# ANTIHYPERTENSIVE PROPERTIES OF STANDARDISED ORTHOSIPHON STAMINEUS BENTH. LEAVES EXTRACTS AND ITS NANO LIPOSOMES IN SPONTANEOUS HYPERTENSIVE RATS

by

# ARMAGHAN SHAFAEI DARESTANI

Thesis submitted in fulfilment of the requirements for the degree of Doctor of Philosophy

December 2016

### ACKNOWLEDGEMENT

"In order to succeed, your desire for success should be greater than your fear of failure." - Bill Cosby. My desire and passion to succeed throughout these years have made my PhD journey nothing short of an amazing and wonderful experience. This thesis presents the invaluable lessons learned in how to view, think, and synthesize every substance of knowledge and its counterparts and compile them as a whole. The successful completion of this dissertation within three years has been made possible with the remarkable individuals whom I wish to acknowledge.

First and foremost, I would like to express my deepest gratitude and appreciation to my dear supervisor Professor Dr.Zhari Ismail for encouraging and allowing me to grow as a researcher. I will be forever grateful for his patience, motivation, enthusiasm, endless support and immense knowledge. I truly appreciate his great mentorship and advice throughout my PhD journey. I would also like to thank my co-supervisors, Associate Professor Dr. Amin Malik Shah Amdul Majid, Dr. Chern Ein Oon and Dr. Khamsah Suryati Binti Mohd who have been actively interested in my work and provided me with their invaluable guidance and advice throughout my PhD studies.

My heartfelt thanks to my beloved husband, Emad, for his encouragement and infinite support. He has always inspired and motivated me for excellence in life. I am grateful for his love, encouragement, and tolerance, the man who has made all the difference in my life. A special word of thanks also goes to my mother (Leila), father (Esmaeil), and my sisters, Elham and Azadeh for their continuous support, love and encouragement. My husband and family are the most important people in my world and I dedicate this thesis to them.

The successful conduct of this research would not have been possible without the support of Universiti Sains Malaysia (USM) through providing USM Fellowship. My appreciation to Institute of Postgraduate Studies (IPS), School of Pharmaceutical Sciences and the academic staff of Universiti Sains Malaysia for their support and assistance. My sincere thanks also goes to Ministry of Agriculture and Argo-Based Industry, Malaysia for funding this project (grant number; 304/PFARMASI/650582/K123). Finally, many thanks to my friends and colleagues for helping me overcome the obstacles throughout my studies and achieve my goals.

"If you want your life to be a magnificent story, then begin by realizing that you are the author and everyday you have the opportunity to write a new page."

- Mark Houlahan

# TABLE OF CONTENTS

ACK	NOWLEDGEMENT	ii
TAB	LE OF CONTENTS	iv
LIST	OF TABLES	xiii
LIST	OF FIGURES	xviii
LIST	OF ABREVIATIONS	xxvi
LIST	OF SYMBOLS	xxxiii
ABST	ΓRAK	xxxiv
ABST	ΓRACT	xxxvii
СНА	PTER 1 INTRODUCTION	1
1.1	Herbal Medicines and Hypertension	1
1.2	Standardisation of Herbal Medicine	3
1.3	Justification of the Research	5
1.4	General Objectives	6
1.5	Specific Objectives	6
1.6	Hypotheses	7
1.7	Significance of Study	7
1.8	Methodology Flowchart	7
СНА	PTER 2 LITERATURE REVIEW	10
2.1	Orthosiphon stamineus	10
	2.1.1 Taxonomy	10
	2.1.2 Ethnopharmacology	11
2.2	Review of Chemical Constituents of Orthosiphon stamineus	12
2.3	Review of Biological and Pharmacological Activities of Orthosip	hon
	stamineus	20
2.4	Hypertension	31

	2.4.1	Definition of Hypertension	31
	2.4.2	The Pathophysiology of Hypertension	32
		2.4.2(a) Cardiac Output and Peripheral Resistance	33
		2.4.2(b) Endothelial Dysfunction	35
		2.4.2(c) Renin-Angiotensin-Aldosterone System	35
		2.4.2(d) Autonomic Nervous System	36
	2.4.3	Treatment of Hypertension	37
		2.4.3(a) Diet and Lifestyle Modification	38
		2.4.3(b) Drug Therapy	38
	2.4.4	Risk Management	41
2.5	Liposo	omal Drug Delivery System	43
2.6	The T	herapeutic Potential of microRNAs	46
		3 MATERIALS AND METHODS	
3.1		ials	
	3.1.1	Materials/Chemicals/Reagents	
	3.1.2	Equipment and Apparatus	
3.2	_	ration of Plant Materials	
3.3		metric Analysis of Orthosiphon stamineus Leaves	
	3.3.1	Microscopic Analysis	
	3.3.2	Determination of Foreign Matter	
	3.3.3	Loss on Drying	54
	3.3.4	Total Ash Content	54
	3.3.5	Acid Insoluble Ash Content	55
	3.3.6	Water Soluble Ash	55
	3.3.7	Sulphated Ash	56
	3.3.8	Extractive Value	56
		3.3.8(a) Cold Method	56
		3.3.8(b) Hot Method	56
	3.3.9	Heavy Metals Content	57
3.4	Micro	bial Limit Test (MLT)	58
	3.4.1	Procedure for Total Aerobic Microbial, Yeast and Mold Count (via	ì
		pour plate)	58
	3.4.2	Procedure for Enterobacteria and Gram-negative	58

	3.4.3	Procedur	es for Specific Microorganisms Test	59
		3.4.3(a)	Escherichia coli	59
		3.4.3(b)	Salmonella sp	59
		3.4.3(c)	Pseudomonas aeruginosa and Staphylococcus aureus	59
3.5	Prepar	ration of th	ne Orthosiphon stamineus Leaf Extracts	59
3.6	Standa	ardisation (	of Orthosiphon stamineus Crude Extracts	60
	3.6.1	Ultra Vic	olet-Visible (UV) Analysis	60
	3.6.2	Fourier T	Fransform Infrared (FT-IR) Spectroscopy	61
	3.6.3	Fourier T	Transform Near-Infrared (FT-NIR) Spectroscopy	61
	3.6.4	High Per	formance Thin Layer Chromatography (HPTLC)	61
	3.6.5	High Per	formance Liquid Chromatography (HPLC)	62
		3.6.5(a)	High Performance Liquid Chromatography Method for	
			Analysis of RA, TMF, SIN and EUP in Orthosiphon	
			stamineus Leaf Extracts	62
		3.6.5(b)	High Performance Liquid Chromatography Method for	
			Analysis of Free Amino Acids in Orthosiphon stamineur	S
			Leaf Extracts	63
		3.6.5(c)	Quantification of Standard Compounds in Orthosiphon	
			stamineus Leaf Extracts by HPLC Methods	66
3.7	Analy	sis of Prim	nary and Secondary Metabolites in Orthosiphon stamineu	S
	Leaf F	Extracts		67
	3.7.1	Determin	nation of Glycosaponins	67
	3.7.2	Determin	nation of Total Proteins	68
	3.7.3	Determin	nation of Total Polysaccharides	69
	3.7.4	Determin	nation of Total Flavonoids	70
	3.7.5	Determin	nation of Total Polyphenolics	71
	3.7.6	Determin	nation of Total Phospholipids	72
3.8	In Viti	ro Antihyp	ertensive Effect of Orthosiphon stamineus Leaf Extracts	73
	3.8.1	Develop	ment of High Performance Liquid Chromatography Metho	od
		for Meas	urement of Angiotensin Converting Enzyme Inhibition	
		Activity.		73
		3.8.1(a)	Chromatographic Conditions	73
		3.8.1(b)	HA Calibration Curve	74
		3.8.1(c)	ACE Calibration Curve	74

		3.8.1(d) Validation of HPLC Method	75
	3.8.2	In Vitro Angiotensin Converting Enzyme Inhibition Activity	76
	3.8.3	Chelation of Zinc Ion (II) by Orthosiphon stamineus Ethanolic Extr	act,
		RA, TMF, SIN, EUP and Captopril	76
3.9	Molec	ular Docking Study	77
	3.9.1	Binding (Antagonistic) Potential of Orthosiphon stamineus Active	
		Markers against Angiotensin Converting Enzyme	77
3.10	In Vive	o Antihypertensive Effect of Orthosiphon stamineus Leaf Extracts or	ì
	Sponta	nneous Hypertensive Rats (SHR)	78
	3.10.1	Experimental Animals	78
	3.10.2	Ethical Approval	79
	3.10.3	Indirect Blood Pressure Measurement by Tail-Cuff Method (Screen	ing
		Study)	79
	3.10.4	Direct Blood Pressure Measurement by Catheterization of Carotid	
		Artery (Screening Study)	80
3.11	Prepar	ration and Characterization of Liposomes of Orthosiphon stamineus	
	Ethano	olic Extract in Deoiled Soya Lecithin	80
	3.11.1	Preparation of Soybean Phospholipids	80
	3.11.2	Preparation of Liposomes of Orthosiphon stamineus Ethanolic Extra	act
			81
	3.11.3	Characterization of Liposomes of Orthosiphon stamineus Ethanolic	
		Extract	81
		3.11.3(a) Aqueous Solubility	81
		3.11.3(b) Effect of pH on OS-EL Stability	82
		3.11.3(c) Determination of Entrapment Efficiency	82
		3.11.3(d) Fourier Transform Infrared Spectroscopy (FT-IR)	83
		3.11.3(e) Measurement of Particle Size and Zeta Potential	83
		3.11.3(f) Transmission Electron Microscopy	83
3.12	Pharm	acokinetics and Bioavailability of Orthosiphon stamineus Ethanolic	
	Extrac	t and its Nano Liposomes in Sprague Dawley Rats	84
	3.12.1	Instrumentation	84
	3.12.2	Plasma Sample Extraction	84
	3.12.3	Development and Validation of HPLC Method	84
		3.12.3(a) Calibration Curves (Linearity Ranges)	84

	3.12.3(b) Limit of Detection (LOD) and Limit of Quantification	
	(LOQ)	. 85
	3.12.3(c) Intra-day and Inter-day Precisions	. 85
	3.12.3(d) Recovery of Plasma Extraction	. 85
	3.12.4 Experimental Animals	. 85
	3.12.5 Ethical Approval	. 86
	3.12.6 Pilot Pharmacokinetic Study of OS-E and OS-EL	. 86
	3.12.7 Pharmacokinetic Study of OS-E and OS-EL	. 87
	3.12.8 Determination of Pharmacokinetic Parameters	. 87
	3.12.9 Statistical Analysis	. 88
3.13	Accelerated Stability Studies of Orthosiphon stamineus Ethanolic Extract	and
	Its Nano Liposomes	. 88
	3.13.1 Stability Study Protocol	. 88
	3.13.2 High Performance Liquid Chromatography	. 89
	3.13.3 Calculations of Chemical Kinetic Parameters	. 89
	3.13.3(a) Order of the Reaction	. 89
	3.13.3(b) Activation Energy	. 90
	3.13.3(c) Shelf Life (t <sub>90</sub> )	. 91
	3.13.4 FTIR Spectrophotometer	. 91
	3.13.4(a) Data Processing for Chemometric Analysis	. 91
3.14	Toxicity Studies of Orthosiphon stamineus Ethanolic Extract and Its Nano	
	Liposomes in Sprague Dawley Rats	. 92
	3.14.1 Experimental animals	. 92
	3.14.2 Ethical Approval	. 92
	3.14.3 Acute Toxicity Study in Rats	. 92
	3.14.4 Repeated Dose 28-days Oral Toxicity Study in Rats	. 93
	3.14.5 Haematological and Biochemical Analysis	. 94
	3.14.6 Histopathological Analysis	. 95
	3.14.7 Analysis of Antimutagenic Effects: Ames Test	. 95
	3.14.8 Statistical Analysis	. 96
3.15	In Vivo Antihypertensive Effect of Orthosiphon stamineus Ethanolic Extra	ct
	and Its Nano Liposomes on Spontaneous Hypertensive Rats (SHR)	. 96
	3.15.1 Indirect Blood Pressure Measurement by Tail Cuff Method	. 96

	3.15.2	Direct Bl	ood Pressure Measurement by Catheterization of Ca	arotid
		Artery		97
	3.15.3	Organ Co	ollection	97
	3.15.4	Determin	nation of ACE Activity in Plasma and Tissues	98
3.16	In Viv	o Antihype	ertensive Effect of Orthosiphon stamineus Ethanolic	Extract
	and Its	s Nano Lip	osomes on Normotensive Wistar Kyoto (WKY) Ra	ts 98
3.17	Micro	RNA Expi	ression Study	99
	3.17.1	Cell Line	es and Culture Conditions	99
	3.17.2	Proliferat	tion Assay	100
	3.17.3	RNA Iso	lation	101
	3.17.4	Quality A	Assessment of Total RNA	102
	3.17.5	Expression	on Analysis	103
	3.17.6	Statistica	l Analysis	104
CHAI	PTER 4	4 RESULT	TS AND DISCUSSION	105
4.1	Gravi	metric Ana	ılysis	105
4.2	Micro	bial Limit	Test (MLT)	108
4.3	Prepar	ration of th	e Orthosiphon stamineus Extracts	109
4.4	Standa	ardisation o	of Orthosiphon stamineus Crude Extracts	110
	4.4.1	Ultra Vic	olet-Visible (UV) Analysis	110
	4.4.2	Fourier T	ransform Infrared (FT-IR) Spectroscopy	111
	4.4.3	Fourier T	ransform Near-Infrared (FT-NIR) Spectroscopy	115
	4.4.4	High Per	formance Thin Layer Chromatography (HPTLC)	117
	4.4.5	High Per	formance Liquid Chromatography	118
		4.4.5(a)	Analysis of RA, TMF, SIN and EUP in Orthosiphe	on
			stamineus Leaf Extracts	118
		4.4.5(b)	Analysis of Free Amino Acids in Orthosiphon stan	nineus
			Leaf Extracts	119
		4.4.5(c)	Validation of Developed HPLC Method	124
		4.4.5(d)	Quantification of Free Amino Acids in Orthosipho	n
			stamineus Leaf Extracts by HPLC Method	133
4.5	Analy	sis of Prim	nary and Secondary Metabolites in Orthosiphon stan	nineus
	Leaf E	Extracts		137
4.6	In Viti	ro Antihyp	ertensive Effect of Orthosiphon stamineus Leaf Ext	racts.139

	4.6.1	Developr	ment of HPLC Method for Measurement of Angiotens	in
		Convertin	ng Enzyme Inhibition Activity	139
	4.6.2	Validatio	on of HPLC Method	139
		4.6.2(a)	Calibration Curves, Linearity Ranges, Limit of Detec	ction
			(LOD) and Limit of Quantification (LOQ)	139
		4.6.2(b)	Intra-day and Inter-day Precisions and Recovery of H	HPLC
			Method	141
	4.6.3	In Vitro A	Angiotensin Converting Enzyme Inhibition Activity	142
	4.6.4	Correlation	on between ACE Inhibition and RA, TMF, SIN and E	UP
		Concentr	rations in Orthosiphon stamineus Leaf Extracts	144
	4.6.5	Chelation	n of Zinc Ion (II) by Orthosiphon stamineus Ethanolic	Extract,
		RA, TMF	F, SIN, EUP and Captopril	145
4.7	Molec	ular Docki	ing Study	148
	4.7.1	Binding (	(Antagonistic) Potential of Orthosiphon stamineus Ac	tive
		Markers	against Angiotensin Converting Enzyme	148
4.8	In Vivo	Antihype	ertensive Effect of Orthosiphon stamineus Leaf Extrac	ets on
	Sponta	neous Hy	pertensive Rats (SHR)	154
4.9	Characterization of Liposomes of Orthosiphon stamineus Ethanolic Extract			
	(OS-E	L)		156
	4.9.1	Aqueous	Solubility	156
	4.9.2	Effect of	pH on OS-EL Stability	157
	4.9.3	Determin	nation of Entrapment Efficiency	158
	4.9.4	Fourier T	Transform Infrared Spectroscopy (FT-IR)	159
	4.9.5	Measurer	ment of Particle Size and Zeta Potential	162
	4.9.6	Transmis	sion Electron Microscopy	163
4.10	Pharm	acokinetic	es and Bioavailability of Orthosiphon stamineus Ethan	olic
	Extrac	t and its N	Jano Liposomes in Sprague Dawley Rats	164
	4.10.1	Developr	ment and Validation of HPLC Method	164
		4.10.1(a)	Calibration curves, linearity ranges, Limit of Detecti	on
			(LOD) and Limit of Quantification (LOQ)	165
		4.10.1(b)	Intra-day and Inter-day Precisions	166
		4.10.1(c)	Recovery of Plasma Extraction	169
	4.10.2	Pilot Pha	rmacokinetic Study of OS-E and OS-EL	170
	4.10.3	Pharmaco	okinetic Study of OS-E and OS-EL	171

4.11	Accelerated Stability Studies of Orthosiphon stamineus Ethanolic Extract	and
	Its Nano Liposomes	179
	4.11.1 High Performance Liquid Chromatography	179
	4.11.2 Chemical Kinetic Parameters	185
	4.11.2(a) Order of the Reaction	185
	4.11.2(b) Activation Energy	189
	4.11.2(c) Shelf Life (t90)	190
	4.11.3 Stability Study of OS-E and OS-EL by Chemical Fingerprinting U	sing
	Fourier Transform Infrared (FTIR) Spectroscopy and Principal	
	Component Analysis (PCA)	191
	4.11.4 Hierarchical Clustering Analysis (HCA) for FTIR Fingerprinting	204
4.12	Toxicity Studies of Orthosiphon stamineus Ethanolic Extract and Its Nano	)
	Liposomes in Sprague Dawley Rats	208
	4.12.1 Acute Toxicity Study in Rats	208
	4.12.2 Repeated Dose 28-days Oral Toxicity Study in Rats	209
	4.12.3 Haematological and Biochemical Analysis	212
	4.12.4 Histopathological Analysis	216
	4.12.5 Bacterial Reverse Mutation Test	222
4.13	In Vivo Antihypertensive Effect of Orthosiphon stamineus Ethanolic Extra	act
	and Its Nano Liposomes on Spontaneous Hypertensive Rats (SHR)	228
	4.13.1 Indirect Blood Pressure Measurement by Tail Cuff Method	228
	4.13.2 Direct Blood Pressure Measurement by Catheterization of Carotid	
	Artery	229
	4.13.3 Determination of ACE Activity in Plasma and Tissues	234
	4.13.3 In Vivo Antihypertensive Effect of Orthosiphon stamineus Ethanol	ic
	Extract and Its Nano Liposomes on Normotensive Wistar Kyoto	
	(WKY) Rats	238
4.14	MicroRNA Expression Study	241
	4.14.1 Proliferation Assay	241
	4.14.2 Quality Assessment of Total RNA	242
	4.14.3 Expression Analysis	245
	4.14.3(a) Unsupervised Analysis	245
	4.14.3(b) Expression Analysis for HR-1 and HR-2 versus HR-NC	250
	4.14.3(c) Expression Analysis for EA-1 and EA-2 versus EA-NC.	254

CHA	PTER 5 GENERAL DISCUSSION AND CONCLUSION	262
5.1	General Discussion	262
5.2	Conclusion	268
5.3	Suggestions for Further Studies	270
REFI	ERENCES	271
APPI	ENDICES	292

# LIST OF TABLES

		Page
Table 2.1	Chemical constituents of Orthosiphon stamineus	12
Table 2.2	Chemical structures of Orthosiphon stamineus	14
Table 2.3	Summary of literatures on biological and pharmacological activities of <i>O. stamineus</i>	27
Table 3.1	Materials used in this study	48
Table 3.2	Equipment and apparatus used in this study	52
Table 3.3	Parameter of microwave digestion	57
Table 3.4	Scheme of elution gradient for HPLC analysis	64
Table 4.1	Gravimetric analysis of O. stamineus raw material	107
Table 4.2	Heavy metal content of Orthosiphon stamineus leaves	108
Table 4.3	Microbial content in the leaves of Orthosiphon stamineus	109
Table 4.4	Percentage yield of extracts from <i>Orthosiphon stamineus</i> leaves. Results are the mean $\pm$ SD (n=3)	109
Table 4.5	Summary of functional groups in FT-IR spectrum of <i>O. stamineus</i> extracts (4000-650 cm <sup>-1</sup> )	114
Table 4.6	Quantification of four marker compounds in <i>Orthosiphon stamineus</i> extracts. Results are shown as average (mg/g; marker compound/extract)	118
Table 4.7	Chemical structures of tested amino acids	122
Table 4.8	Linear correlation between peak area and concentration of marker compounds, limit of detection (LOD) and limit of quantification (LOQ) of the reported HPLC method	125
Table 4.9	Precision of the developed HPLC method for determination of reference compounds in <i>Orthosiphon stamineus leaf</i> extracts	126
Table 4.10	Recovery of spiked reference compounds in different extracts of Orthosiphon stamineus	130

Table 4.11	The content of free amino acids in O. stamineus extracts	136
Table 4.12	Analysis of primary and secondary metabolites content present in different extracts of $O$ . stamineus. The results are expressed as mean $\pm$ SD (n=3)	138
Table 4.13	Linear correlation between peak area and concentration of HA and ACE, limit of detection (LOD) and limit of quantification (LOQ) of the reported HPLC method	141
Table 4.14	Intra-day and inter-day precisions of the reported HPLC method	141
Table 4.15	Recovery of HA with the reported HPLC method	142
Table 4.16	The half-maximal inhibitory concentration (IC <sub>50</sub> ) of <i>O. stamineus</i> extracts, standard compounds and captopril on <i>in vitro</i> ACE inhibitory assay, each value represents mean $\pm$ SD (n=3)	144
Table 4.17	Summary of the docking scores and reported binding affinities of RA, SIN, TMF and EUP compounds with ACE in the docking analysis with LeadIT FlexX Scoring functions	150
Table 4.18	Data of the invasive blood pressure measurements after 14 days treatment with the <i>O. stamineus</i> ethanolic (OS-E), 50% ethanolic (OS-EW) extracts at 250 mg/kg/day, captopril at 5 mg/kg/day, 0.5% carboxymethyl cellulose (CMC) and water (vehicle). The results are shown as mean $\pm$ S.E.M (n = 6). * ( $P \le 0.05$ ) significant vs. water, ** ( $P \le 0.05$ ) significant vs.0.5% CMC	156
Table 4.19	HPLC analysis of marker compounds in <i>O.stamineus</i> ethanolic extract (OS-E) and liposomes of <i>O. stamineus</i> ethanolic extract (OS-EL). Results are shown as average peak area (mAU) $\pm$ SD (n = 3)	157
Table 4.20	Effect of pH on stability of marker compounds in liposomes of O. stamineus ethanolic extract (OS-EL). The results showed percentage of soluble marker compounds in PBS at different pH relative to that in water. The results are expressed as mean $\pm$ SD (n = 3)	157
Table 4.21	Percentage of entrapment efficiency of nano liposomes of O. stamineus ethanolic extract (OS-EL) and its marker compounds. The results are expressed as mean $\pm$ SD (n = 3)	159
Table 4.22	Summary of functional groups from ethanolic extract of <i>O. stamineus</i> (OS-E), soybean phospholipids (SPL), and liposomes of <i>O. stamineus</i> ethanolic extract (OS-EL) by FT-IR	160

Table 4.23	Analysis of particle size and zeta potential by Photon Correlation Spectroscopy	163
Table 4.24	Calibration data, LOD and LOQ of the reported HPLC method	166
Table 4.25	Intra-day and inter-day precision ( $n = 6$ ) for rosmarinic acid (RA), 3'-hydroxy-5,6,7,4'-tetramethoxyflavone (TMF), sinensitin (SIN) and eupatorin (EUP)	167
Table 4.26	Plasma extraction recovery of rosmarinic acid (RA), 3'-hydroxy-5,6,7,4'-tetramethoxyflavone (TMF), sinensitin (SIN) and eupatorin (EUP)	169
Table 4.27	Pharmacokinetic parameters of rosmarinic acid (RA), 3-hydroxy-5,6,7,4 tetramethoxyflavone (TMF), sinensitin (SIN) and eupatorin (EUP) in rat plasma after intravenous administration of <i>O. stamineus</i> ethanolic extract (OS-E) and nano liposomes of <i>O. stamineus</i> ethanolic extract (OS-EL) (n=6)	174
Table 4.28	Pharmacokinetic parameters of rosmarinic acid (RA), 3-hydroxy-5,6,7,4 tetramethoxyflavone (TMF), sinensitin (SIN) and eupatorin (EUP) in rat plasma after oral administration of <i>O. stamineus</i> ethanolic extract (OS-E) and nano liposomes of <i>O. stamineus</i> ethanolic extract (OS-EL) (n=6)	176
Table 4.29	Calibration data of reference markers for the reported HPLC method	181
Table 4.30	Remaining percentage of rosmarinic acid (RA), 3'-hydroxy-5,6,7,4'-tetramethoxyflavone (TMF), sinensitin (SIN) and eupatorin (EUP) of <i>Orthosiphon stamineus</i> ethanolic extract (OS-E) stored for 6 months under different storage conditions	182
Table 4.31	Remaining percentage of rosmarinic acid (RA), 3'-hydroxy-5,6,7,4'-tetramethoxyflavone (TMF), sinensitin (SIN) and eupatorin (EUP) of nano liposomes of <i>Orthosiphon stamineus</i> ethanolic extract (OS-EL) stored for 6 months under different storage conditions	183
Table 4.32	Rate constant (K), activation energy (Ea) and pre-exponential factor (A) of the RA, TMF, SIN and EUP of <i>Orthosiphon stamineus</i> ethanolic extract (OS-E) stored at different temperatures	190
Table 4.33	Rate constant (K), activation energy (Ea) and pre-exponential factor (A) of the RA, TMF, SIN and EUP of nano liposomes of <i>Orthosiphon stamineus</i> ethanolic extract (OS-EL) stored at different temperatures	190

Table 4.34	Shelf life (t90) of the markers in <i>Orthosiphon</i> stamineus ethanolic extract (OS-E) at different storage conditions	191
Table 4.35	Shelf life (t90) of the markers in nano liposomes of <i>Orthosiphon</i> stamineus ethanolic extract (OS-EL) at different storage conditions	191
Table 4.36	Effects of the repeated dose oral administration of OS-EL on organ weights in Sprague Dawley rats	211
Table 4.37	Effects of the repeated dose oral administration of OS-EL on hematological parameters in Sprague Dawley rats	213
Table 4.38	Effects of the repeated dose oral administration of OS-EL on biochemical parameters in Sprague Dawley rats	215
Table 4.39	The number of positive wells scored in a 96-well microplate leading to clear significance in the fluctuation test	222
Table 4.40	Data of the invasive blood pressure measurements after 28 days treatment with the <i>O. stamineus</i> ethanolic extract(OS-E), nano liposomes of <i>O. stamineus</i> ethanolic extract (OS-EL) at 250 mg/kg/day, captopril at 5 mg/kg/day or negative controls of 0.5% carboxymethyl cellulose (CMC), lecithin and water (vehicle). The results are shown as mean $\pm$ S.E.M (n = 6). * ( $P \le 0.05$ ) significant vs. water, ** ( $P \le 0.05$ ) significant vs.0.5% CMC, *** ( $P \le 0.05$ ) significant vs. lecithin	230
Table 4.41	Percent organ weight to body weight ratios after 28 days treatment with the <i>O. stamineus</i> ethanolic extract (OS-E), nano liposomes of <i>O. stamineus</i> ethanolicextract (OS-EL) at 250 mg/kg/day, captopril at 5 mg/kg/day or negative controls of 0.5% carboxymethyl cellulose (CMC), lecithin and water (vehicle). Each value represents the mean $\pm$ S.E.M. (n = 6). * ( $P \le 0.05$ ) significant vs. water, ** ( $P \le 0.05$ ) significant vs.0.5% CMC, *** ( $P \le 0.05$ ) significant vs. lecithin	237
Table 4.42	Data of the invasive blood pressure measurements of Normotensive Wistar Kyoto (WKY) rats after 28 days treatment with the $O$ . stamineus ethanolic extract (OS-E), nano liposomes of $O$ . stamineus ethanolic extract (OS-EL) at 250 mg/kg/day, captopril at 5 mg/kg/day or negative controls of 0.5% carboxymethyl cellulose (CMC), lecithin and water (vehicle). The results are shown as mean $\pm$ S.E.M (n = 6)	240
Table 4.43	The half-maximal effective concentration (EC50) of O. <i>stamineus</i> ethanolic extract (OS-E) and nano liposomes of O. <i>stamineus</i> ethanolic extract (OS-EL) on HRGEC and EAHY 926 cell lines. Each value represents mean ± S.E.M. (n=3)	242

# LIST OF FIGURES

		Page
Figure 1.1	Methodology flowchart	9
Figure 2.1	Pictures of Orthosiphon stamineus leaves and flower	11
Figure 2.2	Pathophysiologic mechanisms of hypertension (Oparil et al., 2003)	33
Figure 2.3	The heart, arteries, and arterioles in hypertension	34
Figure 2.4	Local versus systemic rennin-angiotensin system	34
Figure 2.5	Renin-angiotensin system and effects on blood pressure and aldosterone release	36
Figure 2.6	The autonomic nervous system and its control of blood pressure (Beevers et al., 2001)	37
Figure 2.7	Schematic representation of the different types of liposomal drug delivery systems. (A) Conventional liposome; (B) PEGylated liposome; (C) Ligand-targeted liposome; (D) Theranostic liposome	44
Figure 2.8	Protein inhibition with miRNA (Lauren, 2010)	46
Figure 3.1	Flowchart of analytical analysis of <i>Orthosiphon stamineus</i> extracts used in this study	60
Figure 3.2	Flowchart of miRNA expression study	99
Figure 4.1	Microscopic characters of <i>Orthosiphon stamineus</i> leaves powder (A) Parenchyma cell (40X) (B) Diacytic stomata (40X) (C) Spiral secondary wall (100X) (D) Epidermal cells with diacytic stomata of tracheary elements (100X) (E) Simple, uniseriatetrichome (100X)u	105
Figure 4.2	UV-Vis spectra of <i>Orthosiphon stamineus</i> (OS) extracts and reference compounds. UV-Vis spectra were collected in the wavelength range 500-200 nm	111
Figure 4.3	Figure 4.3: FT-IR fingerprints of <i>Orthosiphon stamineus</i> extracts. FT-IR spectra were collected in the wavenumber range 4000-650 cm <sup>-1</sup>	113

Figure 4.4	Figure 4.4: FT-NIR fingerprints of <i>Orthosiphon stamineus</i> extracts. FT-NIR spectra were collected in the wavenumber range 8000-4000 cm <sup>-1</sup>	116
Figure 4.5	HPTLC fingerprints of <i>Orthosiphon stamineus</i> extracts at 366 nm and reference compounds (TMF, SEN and EUP)	117
Figure 4.6	HPLC chromatograms of (A) reference markers (1: RA, 2: TMF, 3: SIN and 4: EUP), (B) OS-W, (C) OS-E, (D) OS-M, (E) OS-EW and (F) OS-MW at 330 nm	119
Figure 4.7	(A) Chromatogram of amino acid standards, 1) L-aspartic acid, 2) L-glutamic acid, 3) L-serine, 4) L-histidine hydrochloride monohydrate, 5) glycine, 6) L-threonine, 7) L-arginine, 8) L-alanine, 9) L-tyrosine, 10) L-cystine, 11) L-valine, 12) L-methionine, 13) L-phenylalanine, 14) L-isoleucine, 15) L-leucine, 16) L-lysine Hydrochloride, and 17) L-proline; (B) O. <i>stamineus</i> water extract (OS-W); (C) O. <i>stamineus</i> ethanolic extract (OS-E); (D) O. stamineus methanolic extract (OS-M); (E) O. stamineus 50% ethanolic extract (OS-EW) and (F) O. stamineus 50% methanolic extract (OS-MW)	135
Figure 4.8	HPLC chromatograms of (1) HA and (2) HHL standard compounds (A), ACE reaction with negative control (50% ethanol) (B), ACE reaction with <i>O. stamineus</i> water extract (C), ACE reaction with <i>O. stamineus</i> ethanol extract (OS-E) (D), ACE reaction with <i>O. stamineus</i> 50% ethanol extract (OS-EW) (E), ACE reaction with <i>O. stamineus</i> methanol extract (OS-M) (F), ACE reaction with <i>O. stamineus</i> 50% methanol extract (OS-MW) (G) and ACE reaction with captopril (H)	140
Figure 4.9	The dose-response relationship of $O$ . stamineus extracts and standard compounds on in vitro ACE inhibitory assay, data are presented as mean $\pm$ SD, (* $p$ < 0.05)	143
Figure 4.10	The dose-response relationship of captopril on <i>in vitro</i> ACE inhibitory assay, data are presented as mean $\pm$ SD, (* $p$ < 0.05)	143
Figure 4.11	Correlation between (A) RA; (B) TMF; (C) SIN and (D) EUP concentrations and the percentage of ACE inhibition. The graph shows positive correlation which may indicate that the two variables are related ( $P=0.001$ )	145
Figure 4.12	Zn <sup>2+</sup> chelating activity of ethanolic extract of <i>O. stamineus</i> , RA, TMF, SIN, EUP and captopril	146
Figure 4.13	3D visualization of the ligands RA (A), SIN (B), TMF (C), EUP (D) and Captopril (E); and active site residues interaction of protein in ACE	151

Figure 4.14	Surface visualization of proteins in ACE with the ligands RA (A), SIN (B), TMF (C), EUP (D) and Captopril (E). Hydrophobic interaction showed in green region	152
Figure 4.15	Effect of <i>O. stamineus</i> ethanolic (OS-E), 50% ethanolic (OS-EW) extracts at 250 mg/kg/day, captopril at 5 mg/kg/day, 0.5% carboxymethyl cellulose (CMC) and water on systolic blood pressure in SHR rats after 14 days treatment. The results are expresses as mean $\pm$ S.E.M (n = 6). * ( $P \le 0.05$ ) significant vs. water, ** ( $P \le 0.05$ ) significant vs.0.5% CMC and *** ( $P \ge 0.05$ ) no significant vs. Captopril	155
Figure 4.16	FT-IR spectra of ethanolic extract of <i>O. stamineus</i> (OS-E), soybean phospholipids (SPL), and liposomes of <i>O. stamineus</i> ethanolic extract (OS-EL)	162
Figure 4.17	Particle size distribution (A), and zeta potential distribution (B) of liposomes of <i>O. stamineus</i> ethanolic extract (OS-EL)	163
Figure 4.18	Transmission Electron Microscopy (TEM) photograph of liposomes of <i>O. stamineus</i> ethanolic extract (OS-EL) (A), and Soybean phospholipids bilayer (B). TEM indicating presence of round-shaped structures	164
Figure 4.19	Chromatograms of blank rat plasma (A), and rat plasma spiked with 10 $\mu$ g/mL of rosmarinic acid (1), and 3'-hydroxy-5,6,7,4'-tetramethoxyflavone (2), sinensitin (3) and eupatorin (4) (B)	165
Figure 4.20	Chromatograms of rosmarinic acid (1), and 3'-hydroxy-5,6,7,4'-tetramethoxyflavone (2), sinensitin (3) and eupatorin (4) in rat plasma at 1 hour after intravenous administration of 250 mg/kg <i>O. stamineus</i> ethanolic extract (A); rat plasma at 4 hours after oral administration of 1000 mg/kg <i>O. stamineus</i> ethanolic extract (B); rat plasma at 1 hour after intravenous administration of 250 mg/kg nano liposomes of <i>O. stamineus</i> ethanolic extract (C); rat plasma at 4 hours after oral administration of 500 mg/kg nano liposomes of <i>O. stamineus</i> ethanolic extract (D)	171
Figure 4.21	Mean plasma concentration vs. time profiles (mean±S.E.M, n=6) of marker compounds namely; rosmarinic acid (RA), 3-hydroxy-5,6,7,4 tetramethoxyflavone (TMF), sinensitin (SIN) and eupatorin (EUP) after intravenous at 250 mg/kg (A) and oral at 1000 mg/kg (B) administration of <i>O. stamineus</i> ethanolic extract (OS-E)	172

Figure 4.22	Mean plasma concentration vs. time profiles (mean± S.E.M, n=6) of marker compounds namely; rosmarinic acid (RA), 3-hydroxy-5,6,7,4 tetramethoxyflavone (TMF), sinensitin (SIN) and eupatorin (EUP) after intravenous at 250 mg/kg (A) and oral at 500 mg/kg (B) administration of nano liposomes of <i>O. stamineus</i> ethanolic extract (OS-EL)	172
Figure 4.23	(A) HPLC chromatograms of marker compounds; rosmarinic acid (1), 3'-hydroxy-5,6,7,4'-tetramethoxyflavone (2), sinensitin (3) and eupatorin (4); (B) Ethanol extract of <i>Orthosiphon stamineus</i> (OS-E) stored at 30°C/75% RH at month zero; (C) Ethanol extract of <i>Orthosiphon stamineus</i> (OS-E) stored at 30°C/75% RH at month three and (D) Ethanol extract of <i>Orthosiphon stamineus</i> (OS-E) stored at 30°C/75% RH at month six	180
Figure 4.24	(A) HPLC chromatograms of marker compounds rosmarinic acid (1), 3'-hydroxy-5,6,7,4'-tetramethoxyflavone (2), sinensitin (3) and eupatorin (4); (B) Nano liposomes of ethanol extract of <i>Orthosiphon stamineus</i> (OS-EL) stored at 30°C/75% RH at month zero; (C) Nano liposomes of ethanol extract of <i>Orthosiphon stamineus</i> (OS-EL) stored at 30°C/75% RH at month three and (D) Nano liposomes of ethanol extract of <i>Orthosiphon stamineus</i> (OS-EL) stored at 30°C/75% RH at month six	181
Figure 4.25	Plot of ln percentage remaining concentration of the markers in <i>Orthosiphon stamineus</i> ethanolic extract (OS-E) versus time for first order reaction	186
Figure 4.26	Plot of ln percentage remaining concentration of the markers in nano liposomes of <i>Orthosiphon stamineus</i> ethanolic extract (OS-EL) versus time for first order reaction	188
Figure 4.27	Plot of natural log of rate constant versus inverse of temperature (Kelvin-1) of RA, TMF, SIN and EUP in (A) <i>Orthosiphon stamineus</i> ethanolic extract (OS-E) and (B) nano liposomes of <i>Orthosiphon stamineus</i> ethanolic extract (OS-EL) at various temperatures, ln K (natural log of rate constant); 1/T (inverse of temperature)	189
Figure 4.28	Fourier Transform Infrared (FTIR) Spectra of ethanolic extract of <i>Orthosiphon stamineus</i> (OS-E) stored at A: 30°C/75% RH, B: 40°C/75% RH, C: 50°C/75% RH and D: 60°C/75% RH for six months	192
Figure 4.29	Fourier Transform Infrared (FTIR) Spectra of nano liposomes of ethanolic extract of <i>Orthosiphon stamineus</i> (OS-EL) stored at A: 30°C/75% RH, B: 40°C/75% RH, C: 50°C/75% RH and D: 60°C/75% RH for six months.	193

Figure 4.30	PCA plots of ethanolic extract of <i>Orthosiphon stamineus</i> (OS-E) stored at A: 30°C/75% RH, B: 40°C/75% RH, C: 50°C/75% RH and D: 60°C/75% RH for six months. M0-M6 refer to storage months.	195
Figure 4.31	loading plots of PC-1 for ethanolic extract of <i>Orthosiphon stamineus</i> (OS-E) stored at A: 30°C/75% RH, B: 40°C/75% RH, C: 50°C/75% RH and D: 60°C/75% RH	197
Figure 4.32	loading plots of PC-2 for ethanolic extract of <i>Orthosiphon stamineus</i> (OS-E) stored at A: 30°C/75% RH, B: 40°C/75% RH, C: 50°C/75% RH and D: 60°C/75% RH	198
Figure 4.33	PCA plots of nano liposomes of ethanolic extract of <i>Orthosiphon stamineus</i> (OS-EL) stored at A: 30°C/75% RH, B: 40°C/75% RH, C: 50°C/75% RH and D: 60°C/75% RH for six months. M0-M6 refer to storage months	200
Figure 4.34	loading plots of PC-1 for nano liposomes of ethanolic extract of <i>Orthosiphon stamineus</i> (OS-EL) stored at A: 30°C/75% RH, B: 40°C/75% RH, C: 50°C/75% RH and D: 60°C/75% RH	202
Figure 4.35	loading plots of PC-2 for nano liposomes of ethanolic extract of <i>Orthosiphon stamineus</i> (OS-EL) stored at A: 30°C/75% RH, B: 40°C/75% RH, C: 50°C/75% RH and D: 60°C/75% RH	203
Figure 4.36	HCA dendograms of ethanolic extract of <i>Orthosiphon stamineus</i> (OS-E) stored at A: 30°C/75% RH, B: 40°C/75% RH, C: 50°C/75% RH and D: 60°C/75% RH for six months. M0-M6 refer to storage months	205
Figure 4.37	HCA dendograms of nano liposomes of ethanolic extract of <i>Orthosiphon stamineus</i> (OS-EL) stored at A: 30°C/75% RH, B: 40°C/75% RH, C: 50°C/75% RH and D: 60°C/75% RH for six months. M0-M6 refer to storage months	206
Figure 4.38	Body weight changes in acute toxicity study of SD female rats dosed by OS-E and OS-EL at 5000 mg/kg after 14 days	209
Figure 4.39	Body weight changes of female (A) and male (B) Sprague Dawley rats during the 28-day toxicological assessment. The vehicle, water (10 ml/kg/day), was administered to rats in the control group. No significant differences were detected between the treated (250, 500 and 1000 mg/kg) and control (vehicle 10 ml/kg) groups. All values are expressed as the mean $\pm$ S.E.M. (n = 5)	210

Figure 4.40	Representative microscopic findings for the heart and kidneys of female Sprague Dawley rats treated orally with 250, 500 and 1000 mg/kg OS-EL or the control (water) for 28 days	218
Figure 4.41	Representative microscopic findings for the liver and spleen of female Sprague Dawley rats treated orally with 250, 500 and 1000 mg/kg OS-EL or the control (water) for 28 days	219
Figure 4.42	Representative microscopic findings for the heart and kidneys of male Sprague Dawley rats treated orally with 250, 500 and 1000 mg/kg OS-EL or the control (water) for 28 days	220
Figure 4.43	Representative microscopic findings for the liver and spleen of male Sprague Dawley rats treated orally with 250, 500 and 1000 mg/kg OS-EL or the control (water) for 28 days	221
Figure 4.44	Systolic blood pressure changes of Spontaneous Hypertensive Rats (SHR) during 28-day treatment with <i>O. stamineus</i> ethanolic extract (OS-E), nano liposomes of <i>O. stamineus</i> ethanolic extract (OS-EL) at 250 mg/kg/day and captopril at 5 mg/kg/day. The negative controls of 0.5% carboxymethyl cellulose (CMC), lecithin and water (10 ml/kg/day), were administered to rats in the control groups. The results are expresses as mean $\pm$ S.E.M (n = 6). * ( $P \le 0.05$ ) significant vs. water, ** ( $P \le 0.05$ ) significant vs.0.5% CMC and *** ( $P \le 0.05$ ) significant vs. lecithin, **** ( $P \ge 0.05$ ) no significant vs. captopril	228
Figure 4.45	Body weight changes of Spontaneous Hypertensive Rats (SHR) during 28-day treatment with $O$ . stamineus ethanolic extract (OS-E), nano liposomes of $O$ . stamineus ethanolic extract (OS-EL) at 250 mg/kg/day and captopril at 5 mg/kg/day. The negative controls of 0.5% carboxymethyl cellulose (CMC), lecithin and water (10 ml/kg/day), were administered to rats in the control groups. No significant differences were detected between the treated and control (vehicle 10 ml/kg) groups. All values are expressed as the mean $\pm$ S.E.M. (n = 6)	229
Figure 4.46	The activity of angiotensin I-converting enzyme (ACE) in plasma and tissues from Spontaneous Hypertensive Rats (SHR) after 28 days treatment with the <i>O. stamineus</i> ethanolic extract (OS-E), nano liposomes of <i>O. stamineus</i> ethanolic extract (OS-EL) at 250 mg/kg/day, captopril at 5 mg/kg/day or negative controls of 0.5% carboxymethyl cellulose (CMC), lecithin and water (vehicle). The samples were collected four hours after last treatment. The results are shown as mean $\pm$ S.E.M (n = 6). * ( $P \le 0.05$ ) significant vs. water, ** ( $P \le 0.05$ ) significant vs.0.5% CMC, *** ( $P \le 0.05$ ) significant vs. lecithin	236

Figure 4.47	rats (WKY) during 28-day treatment with $O$ . stamineus ethanolic extract (OS-E), nano liposomes of $O$ . stamineus ethanolic extract (OS-EL) at 250 mg/kg/day and captopril at 5 mg/kg/day. The negative controls of 0.5% carboxymethyl cellulose (CMC), lecithin and water (10 ml/kg/day), were administered to rats in the control groups. The results are expresses as mean $\pm$ S.E.M (n = 6)	239
Figure 4.48	Body weight changes of Normotensive Wistar Kyoto rats (WKY) during 28-day treatment with <i>O. stamineus</i> ethanolic extract (OS-E), nano liposomes of <i>O. stamineus</i> ethanolic extract (OS-EL) at 250 mg/kg/day and captopril at 5 mg/kg/day. The negative controls of 0.5% carboxymethyl cellulose (CMC), lecithin and water (10 ml/kg/day), were administered to rats in the control groups. No significant differences were detected between the treated and control (vehicle 10 ml/kg) groups. All values are expressed as the mean $\pm$ S.E.M. (n = 6)	239
Figure 4.49	Effect of <i>O. stamineus</i> ethanolic extract (OS-E) and nano liposomes of <i>O. stamineus</i> ethanolic extract (OS-EL) on (A) HRGEC and (B) EAHY 926 cell lines viability. All extracts inhibit proliferation of HRGEC and EAHY 926 cell lines in dosedependent manner after 48 hours treatment. Values are means of three experiments (n=3). Error bars indicate $\pm$ S.E.M	242
Figure 4.50	Electropherogram of (A) RNA 6000 Nano ladder contains six RNA fragments ranging in size from 0.2 to 6 kb (0.2 kb, 0.5 kb, 1.0 kb, 2.0 kb, 4.0 kb, and 6.0 kb) at a total concentration of 150 ng/μL, (B) RNA isolated from HRGEC treated with <i>O. stamineus</i> ethanolic extract, (C) RNA extracted from HRGEC treated with nano liposomes of <i>O. stamineus</i> ethanolic extract, (D) RNA extracted from HRGEC treated with PBS (negative control), (E) RNA extracted from EAHY 926 cells treated with <i>O. stamineus</i> ethanolic extract, (F) RNA extracted from EAHY 926 cells treated with nano liposomes of <i>O. stamineus</i> ethanolic extract and (G) RNA extracted from EAHY 926 cells treated with PBS (negative control)	244
Figure 4.51	(A) traditional PCA plot and (B) matrix PCA plot. The analysis was performed on all samples, and on the top 50 microRNAs with the highest standard deviation. The normalized log ratio values were used for the analysis	247
Figure 4.52	Heat Map and Unsupervised Hierarchical Clustering was performed on on the top 50 microRNAs. The normalized log ratio values have been used for the analysis	248
Figure 4.53	The volcano plot shows the relation between the logarithm of the p-values and the log fold change between HR and EA groups	249

Figure 4.54 Mechanism of hsa-miR-149-3p and hsa-miR-21-3p miRNAs function in regulating blood pressure

### LIST OF ABREVIATIONS

A Pre-exponential factor

AAS Atomic absorption spectroscopy

ACE Angiotensin converting enzyme

ACE-I Angiotensin converting enzyme inhibitor

AlCl<sub>3</sub> Aluminium chloride

ALP Alkaline phosphatase

ALT Alanine aminotransferase

ANOVA Analysis of variance

A.P.T.T Activated partial thromboplastin time

As Arsenic

AST Aspartate aminotransferase

ARBs Angiotensin II receptor blockers

AT-I Angiotensin I

AT-II Angiotensin II

ATR Attenuated total reflection

AT-R1 Angiotensin II type I receptor

ATRs Angiotensin receptors

AUC Area under plasma concentration-time curve

BSA Bovine serum albumin

C Concentration

Cd Cadmium

CL Clearance

C<sub>max</sub> Maximum concentration

CMC Carboxymethyl cellulose

CO<sub>2</sub> Carbon dioxide

DAD Diode array detector

DBP Diastolic blood pressure

DMEM Dulbecco's modified eagle medium

DMSO Dimethyl sulfoxide

DNA Deoxyribonucleic acid

DPPH 2,2-Diphenyl-1- picrylhydrazil

Ea Activation energy

EA-1 RNA isolated from EAHY 926 cells treated with

Orthosiphon stamineus ethanolic extract

EA-2 RNA isolated from EAHY 926 cells treated with nano

liposomes of Orthosiphon stamineus ethanolic extract

EAHY 926 Human umbilical vein Cell

EA-NC RNA isolated from EAHY 926 cells treated with PBS

EC<sub>50</sub> Half maximal effective concentration

ECGS Endothelial cell growth supplement

ECM Endothelial cell medium

EDTA Ethylenediaminetetraacetic acid

EUP Eupatorin

FBS Fibrinogen and fetal bovine serum

FT-IR Fourier transform infra-red

FT-NIR Fourier transform near-infrared

g Relative centrifugal force or g-force

g Gram

g/kg Gram per kilogram

GGT Gamma-glutamyl transferase

h Hour

H<sub>3</sub>PO<sub>4</sub> Orthosphoshoric acid

HA Hippuric acid

Hb Hemoglobin

HCA Hierarchical clustering analysis

HCl Hydrochloric acid

HDL High-density lipoproteins

Hg Mercury

HHL Hippuryl-histidyl-leucine

HI FBS Heat inactivated foetal bovine serum

HL Histidyl-leucine

HNO<sub>3</sub> Nitric acid

HPLC High performance liquid chromatography

HPTLC High performance thin layer chromatography

HR Heart rate

HR-1 RNA isolated from HRGEC treated with *Orthosiphon* 

stamineus ethanolic extract

HR-2 RNA isolated from HRGEC treated with nano liposomes of

Orthosiphon stamineus ethanolic extract

HRGEC Human renal glomerular endothelial cell

HR-NC RNA isolated from HRGEC treated with PBS

HUVEC Human umbilical vein endothelial cell

IC<sub>50</sub> Half maximal inhibitory concentration

ICH International conference on harmonization

IV Intravenous

J Joule

kb kilobases

KBr Potassium bromide

KCl Potassium chloride

K<sub>e</sub> Elimination rate constant

L Litre

LD<sub>50</sub> Lethal dose

LOD Limit of detection

log Logarithm

LOQ Limit of quantification

LPS Lipopolysaccharide

LSD Least significant difference

M Molar

MAP Mean arterial pressure

MCH Mean corpuscular haemoglobin

MCHC Mean corpuscular hemoglobin concentration

MCV Mean corpuscular volume

MDA Malondialdehyde

MDA-MB-231 Human hormone resistant breast cancer cell line

mg Milligram

mg/g Milligram per gram

mg/kg Milligram per kilogram

MIC Minimum inhibitory concentration

min Minutes

miRNA Micro ribonucleic acid

mL Millilitre

MLT Microbial limit test

mm Millimetre

mM Mill molar

mm<sup>3</sup> Cubic millimetre

MRC Methylripariochromene A

mRNA Messenger ribonucleic acid

MTT 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium

bromide

mU Milli unit

mV Milli volt

n Number of sample

N Normal

Na<sub>2</sub>CO<sub>3</sub> Sodium carbonate

NaCl Sodium chloride

ng Nanogram

nm Nanometre

NO Nitric oxide

OECD Organisation for economic cooperation and development

OPA *O*-Phthaladehyde

OS Orthosiphon stamineus

OS-E Orthosiphon stamineus ethanolic extract

OS-EL Nano liposomes of *Orthosiphon stamineus* ethanolic extract

OS-EW Orthosiphon stamineus 50% ethanolic extract

OS-M Orthosiphon stamineus methanolic extract

OS-MW Orthosiphon stamineus 50% methanolic extract

OS-W Orthosiphon stamineus water extract

Pb lead

PBS Phosphate buffered saline

PCA Principle component analysis

PCR Polymerase chain reaction

PCV Packed cell volume

PDI polydispersity index

PE Plating efficiency

pH Power of hydrogen

Plt Platelet count

PP Pulse pressure

ppm Part per million

PS Penicillin/streptomycin

PT/I.N.R Prothrombin time and international normalized ratio

R<sup>2</sup> Regression correlation coefficient

RA Rosmarinic acid

RAAS Rennin-angiotensin-aldosterone-system

RBC Red blood cells

RDW Red cell distribution width

RH Relative humidity

RIN RNA integrity number

RISC RNA-induced silencing complex

RNA Ribonucleic acid

rpm Round per minutes

RSD Relative standard deviation

RVSEB Rappaport vassiliadis salmonella enrichment broth

S.E.M Standard error of the mean

S/N Signal to noise ratio

SbGTT Subcutaneous glucose tolerance test

SBP Systolic blood pressure

SD Standard deviation

SD Sprague–Dawley rats

Sec Second

SGOT Glutamate oxaloacetate transaminase

SGPT Glutamate pyruvic transaminase

SHR Spontaneously hypertensive rats

SHRSP Stroke-prone spontaneously hypertensive rats

SIN Sinensetin

SPL Soybean phospholipid

t<sub>1/2</sub> Half-life

t<sub>90</sub> Shelf life

TEM Transmission electron microscopy

T<sub>max</sub> Time to reach to maximum concentration

TMF 3'-hydroxy-5,6,7,4'-tetramethoxyflavone

TMM Tetramethylmurexide

UV/Vis Ultraviolet-visible

v/v Volume per volume

V<sub>d</sub> Volume of distribution

WBC White blood cells

WHO Word health organization

WKY Normotensive Wistar Kyoto

Zn Zinc

ZnCl<sub>2</sub> Zinc chloride

### LIST OF SYMBOLS

 $\Delta G$  Gibbs free energy

 $\lambda_{max} \hspace{1.5cm} Lambda \hspace{1mm} max \hspace{1mm} or \hspace{1mm} maximum \hspace{1mm} absorption$ 

 $\mu g \hspace{1cm} \text{Micro gram}$ 

μg/mL Micro gram per millilitre

μL Microliter

μm Micrometre

°C Degree Celsius

°T Temperature in kelvin

% Percent

# CIRI-CIRI ANTIHYPERTENSIF EKSTRAK TERPIAWAI DAUN ORTHOSIPHON STAMINEUS BENTH. DAN NANO LIPOSOMNYA TERHADAP TIKUS HYPERTENSIF SPONTAN

### **ABSTRAK**

Kajian ini dijalankan untuk memenuhi jurang antara amalan herba peribumi dan sains perubatan kontemporari ke atas kesan antihipertensi daun *Orthosiphon stamineus* (OS). Kajian kualiti dan keselamatan bahan mentah tumbuhan diperiksa menggunakan analisis gravimetrik dan ujian had mikrob (MLT). Daun OS didapati memenuhi kualiti dari segi fizikokimia serta pencemaran mikrob. Analisis spektroskopi kualitatif (UV, FT-IR, FT-NIR), kromatografi (HPTLC) dan kromatografi kuantitatif (HPLC) telah dijalankan terhadap ekstrak OS yang berlainan untuk pemiawaian. Hasil kajian menunjukkan bahawa bahan kimia utama dalam ekstrak OS adalah fenolik dan flavonoid seperti asid rosmarinik (RA), 3-hidroksi-5,6,7,4-metoksiflavon (TMF), sinensetin (SIN) dan eupatorin (EUP). Tambahan lagi, teknik HPLC-DAD gradien yang digabungkan dengan pengekstrakan fasa pepejal telah dibangunkan dan disahkan untuk pengenalpastian dan pengkuantitian 17 asid amino bebas dalam ekstrak OS. Hasil kajian menunjukkan bahawa asid L-aspartik dan asid L-glutamik adalah asid amino bebas utama dalam ekstrak OS dengan  $0.93 \pm 0.01$ nmol/mg dan  $4.01 \pm 0.12$  nmol/mg. Metabolit primer dan sekunder ekstrak OS telah dianalisis untuk menentukan jumlah flavonoid, polifenol, fosfolipid, protein, polisakarida dan glikosaponin. Perbezaan peratusan metabolit ini dalam setiap ekstrak telah ditunjukkan. Ekstrak OS yang berlainan dan sebatian piawai (RA, TMF, SIN dan EUP) telah dinilai dalam assai perencatan enzim penukaran angiotensin (ACE-I) secara in vitro. Hasil kajian menunjukkan bahawa OS-E dan EUP, pada kepekatan 50

μg/mL mempamerkan perencatan tertinggi (masing-masing pada 52.67 ± 0.89 dan 73.11 ± 2.39%) terhadap ACE berbanding dengan ekstrak dan sebatian piawai lain. Captopril telah digunakan sebagai kawalan positif dan menunjukkan perencatan sebanyak  $86.14 \pm 2.98\%$  pada kepekatan 6.8 ng/mL. Keupayaan mengkelat  $Zn^{2+}$  oleh RA, TMF, SIN, EUP, captopril dan OS-E telah dijalankan menggunakan reagen tetrametilmureksida (TMM). Hasil kajian menunjukkan OS-E dan captopril mempunyai keupayaan tinggi (79.42  $\pm$  1.91 dan 100  $\pm$  1.59) untuk mengikat dengan Zn<sup>2+</sup> pada kepekatan 5 mg/mL. Antara sebatian piawai yang diuji, EUP menunjukkan keupayaan tertinggi dalam mengikat  $Zn^{2+}$  (56.03 ± 1.26%) pada kepekatan 5 mg/mL. Tambahan lagi, skor cantuman dan afiniti ikatan bagi sebatian penanda Zn<sup>2+</sup> pada ACE telah dinilai. Hasil kajian menunjukkan bahawa EUP mempunyai tenaga afiniti pengikatan (ΔG) dan kecekapan ligan tertinggi dengan -6.93 kcal/mol. Seterusnya, ekstrak OS-E telah disediakan dalam formulasi liposom menggunakan lesitin soya nyah minyak (OS-EL) dan dicirikan melawan parameter berbeza. OS-EL dicirikan melawan parameter yang berbeza. Hasil kajian menunjukkan penghasilan liposom nano daripada OS-E. Farmakokinetik dan bioavailabiliti oral bagi RA, TMF, SIN dan EUP dalam OS-E dan OS-EL dikaji ke atas tikus Sprague Dawley (SD) menunjukkan peningkatan signifikan dalam keterlarutan akueus dan bioavailabiliti oral bagi RA, TMF, SIN dan EUP dengan masing-masing pada  $70.64 \pm 3.87$ ,  $66.26 \pm 5.95$ ,  $76.61 \pm$ 3.99 and 81.39  $\pm$  2.46% dalam OS-EL berbanding OS-E. OS-E dan OS-EL kemudiannya dikaji untuk aktiviti antihipertensi in vivo ke atas tikus SHR pada dos 250 mg/kg/hari selama 28 hari. Captopril digunakan sebagai kawalan positif pada dos 5 mg/kg/hari. Selain itu, aktiviti ACE-I ekstrak-ekstrak ini dalam plasma dan tisu SHR yang berbeza juga diukur selepas 28 hari rawatan melalui kaedah HPLC-UV yang dibangunkan. Keputusan menunjukkan bahawa, kedua-dua OS-E dan OS-EL mampu

menurunkan SBP (-23.08  $\pm$  5.16 dan -28.96  $\pm$  6.65) dan aktiviti ACE dalam plasma, jantung, paru-paru dan buah pinggang SHR secara signifikan. aorta. Walaubagaimanapun, OS-EL menunjukkan kesan antihipertensi yang lebih kuat (dan aktiviti penindasan ACE) berbanding OS-E. Kajian kestabilan dipercepatkan bagi OS-E dan OS-EL yang disimpan pada empat suhu yang berbeza (30, 40, 50 dan 60 ° C) selama enam bulan telah dilakukan dengan menggunakan HPLC dan spektroskopi FT-IR yang digabungkan dengan pendekatan kemometrik. Berdasarkan hasil kajian, sebatian penanda (RA, TMF, SIN dan EUP) dalam OS-E dan OS-EL adalah lebih stabil pada suhu rendah (30°C dan ke bawah). Kajian genotoksisiti akut dan ketoksikan subkronik OS-EL menunjukkan pengambilan OS-EL tidak menyebabkan kematian, tanda-tanda keracunan atau fungsi fisiologi dalam kedua-dua jantina haiwan. Data yang diperolehi daripada analisis pengekspresan miRNA menunjukkan bahawa dua miRNA; hsa-miR-149-3p dan hsa-miR-21-3p telah diekspres secara berbeza (kawal selia menaik) dalam HRGEC yang dirawat dengan OS-E dan OS-EL. miR-149 dan miR-21 mampu mensasarkan gen ACE dan AGTR1 secara langsung dan bertindak sebagai pengatur gen negatif. Oleh itu, boleh dicadangkan bahawa, OS boleh bertindak sebagai perencat ACE (ACE-I) dan penghalang reseptor angiotensin (ARBs). Sebagai kesimpulan, kajian ini memberikan bukti ke atas aktiviti antihipertensi, hubungan struktur aktiviti bagi sebatian penanda, kualiti dan keselamatan ekstrak terpiawai OS.

# ANTIHYPERTENSIVE PROPERTIES OF STANDARDISED ORTHOSIPHON STAMINEUS BENTH. LEAVES EXTRACTS AND ITS NANO LIPOSOMES IN SPONTANEOUS HYPERTENSIVE RATS

#### **ABSTRACT**

This study was conducted to fulfill gaps between indigenous herbal practices and contemporary medicinal sciences on antihypertensive effect of Orthosiphon stamineus (OS) leaves. Quality and safety of the plant raw material were examined using gravimetric analysis and microbial limit test (MLT). OS leaves were found to be qualified in terms of physicochemical properties as well as microbial contamination. Spectroscopic (UV, FT-IR, FT-NIR) and chromatographic (HPTLC and HPLC) analyses were carried out on different extracts of OS for standardisation. The results showed that the major chemical constituents in OS extracts are phenolics and flavonoids such as rosmarinine acid (RA), 3-hydroxy-5, 6, 7, 4-methoxyflavone (TMF), sinensetin (SIN) and eupatorin (EUP). Furthermore, a gradient HPLC-DAD combined with solid-phase extraction technique was developed and validated for identification and quantification of 17 free amino acids in OS extracts. The results demonstrated that L-aspartic acid with  $0.93 \pm 0.01$  nmol/mg and L-glutamic acid with  $4.01 \pm 0.12$  nmol/mg are the major free amino acid in OS extracts. Primary and secondary metabolites of OS extracts were analysed for their total flavonoids, polyphenols, phospholipids, proteins, polysaccharides and glycosaponins. The variation in the percentage of these metabolites in each extracts was indicated. Different extracts of OS and standard compounds (RA, TMF, SIN and EUP) were evaluated for in vitro assay of angiotensin converting enzyme inhibition (ACE-I). The results showed that, at final concentration of 50 µg/mL OS-E and EUP exhibit the

highest inhibition (52.67  $\pm$  0.89 and 73.11  $\pm$  2.39%, respectively) against ACE among extracts and tested standard compounds. Captopril was used as positive control and showed 86.14 ± 2.98% inhibition at concentration of 6.8 ng/mL. Chelation ability of Zn<sup>2+</sup> by RA, TMF, SIN, EUP, captopril and OS-E was conducted using tetramethylmurexide (TMM) reagent. The results demonstrated that OS-E and captopril have a high ability (79.42  $\pm$  1.91 and 100  $\pm$  1.59%, respectively) to bind with Zn<sup>2+</sup> at concentration of 5 mg/mL. Among the standard compounds, EUP shows the highest binding ability with  $Zn^{2+}$  (56.03  $\pm$  1.26 %) at concentration of 5 mg/mL. In addition, the docking scores and binding affinities of marker compounds with Zn<sup>2+</sup> in ACE were evaluated. The results showed that EUP has the highest binding affinity energy (ΔG) and ligand efficiency with -6.93 kcal/mol. Subsequently, OS-E extract was prepared in liposomal formulation using deoiled soya lecithin (OS-EL) and characterized versus different parameters. The results revealed production of nano liposomes of OS-E. The pharmacokinetics and oral bioavailability of RA, TMF, SIN and EUP in OS-E and OS-EL on Sprague Dawley (SD) rats indicated significant improvement in aqueous solubility and oral bioavailability of RA, TMF, SIN and EUP with  $70.64 \pm 3.87$ ,  $66.26 \pm 5.95$ ,  $76.61 \pm 3.99$  and  $81.39 \pm 2.46\%$ , respectively in OS-EL compared to OS-E. OS-E and OS-EL were then studied for their in vivo antihypertensive activity on Spontaneous Hypertensive Rats (SHR) at dose of 250 mg/kg/day for 28 days. Captopril was used as positive control at dose of 5 mg/kg/day. Moreover, ACE-I activity of these extracts in plasma and different tissues of SHR after 28 days treatment was also measured using developed HPLC-UV method. The results demonstrated that both OS-E and OS-EL were able to reduce SBP (-23.08  $\pm$  5.16 and -28.96 ± 6.65) and ACE activity in plasma, aorta, heart, lung and kidney of SHR significantly. However, OS-EL showed stronger antihypertensive effect (and suppression of ACE activity) than OS-E. The accelerated stability studies of OS-E and OS-EL stored at four different temperatures (30, 40, 50 and 60°C) for six months were performed using HPLC and FT-IR spectroscopy combined with chemometric approach. Based on the study findings, the marker compounds (RA, TMF, SIN and EUP) in both OS-E and OS-EL were more stable at lower temperature (30°C and below). The genotoxicity, acute and repeated dose oral toxicity study of OS-EL showed that administration of OS-EL does not cause death, visible signs of toxicity or other physiological functions in any animals of both sexes. The data obtained from miRNA expression analysis revealed that two miRNA; hsa-miR-149-3p and hsa-miR-21-3p are expressed differently (up regulated) in HRGEC treated with OS-E and OS-EL. miR-149 and miR-21 are able to directly target ACE and AGTR1 genes, respectively and act as negative gene regulators. It is suggested that OS could act ac ACE inhibitor (ACE-I) and angiotensin receptor blockers (ARBs). In conclusion, the current study provides evidence on antihypertensive activity, structure activity relationship of the active marker compounds, quality and safety of the standardised OS extract.

# CHAPTER 1 INTRODUCTION

#### 1.1 Herbal Medicines and Hypertension

Herbal products as sources of medicine have been used for many years practically in all cultures. Consumption of herbal products has been increased significantly over the last decade (Ekor, 2014). This is probably because herbs are remarked as natural and therefore they are safe to use. Plants as an important source of new drugs development are still used, despite the extensive developments in synthetic chemistry. Several well-known medicines derived from plants include L-hyoscyamine, morphine, colchincine, taxol and digitoxin. Traditional medicines and herbal products for the primary health care are used by more than 80% of world population and mostly in developing countries (Hussain et al., 2009). In a period of 1983-1994 in North America, about 40% of the new drugs were derived from natural compounds (Simmonds, 2003). Interestingly, over 70% of the new reported chemical in 1981-2006 were from the study of natural products (Newman and Cragg, 2007). Promoting and encouraging of the use of herbal products and remedies are recommended by World Health Organization (WHO) in the National Health Care Program (Hussain et al., 2009).

Herbal products refer to organic chemicals that might come from a single plant or combination of more than one plant, from any raw or processed part of plant such as stems, leaves, roots, flowers and seeds. However, if they are combined with synthetic chemicals or other active substances, they are not considered as herbal medicines (WHO, 1996). Due to the presence of molecules and products that combat diseases in plants, they have played an important role in maintaining health. Many

products derived from plants in the form of dried plant materials, fresh or extracts are used as folk remedies. Different compounds derived from plant can be used as starting material for preparation of novel synthetic drugs. The potential of herbs as a source of new drugs has not been explored yet and only a limited number of plant species among 250,000 have been investigated for their bioactive compounds (Borchardt, 2002)

Traditional medicine has a number of proven benefits for prevention and cure of different ailments. Application of modern medicines along with traditional medicines made a strong comeback of herbs in many countries in the last decade (WHO, 1998). Appreciation of natural remedies might be due to the increment of the cost of treatment with modern medicines and fear of their side effects, which represents alternative healthcare movement. Due to this reason, the demand for herbal products has increased tremendously in the world market especially among young generation.

The development of spectroscopic methods for the elucidation of natural compound structure together with development of biological sciences have opened a new era to study structure activity relationship. These developments have allowed for preparing derivatives or synthetic analogues using natural compounds as model.

Hypertension or high blood pressure is one of the most important concerns in developed countries. It causes heart to work harder to maintain high blood pressure. Moreover, it contributes to atherosclerosis (hardening of arteries), besides increasing the risk of heart disease and stroke. Local plants, from different countries such as *Agathosma betulina* from Rutaceae family, *Allium sativum* from Alliaceae or Liliaceae family, *Annona muricata* from Annonaceae family, *Apium graveolens* from Apiaceae family, *Aristolochia manshuriensis* from Aristolochiaceae family, *Artocarpus altilis* 

from Moraceae family, *Coleus forskohlii* from Lamiaceae family, have been widely used with hypotensive and antihypertensive therapeutic values to reduce high blood pressure (Tabassum and Ahmad, 2011). In Malaysia, *Orthosiphon stamineus* plant from Lamiaceae family have been used traditionally to cure hypertension (Perry, 1998). However, scientific data to support its effect is not available. Therefore, more research coupled with modern medicine needs to be carried out to verify its effectiveness, and elucidate the safety profile of such herbal remedies for their hypotensive/antihypertensive potential.

#### 1.2 Standardisation of Herbal Medicine

Standardisation is prescribing a set of standards or inherent characteristics, constant parameters, definitive qualitative and quantitative values that carry an assurance of quality, efficacy, safety and reproducibility. It is the process of developing and agreeing upon technical standards. Specific standards work out by experimentation and observations, which would lead to the process of prescribing a set of characteristics exhibited by a particular herbal medicine. Hence, standardisation is a tool in the quality control process.

Several problems that are not applicable to synthetic drugs often influence the quality of herbal drugs. For instance: 1) herbal drugs are usually mixture of many constituents, 2) the active principle(s) is (are), in most cases unknown, 3) selective analytical methods or reference compounds may not be available commercially, 4) plant materials are chemically and naturally variable, 5) Chemo-varieties and chemo cultivars exist, 6) the source and quality of the raw materials are variable. Moreover, the light exposure, temperature, nutrients, use of fresh plants, age, part of the collected plant, water availability, period and time of collection, method of collecting, drying,

packing, storage, transportation, contamination with microorganism, heavy metals or pesticides, and processing (for example, mode of extraction and polarity of the extracting solvent, instability of constituents) might impact the quality, safety and efficacy of the herbal drugs (Calixto, 2000). In spite of many proven benefits of natural products, they cannot be widely accepted in the main stream of pharmaceuticals due to lack of standardisation. Therefore, it is necessary to provide scientific evidence on standardisation to support their efficacy and bring these remedies into the mainstream pharmaceutical market (Barnes, 2003).

Due to the long-history of the use of herbal products in various cultures, they are usually considered safe. However, series of harmful effects after consumption of herbal products have been reported including direct toxic effect especially because of presence of heavy metal, allergic reactions, and mutagenic effects. In addition, the toxic effects of herbal preparation may be attributed to inherent toxicity of plant constituents, ingredients, manufacturing malpractice, and contamination (Mosihuzzaman and Choudhary, 2008). Therefore, standardisation and quality control of the raw materials and herbal preparation are necessary to be carried out.

Numbers of International Pharmacopoeia such as British Pharmacopoeia and United States Pharmacopeia, which contained collection of recommended procedures such as macroscopic and microscopic examination, determination of total ash, acid-insoluble ash and water-soluble ash, determination of pesticide residue, determination of swelling and foaming index, limit test for heavy metals, limit test for microorganisms, and test for determination of extractable matter, water and volatile matterfor specifications and quality control of plant raw materials, have been conducted by WHO (WHO, 1973). Moreover, many international authorities and

agencies including the European Agency for Evaluation of Medicinal Products and the European Scientific Cooperation of Phytomedicine, The US Agency for Healthcare Policy, and Research the European Pharmacopoeia Commission have started creating a new mechanism for quality control and standardisation of botanical medicines (Sharma et al., 2010).

#### 1.3 Justification of the Research

In this study, Orthosiphon stamineus (OS) a local plant was selected for studying its antihypertensive effect. O. stamineus known as misai kucing is a medicinal plant grown in South East Asia and currently cultivated in Indonesia and Malaysia. In Malaysia, the leaves of this plant (misai kucing) have been used traditionally in treating angiogenesis related diseases, urinary lithiasis, edema, inflammation, eruptive fever, influenza, hepatitis, jaundice, rheumatism, diabetes and hypertension (Mukesh et al., 2015; Perry, 1998). Recent scientific findings also showed that O. stamineus have the potential for different pharmacological properties. Although a number of products manufactured from O. stamineus are available in the market, there is still lack of information in terms of chemical components related to primary metabolites such as content of specific amino acids. The basis for the traditional use of this herb as antihypertensive and structure activity relationship has not yet been scientifically verified. Furthermore, a new step in development of new generation of standardised herbal medicine is preparation of botanical formulation to increase the solubility and bioavailability of the active constituents with therapeutic activity. In addition, microRNAs, as candidates for diagnostic and prognostic biomarkers and predictors of this herb response, have not yet been investigated. Therefore, this research aims to fulfil the gaps between indigenous herbal practices and contemporary medicinal sciences.

#### 1.4 General Objectives

This study generally seeks to standardise *O. stamineus* leaves extracts by developing new analytical techniques to measure the content of primary and secondary metabolites in *O. stamineus* extracts. Moreover, it aims to demonstrate the *in vitro* and *in vivo* antihypertensive properties of standardised *O. stamineus* extracts based on structure activity relationship of marker compounds. In addition, it seeks to prepare new formulation from standardised *O. stamineus* extract using soy bean phospholipids in order to improve the solubility and bioavailability of the active constituents with therapeutic activity. It also aims to utilize expression of miRNA subsets as a new tool to elucidate the mechanism of plant in treatment of hypertension.

## 1.5 Specific Objectives

- 1) To standardise *Orthosiphon stamineus* extracts using selected markers by developing new analytical methods.
- 2) To evaluate antihypertensive properties of the various standardised extracts of Orthosiphon stamineus based on structure activity relationship of marker compounds.
- 3) To prepare nano liposomes of the most active extract in order to improve antihypertensive activity, solubility and bioavailability of the active marker compounds.
- 4) To determine pharmacokinetic, stability, acute and sub chronic toxicity studies of the nano liposomes of *Orthosiphon stamineus* extract.
- To investigate the expression of miRNA subsets as potential biomarkers for antihypertensive activities.

#### 1.6 Hypotheses

O. stamineus has been used in traditional medicine to treat hypertension. Subsequently, presence of high content of phenolics and flavonoids compounds such as sinensetin (SIN), eupatorin (EUP), 3'-hydroxy-5, 6, 7, 4'-tetramethoxy flavone (TMF) and rosmarinic acid (RA) were detected in O. stamineus extract in previous studies. Whereas, inhibition activity of angiotensin converting enzyme (ACE) has been attributed to the presence of flavonoids in the plant extract, due to the generation of chelate complexes within the active centre of ACE. Therefore, possible research hypothesis would be that there is a correlation between the presence of these compounds and antihypertensive properties of the plant.

## 1.7 Significance of Study

The findings of this study provide knowledge on application of analytical methods for standardisation of plant materials and extract to produce, safe and high quality herbal medicinal products for manufacturers and consumers. Moreover, this research fills the gaps between indigenous herbal practices and contemporary medicinal sciences on antihypertensive effect of medicinal plant. Additionally, this study is a significant endeavour in promoting the use of natural phospholipid bilayer obtained from food grade soybean lecithin to prepare new botanical formulation to improve the extract's solubility and permeability as the major factors for improving oral bioavailability.

# 1.8 Methodology Flowchart

The overall methodology consists of many steps, which includes quality test of raw material by gravimetric analysis, extractive value, heavy metals and microbial limit test. The plant raw materials will be extracted by maceration method to prepare water (OS-W), ethanolic (OS-E), methanolic (OS-M), 50% ethanolic (OS-EW) and 50% methanolic (OS-MW) extracts. All extracts will be standardised by different spectroscopic and chromatographic techniques (UV-Vis, FT-IR, FT-NIR, HPTLC and HPLC). Moreover, the contents of primary and secondary metabolites also will be quantified in the extracts. The molecular docking study will be carried out on the marker compounds of extracts to determine the docking scores and binding affinities of marker compounds with Zn<sup>2+</sup> in angiotensin converting enzyme. Then in vitro angiotensin converting enzyme inhibitory (ACE-I) assays including enzymatic ACE-I assay and chelating activity of Zn<sup>+2</sup> will be studied on extract and marker compounds. Two extracts with highest ACE-I activity will be screened for *in vivo* antihypertensive properties on Spontaneous Hypertensive Rats (SHR) in order to select the best extract with highest antihypertensive activity. The most active extract will be formulated using soy been phospholipids in order to improve the solubility and bioavailability of the active constituents with therapeutic activity. Subsequently, the nano formulated extract will be characterized for different parameters. Moreover, pharmacokinetic, stability and toxicology studies will be done on nano formulated extract. Then, in vivo antihypertension activity of nano formulated extract will be studied on SHR. At the end, miRNA expression study will be done to investigate the expression of miRNA subsets as potential biomarkers for antihypertensive activities. The methodology of this study is summarized in the following flowchart (Figure 1.1).

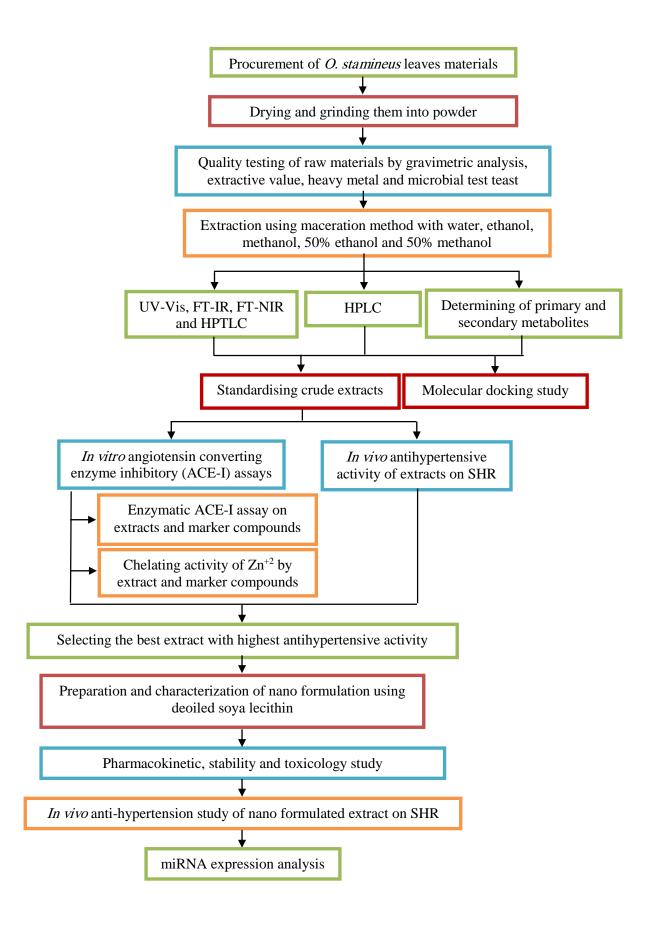


Figure 1.1: Methodology flowchart

#### **CHAPTER 2**

#### LITERATURE REVIEW

#### 2.1 Orthosiphon stamineus

## 2.1.1 Taxonomy

Taxonomically, this plant is classified as the following scheme:

Family Lamiaceae

Genus Orthosiphon

Scientific name *Orthosiphon stamineus*. (Benth)

Local name Misai Kucing

Synonyms O. aristatus (Bl.); O. grandiflorus, Bold., O. spicatus

Common name Java tea, cat's whiskers

(Comprehensive information about *Orthosiphon stamineus* plant can be found at http://www.ars-grin.gov/cgi-bin/npgs/html/taxon.pl?423487)

Orthosiphon stamineus Benth is a well-known medicinal plant belonging to Lamiaceae family. This plant is native in Southeast Asian countries. It is a perennial herb, 25-200 cm tall with quadrangular, poorly ramified and ascending stem. The plant is herbaceous shrub and it can be found in tropical and sub tropical regions. The stem is acutely quadrangular, reddish in colour, erect and branches profusely. The leaves are simple, green and glabrous with a lanceolate leaf blade and serrate margin. The leaves are arranged in opposite pairs and the petiole is short about 1.3 cm in length and reddish purple in colour. The flowers are hermaphrodite, about 6.2 cm in length including the staments, with very irregular flower symmetry (Almatar et al., 2013). In South East Asia, this plant is known as misai kucing (Malaysia), kumis kucing and

remujung (Indonesia) and yaa nuat maeo (Thailand). In Malaysia, the local name of this plant refers to the white or blue colour of flowers with long filaments over midgreen foliage which makes the flower look like cat's whiskers in Malay Misai (whiskers) kucing (cat) (Figure 2.1).

# 2.1.2 Ethnopharmacology

*O. stamineus* has been traditionally used for treating ranges of diseases such as edema, inflammation, urinary, lithiasis, hepatitis, rheumatism, eruptive fever, diabetes, influenza, jaundice, as a remedy for kidney stones and nephritis, pain in the bladder with frequent urination, diuretic, biliary and hypertension (Awale et al., 2001; Dat et al., 1992; Goh et al., 1995; Tezuka et al., 2000). Leaves of this plant in Southeast Asian and European countries are used popularly as herbal tea, known as "Java tea".



Figure 2.1: Pictures of *Orthosiphon stamineus* leaves and flower

# 2.2 Review of Chemical Constituents of *Orthosiphon stamineus*

Phytochemically, the plant is rich in flavonoids especially polymethoxylated flavone, terpenes, diterpenoids and triterpenes such as hydroxyl betulinic acid, betulinic acid, oleanolic acid, ursolic acid and caffeic acid derivatives like rosmarinic acid (Sumaryono et al., 1991). The chemical constituents and chemical structure identified in the aerial parts of *O. stamineus* are illustrated in Table 2.1 and Table 2.2.

Table 2.1: Chemical constituents of *Orthosiphon stamineus* 

Class of compounds	Part of plant	<b>Chemical constituents</b>	Reference
Diterpenes	Aerial	Orthisiphols F [1], orthosiphols G [2], orthosiphols H [3], orthosiphol I [4], orthosiphol J [5], orthosiphol S [6], staminols A [7], staminols B [8], staminolactones A [9], staminolactones B [10], norstaminol A [11], orthosiphonone A [12], orthosiphonone B [13]	(Awale et al., 2001; Tezuka et al., 2000)
Triterpenes	Aerial	Oleanolic acid [14], ursolic acid [15], betulinic acid [16], β-sitosterol [17]	(Tezuka et al., 2000)
Flavones	Aerial	7,3',4'-tri-O-methylluteolin [18], eupatorin [19], sinensetin [20], 3'-hydroxy-5,6,7,4'-tetramethoxyflavone [21], salvigenin [22], ladanein [23], scutellarein tetramethyl ether [24], 6-hydroxy-5,7,4'-trimethoxyflavone [25], kaempferol-3-O-β-glucoside [26], quercetin-3-O-β-glucoside [27]	(Sumaryono et al., 1991; Tezuka et al., 2000)
Phenolic acids	Leaves	Caffeoyl tartrate [28], rosmarinic acid [29], aurantiamide acetate [30], vomifoliol [31], caffeic acid [32], 2,3-dicaffeoyl tartrate [33]	(Sumaryono et al., 1991)

Table 2.1: Continued

Benzochromene	Leaves	Methylripariochromene A [34], acetovanillochromene [35], orthochromene A [36]	(Shibuya et al., 1999)

Number in brackets indicate the number of the structure

Table 2.2: Chemical structures of *Orthosiphon stamineus* 

Orthosiphol F [1]	Orthosiphol G [2]	Orthosiphol H [3]
Orthosiphol I [4]	Orthosiphol J [5]	Orthosiphol S [6]

Table 2.2: Continued

Table 2.2: Continued

	HO HO OH	HO HO OH
Orthosiphonone B [13]	Oleanolic acid [14]	Ursolic acid [15]
HO HO OH	HO HO	OH O
Betulinic acid [16]	β-sitosterol [17]	7,3',4'-tri- <i>O</i> -methylluteolin [18]

Table 2.2: Continued

OH O		ОН
Eupatorin [19]	Sinensetin [20]	3'-hydroxy-5,6,7,4'-tetramethoxyflavone
		[21]
OH O	HO OH O	
Salvigenin [22]	Ladanein [23]	Scutellarein tetramethyl ether [24]

Table 2.2: Continued

6-Hydroxy-5,7,4'-trimethoxyflavone [25]	OH OOH OOH OOH OOH OOH OOH OOH OOH OOH	OH O
HO OH O OH O OH O OH OH OH OH OH OH OH O	HO OH OH Rosmarinic acid [29]	Aurantiamide acetate [30]