 & \dots & x_{N-1}^1 & x_N^1 \\ x_1^2 & x_2^2 & \dots & x_{N-1}^2 & x_N^2 \\ \vdots & \dots & \dots & \dots \\ x_1^{HMS-1} & x_2^{HMS-1} & \dots & x_{N-1}^{HMS-1} & x_N^{HMS-1} \\ x_1^{HMS} & x_2^{HMS} & \dots & x_{N-1}^{HMS} & x_N^{HMS} \end{bmatrix} \Rightarrow f(x^1)$$

$$\Rightarrow f(x^2)$$

$$\Rightarrow \vdots$$

$$\Rightarrow f(x^{HMS-1})$$

$$\Rightarrow f(x^{HMS-1})$$

$$\Rightarrow f(x^{HMS})$$

Where each $x' = (x_1^1 x_2^1 \dots x_N^1)$ and $f(x^1)$ represents a feasible solution vector and it's corresponding objective function respectively.

2.1.2(c) Improvise a New Harmony

A new harmony vector $x' = (x'_1 x'_2 \dots x'_N)$, is generated based on three parameters: memory consideration, pitch adjustment and random selection as follows (Geem et al., 2001):

i) for each component $x_i^{'}$, pick up the corresponding component of $x_i^{'}$ randomly from any of

the values in the specified HM range $(x_i^{'1} - x_i^{'HMS})$ with the probability of P_{hmcr} .

$$x_{i}^{'} \leftarrow \begin{cases} x_{i}^{'} \in \{x^{'} = (x_{i}^{1}, x_{i}^{2}, \dots : x_{i}^{HMS}\} & with \ probability \ HMCR \\ x_{i}^{'} \in Xi & with \ probability \ (1 - HMCR) \end{cases}$$

$$(2.3)$$

- ii) the rest of the components of x_i' are picked randomly from the range of allowed values with the probability of $1 P_{hmcr}$. For example, HMCR of 0.95 indicates that the probability of HS algorithm to choose the decision variable values from historically stored values in the HM is 95% and the probability of choosing a new random value from the allowed range is (100-95)%.
- iii) change $x_i^{'}$ with the probability of P_{par} . The pitch adjustment is applied only if the value is chosen from the HM.

Pitch adjusting decision for
$$x_{i}^{'} \leftarrow \begin{cases} Yes \ with \ probability \ PAR, \\ No \ with \ probability \ (1-PAR). \end{cases}$$
 (2.4)

If the pitch adjustment decision for $x_i^{'}$ is yes, then small amount (bw) of changes takes place for pitch adjustments:

$$x_{i}^{'} \leftarrow x_{i}^{'} \pm bw * rand(). \tag{2.5}$$

2.1.2(d) Harmony Memory Update

Evaluate the new harmony $x' = (x'_1 x'_2 \dots x'_N)$ by calculating it's objective function. If the value of its objective function is better than that of the objective function of the worst harmony in the

HM, the new harmony is included in the HM and the existing worst harmony is excluded from the HM. Subsequently, the vectors are sorted out based-on their objective function values.

2.1.2(e) Termination Criterion Check

Stop the search process if a maximum number of iterations (number of improvisations) is reached. Otherwise, repeat steps three and four.

2.1.3 Adaptive Parameters

The study of the adaptive parameters in HS started in 2007 by Mahdavi et al. (2007) with his algorithm called Improved Harmony Search (IHS). Many subsequent studies were inspired by IHS. However, some of these studies disagreed with IHS. In IHS the fine-tuning was done for two parameters, PAR and BW. These two parameters control the convergence rate of HS. Figure 2.2 shows the fine-tuning that has been applied dynamically by increasing and decreasing the values of PAR and BW respectively. They claimed that IHS overcomes the drawbacks of using fixed values of PAR and BW in the simple HS algorithm. Formally, PAR and BW are updated dynamically according to Equation 2.6 and Equation 2.7 respectively.

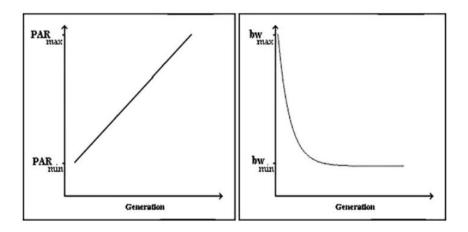


Figure 2.2: Variation of PAR and bw versus generation number (Mahdavi et al., 2007).

$$PAR(g) = PAR_{min} + \frac{PAR_{max} - PAR_{min}}{NI} \times g$$
 (2.6)

$$BW(g) = BW_{max} \exp\left(\frac{\ln \frac{BW_{min}}{BW_{max}}}{NI} \times g\right), \tag{2.7}$$

where PAR(g) and BW(g) are the pitch adjustment rate and the distance bandwidth in generation g respectively; NI is the maximum number of iterations, and g is the current iteration; PAR_{min} and PAR_{max} are the minimum and the maximum pitch adjustment rate respectively; BW_{min} and BW_{max} are the minimum and the maximum bandwidth respectively.

IHS algorithm has critical drawbacks (Wang and Huang, 2010). For instance, there is difficultly in setting suitable values of BW_{min} and BW_{max} and, on the other hand, PAR should be decreased with search time to limit perturbation. Surprisingly, Omran and Mahdavi (2008) in their subsequent research claimed that they achieved better results, in spite of giving PAR a small constant value.

Later, Omran and Mahdavi (2008) developed another version of HS called the Global-best Harmony Search (GHS). GHS is different from the simple HS in the improvisation step by modifying the pitch adjustment rule. The idea was inspired from swarm intelligence to enhance the performance of HS. To improvise new harmony, the pitch adjustment of the GHS was modified such that a new harmony is affected by the best harmony in the harmony memory. GHS simplifies the pitch adjustment step and BW is not used anymore. Formally, the rule to

adjust the pitch is given in Equation 2.8 as follows:

$$X_{new}(j) = X_B(k), j = 1, 2, ..., n \text{ and } k = Rand(1, n),$$
 (2.8)

where X_{new} is the new harmony, X_B is the best harmony in harmony memory and k is a random integer between 1 and n. According to Omran and Mahdavi (2008), this modification allows the GHS algorithm to work more efficiently on both continuous and discrete problems.

The GHS was also criticized by Wang and Huang (2010). They listed a number of disadvantages of GHS. It suffered from premature convergence. Moreover, there are some obvious mistakes in the GHS and so the reliability of the numerical results is decreased.

Later, dos Santos Coelho and Mariani (2009) proposed a modified version of HS. They inspired the concept from Mahdavi (Mahdavi et al., 2007) for using variable PAR with small changes to the Equation 2.2. The modification is the inclusion of the grade of the solution vectors into Equation 2.6. The grade is updated according to the following expression:

$$Grade = \frac{F_{max}(g) - mean(F)}{F_{max}(g) - F_{min}(g)},$$
(2.9)

where $F_{max}(g)$ and $F_{min}(g)$ are the maximum and minimum objective function values in generation g, respectively; mean (F) is the mean of the objective function value of the harmony memory. The new PAR rule shown in Equation 2.10 as follows:

$$PAR(g) = PAR_{min} + \frac{PAR_{max} - PAR_{min}}{NI} \times g \times grade$$
 (2.10)

Then, Pan et al. (2010c) proposed a variant of HS called A Self-Adaptive Global Best

Harmony Search Algorithm for continuous optimization problems (SGHS). Unlike GHS, in SGHS the value of the decision variable $X_B(j)$ in X_B is assigned to $X_{new}(j)$, while in GHS, $X_{new}(j)$ is determined by selecting one of the decision variables of X_B randomly. According to Equation 2.11, PAR is updated as follows:

$$X_{new}(j) = X_B(j), j = 1, 2, ..., n$$
 (2.11)

where X_{new} is the new harmony, X_B is the best harmony in harmony memory and j is an integer between 1 and n which refers to the current location in the corresponding harmony. The results showed that the SGHS algorithm outperforms the existing HS, IHS and GHS algorithms.

2.1.4 Multiple Harmony Memories Models

According to Yang (2009), since HS algorithm is a population-based metaheuristic, a group of multiple harmonies can be used in parallel. Proper parallelism could result in a better performance with higher efficiency. Conducting a balance between intensification and diversification could also be achieved with the use of parallelism and elitism.

Pan et al. (2010b,a) proposed two variants. The first one is called referred to them as a local-best harmony search algorithm with dynamic subpopulations for solving continuous optimization problems; and the other is called a local-best harmony search algorithm with dynamic sub-harmony memories for lot-streaming flow shop scheduling problem. These two are the only methods that take advantage of multiple harmony memories to improve the HS performance. Numerical experiments showed that these techniques overcome the existing HS, IHS, GHS, and MHS algorithms. The methods have some limitations due to the incomplete model of multiple harmony memories and the effects of the parameters of the multiple harmony memories model was neglected.

2.2 RNA Secondary Structure

The linear sequence of RNA molecule consists of a single stranded sequence of four nucleotides. This linear sequence is the primary structure of RNA molecule. The RNA strand has the ability to fold back upon itself. During the folding process, the hydrogen bonds which lie between different nucleotides form base pairs. These hydrogen bonds, which occur mostly between G and C or A and U, are called the Watson-Crick base pairs and the bond between G and U is called the wobble base pair. These base pairs- GC, AU, and GU, and their mirrors, CG, UA, and UG- are called the canonical base pairs.

Definition 2.1. Given a single stranded RNA sequence of length $L, x = (x_1, x_2, ..., x_L)$, with $x_i \in \{A, C, G, U\}$ for all i, the RNA secondary structure for x is defined as a set P of ordered base pairs, written as (i, j), with $1 \le i \le j \le L$, which satisfy the following constraints (Wiese and Hendriks, 2006; Mathews, 2006a; Zuker, 1994):

- *i)* for (*i,j*), it must be a canonical base pair;
- ii) each base pair cannot share more than one base (nucleotide);
- iii) pairing bases must be at least three bases apart i j > 3; and
- iv) two base pairs must not cross, i.e.: $\{i,j\} \cap \{i',j'\} = \Phi$ or for all (i,j), (i',j') either i < i' < j' < j or i' < i < j' holds.

The RNA secondary structure has a number of elements including stacked base pairs which form helices, hairpin loops, internal loops, bulges, multi-branched loops and external bases.

• Hairpin loop is a group of nucleotides which are enclosed by a helix but not canonically paired. Formally, in a given secondary structure, the tuple (i, j) defines a hairpin loop if