SYNTHESIS, CHARACTERIZATION AND BIOLOGICAL EVALUATION OF NAPHTHALENE- AND COUMARIN-BASED CHALCONES

by

SUMITHDA LIM YEONG HUI

Thesis submitted in fulfillment of the requirements for the degree of

Master of Science

March 2016

ACKNOWLEDGMENT

I am grateful to many people who have assisted me to bring this thesis into reality. Foremost, I am highly thankful to my supervisor, Dr. Oo Chuan Wei for his guidance, comments, motivation, forgiveness and his unwavering support throughout the period of my candidature. I also would like to express my appreciation to Dr. Quah Ching Kheng (School of Physical Sciences) and Dr. Yam Mun Fei (School of Pharmaceutical Sciences) for their assistance and dedication in helping me accomplishing this project.

I wish to take this opportunity to thank Universiti Sains Malaysia (USM) for providing me the scholarship (Graduate Assistant Scheme) and giving me the chance to enroll as an MSc student in USM. I would like to thank School of Chemical Sciences, USM for providing me with the research facilities and research funding from TWAS Research Grant (12-312RG/PHA/AS_C). My gratitude also goes to the staff of School of Chemical Sciences especially to Mr. Megat, Mr. Zahari, Mr. Ong Chin Hin and Mr. Chow Cheng Por for their great assistance in my research. I would also like to thank Ms. Loh Wan Sin for her help in my XRD studies. Last but not least, I wish to convey my best regards and gratitude to my beloved parent, Ms. Lim Gaik Peng and my sisters, who have always gives me such amazing support. Thanks for always encouraging me in my interests and teaching me about values and respects.

TABLE OF CONTENTS

		Page
Ackn	owledgement	ii
Table	e of Contents	iii
List o	of Tables	vii
List o	of Figures	xii
List o	of Abbreviations	xxi
Abstı	ak	xxii
Absti	act	xxiv
СНА	PTER 1: INTRODUCTION	
1.1	Chalcone	1
1.2	Problem Statement	4
1.3	Objectives of the research	4
1.4	Scope of Study	. 5
СНА	PTER 2: LITERATURE REVIEW	
2.1	Staring materials.	6
	2.1.1 Coumarin.	6
	2.1.2 Naphthalene.	8
2.2	Methods to synthesize chalcone	. 10
	2.2.1 Claisen-Scmidt reaction.	11
	2.2.2 Suzuki reaction.	. 12
	2.2.3 Heck reaction.	13
2.3	Bioactivities of chalcones	14

CHAPTER 3:EXPERIMENTAL

3.1	Chemicals	20
3.2	Instruments	21
3.3	Synthesis	
	3.3.1 Preparation of 2-alkyloxy-1-naphthaldehyde, 2-8	22
	3.3.2 Preparation of 3-acetyl-4-hydroxycoumarin, 9	23
	3.3.3 Preparation of 3-acetylcoumarin, 10	24
	3.3.4 Preparation of 3-acetyl-6,7-dimethyl-4-hydroxycoumarin,	11
		24
	3.3.5 Preparation of (E)-1-(4-hydroxy-1-benzopyran-2-one-3-yl)	-3-
	(2-alkoxy naphthyl)-prop-2-en-1-one, 12-19	25
	3.3.6 Preparation of (E)-1-(1-benzopyran-2-one-3-yl)-3-(2-alkox	.y
	naphthyl)-prop-2-en-1-one, 20-26	26
	3.3.7 Preparation of (E)-1-(6,7-dimethyl-4-hydroxy-1-benzopyra	ın-
	2-one-3-yl)-3-(2-alkoxynaphthyl)-prop-2-en-1-one, 27-33 .	. 27
	3.3.8 Preparation of (E)-1-(2-hydroxyphenyl)-3-(2-alkoxynaphth	ıyl)-
	prop-2-en-1-one, 34-39	28
	3.3.9 Preparation of (E)-1-(4-fluro-2-hydroxyphenyl)-3-(2-alkox	y
	naphthyl)-prop-2-en-1-one, 40-46	29
3.4	X-ray crystallographic	31
3.5	Evaluation of anti-cancer activity	31

CHAPTER 4: RESULTS AND DISCUSSION

4.1	2-Alk	yloxy-1-naphthaldehyde, 2-8	33
	4.1.1	Fourier transform infrared spectroscopy (FT-IR)	34
	4.1.2	Fourier transform nuclear magnetic resonance spectr	oscopy
		(FT-NMR)	37
	4.1.3	X-ray crystallography analysis	43
4.2	(E)-1-	(4-hydroxy-1-benzopyran-2-one-3-yl)-3-(2-alkoxynap	hth-1-
	yl) -p	rop-2-en-1-one, 12-19	48
	4.2.1	Fourier transform infrared spectroscopy (FT-IR)	49
	4.2.2	Fourier transform nuclear magnetic resonance spectr	oscopy
		(FT-NMR)	51
4.3	(E)-1-	(1-benzopyran-2-one-3-yl)-3-(2-alkoxynaphth-1-yl)-p	rop-2-
	en- 1-	-one, 20-26	69
	4.3.1	Fourier transform infrared spectroscopy (FT-IR)	70
	4.3.2	Fourier transform nuclear magnetic resonance spectr	oscopy
		(FT-NMR)	72
	4.3.3	X-ray crystallography analysis	89
4.4	(E)-1-	(6,7-dimethyl-4-hydroxy-1-benzopyran-2-one-3-yl)-3-	-(2-
	alkox	ynaphth-1-yl)-prop-2-en-1-one, 27-33	95
	4.4.1	Fourier transform infrared spectroscopy (FT-IR)	96
	4.4.2	Fourier transform nuclear magnetic resonance spectr	oscopy
		(FT-NMR)	98
	4.4.3	X-ray crystallography analysis	114

4.5	(E)-1-	(2-hydroxyphenyl)-3-(2-alkoxynaphth-1-yl)-prop-2-en-1-
one, 3	4-39	121
	4.5.1	Fourier transform infrared spectroscopy (FT-IR)122
	4.5.2	Fourier transform nuclear magnetic resonance spectroscopy
		(FT-NMR)124
	4.5.3	X-ray crystallography analysis141
4.6	(E)-1-	(4-fluro-2-hydroxyphenyl)-3-(2-alkoxynaphth-1-yl)-prop-2-
	en-1-0	one, 40-46 147
	4.6.1	Fourier transform infrared spectroscopy (FT-IR)148
	4.6.2	Fourier transform nuclear magnetic resonance spectroscopy
		(FT-NMR)150
	4.6.3	X-ray crystallography analysis167
4.7	Anti-c	ancer Activity
CHAPTER 5: CONCLUSION 179		
5.1	Recon	nmendations for future studies
REFERENCES		
A DDE	NDIY	192

LIST OF TABLES

		Page
Table 2.1	Biological applications of chalcones and their derivatives.	15
Table 3.1	List of chemicals used for this study.	21
Table 3.2	Scientific name and respective R group of 2-8 .	24
Table 3.3	Scientific name and respective R group of 12-19.	27
Table 3.4	Scientific name and respective R group of 20-26 .	28
Table 3.5	Scientific name and respective R group of 27-33 .	29
Table 3.6	Scientific name and respective R group of 34-39 .	30
Table 3.7	Scientific name and respective R group of 40-46 .	31
Table 4.1	CHN micro analytical data of 2-8.	34
Table 4.2	Physical appearance, molecular formula, molecular weights, percentage yields and melting points of 2-8 .	35
Table 4.3	Selected FT-IR wavenumbers, v/cm^{-1} and relative intensities of 2-8 .	36
Table 4.4	¹ H-NMR chemical shifts, δ /ppm of 2-8.	41
Table 4.5	¹³ C-NMR chemical shifts, δ/ppm of 2-8 .	43
Table 4.6	Crystal and structure refinemental data of 3.	46

Table 4./	Atomic coordinates and equivalent isotropic displacement parameters (\mathring{A}^2) of 3 .	47
Table 4.8	Bond lengths (Å), bond angles (°) and torsion angles (°) of $\bf 3$.	48
Table 4.9	CHN micro analytical data of 12-19.	49
Table 4.10	Physical appearance, molecular formula, molecular weights, percentage yields and melting points of 12-19.	49
Table 4.11	Selected FT-IR wavenumbers, v/cm^{-1} and relative intensities of 12-19.	50
Table 4.12	¹ H-NMR chemical shifts, δ/ppm of 12-19 .	57
Table 4.13	¹³ C-NMR chemical shifts, δ/ppm of 12-19 .	61
Table 4.14	¹ H- ¹ H correlations as inferred from 2D COSY experiment for 14 .	69
Table 4.15	¹ H- ¹³ C correlations as inferred from 2D HMQC and HMBC experiments for 14 .	69
Table 4.16	CHN micro analytical data of 20-26 .	70
Table 4.17	Physical appearance, molecular formula, molecular weights, percentage yields and melting points of 20-26 .	70
Table 4.18	Selected FT-IR wavenumbers, v/cm ⁻¹ and relative intensities of 22-26 .	71
Table 4.19	¹ H-NMR chemical shifts, δ/ppm of 20-26 .	77

Table 4.20	¹³ C-NMR chemical shifts, δ/ppm of 20-26 .	81
Table 4.21	¹ H- ¹ H correlations as inferred from 2D COSY experiment for 23 .	89
Table 4.22	¹ H- ¹³ C correlations as inferred from 2D HMQC and HMBC experiments for 23 .	89
Table 4.23	Crystal and structure refinemental data of 23.	92
Table 4.24	Atomic coordinates and equivalent isotropic displacement parameters (\mathring{A}^2) of 23 .	93
Table 4.25	Bond lengths (Å), bond angles (°) and torsion angles (°) of 23.	94
Table 4.26	CHN micro analytical data of 27-33.	96
Table 4.27	Physical appearance, molecular formula, molecular weights, percentage yields and melting points of 27-33 .	96
Table 4.28	Selected FT-IR wavenumbers, v/cm^{-1} and relative intensities of 27-33.	97
Table 4.29	¹ H-NMR chemical shifts, δ/ppm of 27-33 .	102
Table 4.30	¹³ C-NMR chemical shifts, δ/ppm of 27-33 .	106
Table 4.31	¹ H- ¹ H correlations as inferred from 2D COSY experiment for 28 .	114
Table 4.32	¹ H- ¹³ C correlations as inferred from 2D HMQC and HMBC experiments for 28 .	114

Table 4.33	Crystal and structure refinemental data of 28.	118
Table 4.34	Atomic coordinates and equivalent isotropic displacement parameters (\mathring{A}^2) of 28 .	119
Table 4.35	Bond lengths (Å), bond angles (°) and torsion angles (°) of 28 .	120
Table 4.36	CHN micro analytical data of 34-39 .	122
Table 4.37	Physical appearance, molecular formula, molecular weights, percentage yields and melting points of 34-39 .	122
Table 4.38	Selected FT-IR wavenumbers, v/cm^{-1} and relative intensities of 34-39 .	123
Table 4.39	¹ H-NMR chemical shifts, δ/ppm of 34-39 .	129
Table 4.40	¹³ C-NMR chemical shifts, δ /ppm of 34-39 .	133
Table 4.41	¹ H- ¹ H correlations as inferred from 2D COSY experiment for 34 .	141
Table 4.42	¹ H- ¹³ C correlations as inferred from 2D HMQC and HMBC experiments for 34 .	141
Table 4.43	Crystal and structure refinemental data of 34 .	144
Table 4.44	Atomic coordinates and equivalent isotropic displacement parameters (\mathring{A}^2) of 34 .	145
Table 4.45	Bond lengths (Å), bond angles (°) and torsion	146

angles (°) of 34.

Table 4.46	CHN micro analytical data of 40-46 .	148
Table 4.47	Physical appearance, molecular formula, molecular weights, percentage yields and melting points of 40-46 .	148
Table 4.48	Selected FT-IR wavenumbers, v/cm ⁻¹ and relative intensities of 40-46 .	149
Table 4.49	¹ H-NMR chemical shifts, δ/ppm of 40-46 .	154
Table 4.50	¹³ C-NMR chemical shifts, δ/ppm of 40-46 .	158
Table 4.51	¹ H- ¹ H correlations as inferred from 2D COSY experiment for 41 .	166
Table 4.52	¹ H- ¹³ C correlations as inferred from 2D HMQC and HMBC experiments for 41 .	166
Table 4.53	Crystal and structure refinemental data of 41.	169
Table 4.54	Atomic coordinates and equivalent isotropic displacement parameters (\mathring{A}^2) of 41 .	170
Table 4.55	Bond lengths (Å), bond angles (°) and torsion angles (°) of 41.	171
Table 4.56	IC ₅₀ values of the test samples against breast cancer cell, 4T1 and fibroblast cell, NIH-3T3.	176
Table 4.57	Percentage of 4T1 cell viability after treated with potential selected compounds and cisplatin for 72 hours.	179

LIST OF FIGURES

		Page
Figure 1.1	General structure of chalcone.	1
Figure 1.2	Chemical structure of 1-(4-hydroxyphenyl)-3-(2',4'-dihydroxyphenyl)-2- propen-1-one labeled with universal numbering system.	2
Figure 1.3	Chalcone as an intermediate in biosynthetic pathways leading to different classes of flavonoids.	3
Figure 2.1	Structure of a coumarin.	5
Figure 2.2	Chemical structures of some naturally occurring coumarins with potent anticancer activity.	6
Figure 2.3	Structure of naphthalene.	7
Figure 2.4	Chemical structure of 6-hydroxy-4-(4-hydroxy-3-methoxyphenyl)-3-hydroxymethyl-7-methoxy-3,4-dihydro-2-naphthaldehyde extracted from seeds of Vitex Negundo.	8
Figure 2.5	Synthesis of various substituted chalcones in the presence of water and hydroxyapatite as catalyst via Claisen–Schmidt condensation using microwave irradiation.	10
Figure 2.6	Proposed reaction mechanism for the chalcone formation under alkaline condensation in 65% EtOH-H2O by Gasull et al. (2000).	11
Figure 2.7	Synthesis of chalcones by Suzuki coupling between cinnamonyl chloride and phenylboronic acid.	12
Figure 2.8	The reaction scheme by Tsutomo Mizoroki (1971).	13

Figure 2.9	The reaction scheme by Richard F. Heck (1972).	13
Figure 2.10	Chalcone synthesis by Heck coupling reaction.	14
Figure 3.1	Synthetic scheme for the formation of 2-alkyloxy-1-naphthaldehyde, 2-8 .	23
Figure 3.2	Synthetic scheme for the formation of 3-acetyl-4-hydroxycoumarin, 9 .	24
Figure 3.3	Synthetic scheme for the formation of 3-acetyl coumarin, 10.	25
Figure 3.4	Synthetic scheme for the formation of 3-acetyl-6,7-dimethyl-4-hydroxycoumarin, 11 .	25
Figure 3.5	Synthetic scheme for the formation of (E) -1-(4-hydroxy-1-benzopyran-2-one-3-yl)-3-(2-alkoxynaphthyl)-prop-2-en-1-one, 12-19 .	26
Figure 3.6	Synthetic scheme for the formation of (E) -1-(1-benzopyran-2-one-3-yl)-3-(2-alkoxynaphthyl)-prop-2-en-1-one, 20-26 .	27
Figure 3.7	Synthetic scheme for the formation of (E) -1- $(6,7$ -dimethyl-4-hydroxy-1-benzopyran-2-one-3-yl)-3- $(2$ -alkoxynaphthyl)-prop-2-en-1-one, 27-33 .	28
Figure 3.8	Synthetic scheme for the formation of (E) -1- $(2$ -hydroxyphenyl)-3- $(2$ -alkoxynaphthyl)-prop-2-en-1-one, 34-39 .	29
Figure 3.9	Synthetic scheme for the formation of (<i>E</i>)-1-(4-fluro-2-hydroxyphenyl)-3-(2-alkoxynaphthyl)-prop-2-en-1-one, 40-46 .	30
Figure 4.1	FT-IR spectrum of 2-butoxy-1-naphthaldehyde 3	37

Figure 4.2	Structure of compound 3 with complete atomic numbering.	38
Figure 4.3	¹ H NMR spectrum of 2-butoxy-1-naphthaldehyde, 3 .	40
Figure 4.4	¹³ C NMR spectrum of 2-butoxy-1-naphthaldehyde, 3 .	42
Figure 4.5	Molecular structure with atomic numbering scheme for 2-butoxy-1-naphthaldehyde, 3 .	44
Figure 4.6	Molecular packing of 3, showing the intra- and intermolecular bonding.	45
Figure 4.7	FT-IR spectrum of (<i>E</i>)-1-(4-hydroxy-1-benzopyran-2-one-3-yl)-3-(2-butoxynaphth-1-yl)-prop-2-en-1-one, 14 .	51
Figure 4.8	Structure of compound 14 with complete atomic numbering.	52
Figure 4.9	¹ H -NMR spectrum of (<i>E</i>)-1-(4-hydroxy-1-benzopyran-2-one-3-yl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 14 .	56
Figure 4.10	¹³ C -NMR spectrum of (<i>E</i>)-1-(4-hydroxy-1-benzopyran-2-one-3-yl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 14 .	59
Figure 4.11	DEPT-135 NMR spectrum of (<i>E</i>)-1-(4-hydroxy-1-benzopyran-2-one-3-yl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 14 .	60
Figure 4.12	COSY spectrum of (<i>E</i>)-1-(4-hydroxy-1-benzopyran-2-one-3-yl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 14 .	63

Figure 4.13	COSY (expand) spectrum of (<i>E</i>)-1-(4-hydroxy-1-benzopyran-2-one-3-yl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 14 .	64
Figure 4.14	HMQC spectrum of (<i>E</i>)-1-(4-hydroxy-1-benzopyran-2-one-3-yl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 14 .	65
Figure 4.15	HMQC (expand) NMR spectrum of (<i>E</i>)-1-(4-hydroxy-1-benzopyran-2-one-3-yl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 14 .	66
Figure 4.16	HMBC spectrum of (E) -1- $(4$ -hydroxy-1-benzopyran-2-one-3-yl)-3- $(2$ -butoxynaphthyl)-prop-2-en-1-one, 14 .	67
Figure 4.17	HMBC (expand) NMR spectrum of (<i>E</i>)-1-(4-hydroxy-1-benzopyran-2-one-3-yl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 14 .	68
Figure 4.18	FT-IR spectrum of (E) -1-(1-benzopyran-2-one-3-yl)-3-(2-octoxynaphth-1-yl)-prop-2-en-1-one, 23 .	72
Figure 4.19	Structure of compound 23 with complete atomic numbering.	73
Figure 4.20	The resonance structure of 23 , which greatly affect the electron cloud of C14 atom.	75
Figure 4.21	1 H -NMR spectrum of (<i>E</i>)-1-(1-benzopyran-2-one-3-yl)-3-(2-octoxynaphthyl)-prop-2-en-1-one, 23 .	76
Figure 4.22	13 C -NMR spectrum of (<i>E</i>)-1-(1-benzopyran-2-one-3-yl)-3-(2-octoxynaphthyl)-prop-2-en-1-one, 23 .	79
Figure 4.23	DEPT-135 NMR spectrum of (<i>E</i>)-1-(1-benzopyran-2-one-3-yl)-3-(2-octoxynaphthyl)-prop-2-en-1-one, 23 .	80

Figure 4.24	COSY spectrum of (E) -1- $(1$ -benzopyran-2-one-3-yl)-3- $(2$ -octoxynaphthyl)-prop-2-en-1-one, 23 .	83
Figure 4.25	COSY (expand) NMR spectrum of (<i>E</i>)-1-(1-benzopyran-2-one-3-yl)-3-(2-octoxynaphthyl)-prop-2-en-1-one, 23 .	84
Figure 4.26	HMQC spectrum of (E) -1- $(1$ -benzopyran-2-one-3-yl)-3- $(2$ -octoxynaphthyl)-prop-2-en-1-one, 23 .	85
Figure 4.27	HMQC (expand) NMR spectrum of (<i>E</i>)-1-(1-benzopyran-2-one-3-yl)-3-(2-octoxynaphthyl)-prop-2-en-1-one, 23 .	86
Figure 4.28	HMBC spectrum of (E) -1- $(1$ -benzopyran-2-one-3-yl)-3- $(2$ -octoxynaphthyl)-prop-2-en-1-one, 23 .	87
Figure 4.29	HMBC (expand) NMR spectrum of (<i>E</i>)-1-(1-benzopyran-2-one-3-yl)-3-(2-octoxynaphthyl)-prop-2-en-1-one, 23 .	88
Figure 4.30	Molecular structure with atomic numbering scheme for (E) -1- $(1$ -benzopyran-2-one-3-yl)-3- $(2$ -alkoxynaphthyl)-prop-2-en-1-one, 23 .	90
Figure 4.31	Molecular packing of 23, showing intra- and intermolecular bonds. H atoms not involved in hydrogen bonds (dashed lines) have been omitted for clarity.	91
Figure 4.32	FT-IR spectrum of (E) -1- $(6,7$ -dimethyl-4-hydroxy-1-benzopyran-2-one-3-yl)-3- $(2$ -butoxynaphth-1-yl)-prop-2-en-1-one, 28 .	98
Figure 4.33	Structure of compound 28 with complete atomic numbering.	99

Figure 4.34	¹ H- NMR spectrum of (<i>E</i>)-1-(6,7-dimethyl-4-hydroxy-1-benzopyran-2-one-3-yl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 28 .	101
Figure 4.35	¹³ C- NMR spectrum of (<i>E</i>)-1-(6,7-dimethyl-4-hydroxy-1-benzopyran-2-one-3-yl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 28 .	104
Figure 4.36	DEPT-135 NMR spectrum of (<i>E</i>)-1-(6,7-dimethyl-4-hydroxy-1-benzopyran-2-one-3-yl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 28 .	105
Figure 4.37	COSY spectrum of (E) -1- $(6,7$ -dimethyl-4-hydroxy-1-benzopyran-2-one-3-yl)-3- $(2$ -butoxynaphthyl)-prop-2-en-1-one, 28 .	108
Figure 4.38	COSY (expand) NMR spectrum of (<i>E</i>)-1-(6,7-dimethyl-4-hydroxy-1-benzopyran-2-one-3-yl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 28 .	109
Figure 4.39	HMQC spectrum of (<i>E</i>)-1-(6,7-dimethyl-4-hydroxy-1-benzopyran-2-one-3-yl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 28 .	110
Figure 4.40	HMQC (expand) NMR spectrum of (<i>E</i>)-1-(6,7-dimethyl-4-hydroxy-1-benzopyran-2-one-3-yl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 28 .	111
Figure 4.41	HMBC spectrum of (<i>E</i>)-1-(6,7-dimethyl-4-hydroxy-1-benzopyran-2-one-3-yl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 28 .	112
Figure 4.42	HMBC (expand) NMR spectrum of (<i>E</i>)-1-(6,7-dimethyl-4-hydroxy-1-benzopyran-2-one-3-yl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 28 .	1113
Figure 4.43	Molecular structure with atomic numbering scheme for (<i>E</i>)-1-(6,7-dimethyl-4-hydroxy-1-benzopyran-2-one-3-vl)-3-(2-alkoxynaphthyl)-prop-2-en-1-one	115

Figure 4.44	Molecular packing of 28 , showing intra- and intermolecular bonds. H atoms not involved in hydrogen bonds (dashed lines) have been omitted for clarity.	116
Figure 4.45	Side view of molecule 28 with dihedral angle (73.60°) between two planes.	117
Figure 4.46	FT-IR spectrum of (E) -1- $(2$ -hydroxyphenyl)-3- $(2$ -butoxynaphth-1-yl)-prop-2-en-1-one, 34 .	124
Figure 4.47	Structure of compound 34 with complete atomic numbering.	125
Figure 4.48	Resonance structure of 34 within phenyl ring.	126
Figure 4.49	¹ H- NMR spectrum of (<i>E</i>)-1-(2-hydroxyphenyl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 34 .	128
Figure 4.50	13 C- NMR spectrum of (<i>E</i>)-1-(2-hydroxyphenyl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 34 .	131
Figure 4.51	DEPT-135 NMR spectrum of (<i>E</i>)-1-(2-hydroxyphenyl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 34 .	132
Figure 4.52	COSY spectrum of (E) -1- $(2$ -hydroxyphenyl)-3- $(2$ -butoxynaphthyl)-prop-2-en-1-one, 34 .	135
Figure 4.53	COSY (expand) NMR spectrum of (<i>E</i>)-1-(2-hydroxyphenyl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 34 .	136
Figure 4.54	HMQC spectrum of (E) -1- $(2$ -hydroxyphenyl)-3- $(2$ -butoxynaphthyl)-prop-2-en-1-one, 34 .	137

Figure 4.55	HMQC (expand) NMR spectrum of (<i>E</i>)-1-(2-hydroxyphenyl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 34 .	138
Figure 4.56	HMBC spectrum of (<i>E</i>)-1-(2-hydroxyphenyl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 34 .	139
Figure 4.57	HMBC (expand) NMR spectrum of (<i>E</i>)-1-(2-hydroxyphenyl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 34 .	140
Figure 4.58	Molecular structure with atomic numbering scheme for (<i>E</i>)-1-(2-hydroxyphenyl)-3-(2-alkoxynaphthyl)-prop-2-en-1-one, 34 .	142
Figure 4.59	Molecular packing of 34 , showing intra- and intermolecular bonds. H atoms not involved in hydrogen bonds (dashed lines) have been omitted for clarity.	143
Figure 4.60	FT-IR spectrum of (<i>E</i>)-1-(4-fluro-2-hydroxyphenyl)-3-(2-butoxynaphth-1-yl)-prop-2-en-1-one, 41 .	150
Figure 4.61	Structure of compound 41 with complete atomic numbering.	151
Figure 4.62	¹ H- NMR spectrum of (<i>E</i>)-1-(4-fluro-2-hydroxyphenyl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 41 .	153
Figure 4.63	¹³ C- NMR spectrum of (<i>E</i>)-1-(4-fluro-2-hydroxyphenyl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 41 .	156
Figure 4.64	DEPT-135 NMR spectrum of (<i>E</i>)-1-(4-fluro-2-hydroxyphenyl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 41 .	157

Figure 4.65	COSY spectrum of (<i>E</i>)-1-(4-fluro-2-hydroxyphenyl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 41 .	160
Figure 4.66	COSY (expand) NMR spectrum of (<i>E</i>)-1-(4-fluro-2-hydroxyphenyl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 41 .	161
Figure 4.67	HMQC spectrum of (E) -1- $(4$ -fluro-2-hydroxyphenyl)-3- $(2$ -butoxynaphthyl)-prop-2-en-1-one, 41 .	162
Figure 4.68	HMQC (expand) NMR spectrum of (<i>E</i>)-1-(4-fluro-2-hydroxyphenyl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 41 .	163
Figure 4.69	HMBC spectrum of (E) -1- $(4$ -fluro-2-hydroxyphenyl)-3- $(2$ -butoxynaphthyl)-prop-2-en-1-one, 41 .	164
Figure 4.70	HMBC (expand) NMR spectrum of (E) -1-(4-fluro-2-hydroxyphenyl)-3-(2-butoxynaphthyl)-prop-2-en-1-one, 41 .	165
Figure 4.71	Molecular structure with atomic numbering scheme for (E) -1- $(4$ -fluro-2-hydroxyphenyl)-3- $(2$ -alkoxynaphthyl)-prop-2-en-1-one, 41 .	167
Figure 4.72	Molecular packing of 41 , showing intra- and intermolecular bonds. H atoms not involved in hydrogen bonds (dashed lines) have been omitted for clarity.	168
Figure 4.73	Percentage of 4T1 cell viability at 12.5 μM by selected compounds and cisplatin.	178

LIST OF ABBREVIATION

¹³C-NMR Carbon nuclear magnetic resonance

¹H-NMR Proton nuclear magnetic resonance

4T1 Breast cancer cell

AcOH Acetic acid

COSY Correlation spectroscopy

DMF Dimethylformamide

DMSO Dimethylsulfoxide

EtOH Ethanol

FT-IR Fourier transform infrared

FT-NMR Fourier transform nuclear magnetic resonance

HMBC Heteronuclear multiple bond correlation

HMQC Heteronuclear multiple quantum correlation

IC₅₀ Inhibitory concentration at 50%

K₂CO₃ Potassium carbonate

MTT Dimethyl thiazolyl diphenyl tetrazolium

NIH-3T3 Fibroblast cell

POCl₃ Phosphorylchloride

SI Selectivity Index

SINTESIS, PENCIRIAN DAN PENILAIAN BIOLOGI KALKON BERASASKAN NAFTALENA DAN KUMARIN

ABSTRAK

Kajian ini dimulakan dengan sintesis dan pencirian dua siri utama bahan pemula: pengalkilan naftaldehid (2-8) dan pengasetilan kumarin tertukar ganti (9-11). Bahan pemula yang diperoleh dikenalpasti melalui kaedah spektroskopi seperti inframerah transformasi Fourier (FTIR), resonans magnet nukleus (NMR) dan mikroanalisis CHN. Analisa pembelauan sinar-X hablur tunggal (XRD) juga dijalankan dengan hablur bagi sebatian 3, 34, dan 41 menghablur dalam sistem kristal triklinik. Lima siri kalkon kemudiannya disintesis melalui kaedah kondensasi Claisen-Schmidt dengan tiga siri diterbitkan daripada kumarin asetil tertukar ganti (12-33) dan dua siri yang lain diterbitkan daripada asetofenon tertukar ganti (34-46). Semua kalkon yang disintesis telah dicirikan dengan analisa unsur (CHN), spektroskopi FTIR, ¹H-NMR, ¹³C-NMR, DEPT 135, ¹H-¹H COSY, ¹H-¹³C HMQC dan ¹H-¹³C HMBC. Kalkon 23, 28, 34, dan 41 telah dipilih untuk analisis kristolografi. Kalkon yang disintesis telah dinilai bagi ciri farmakologi (anti-kanser) terhadap sel kanser payudara (4T1) dan sel fibroblas (NIH - 3T3) dengan cisplatin sebagai kawalan positif. Berdasarkan data yang diperoleh, kalkon yang mengandungi gelang kumarin menunjukkan kesitotoksi yang lebih kuat terhadap sel kanser payudara berbanding dengan kaklon yang mengandungi gelang fenil biasa. Sebatian 17 menunjukkan aktiviti perencatan tertinggi terhadap titisan sel kanser 4T1 di kalangan kalkon yang disintesis dan lima kali ganda (IC₅₀ = 3 ± 0.00) lebih aktif berbanding dengan cisplatin (IC₅₀ = 16.5 ± 0.71). Kalkon 13, 14, 15, 16, 24 dan 31 juga menunjukkan aktiviti perencatan tinggi terhadap titisan sel kanser 4T1 dengan IC₅₀ < 20.5 μ M. Keputusan juga menunjukkan bahawa kalkon terfluorin (nilai IC₅₀ antara 28-62 μ M) mungkin menjadi ejen anti-kanser payudara yang lebih baik berbanding dengan kalkon tanpa fluorin (IC₅₀ > 92.5 μ M).

SYNTHESIS, CHARACTERIZATION AND BIOLOGICAL EVALUATION OF NAPHTHALENE- AND COUMARIN-BASED CHALCONES

ABSTRACT

This study was started by the synthesis and characterization of two main series of starting materials: alkylation of naphthaldehydes (2-8) and acetylation of substituted coumarins (9-11). The starting materials thus obtained were elucidated by the spectroscopic methods such as fourier transform infrared (FTIR), nuclear magnetic resonance (NMR) and CHN microanalyses. Single crystal X-ray diffraction (XRD) analyses have also been carried out wherein the crystals of compound 3, 34 and 41 crystallized in a triclinic crystal system. Five novel series of chalcones were subsequently synthesized by means of Claisen-Schmidt condensation wherein three series were derived from substituted acetyl coumarins (12-33) and the remaining two series was derived from substituted acetophenones (34-46). All the synthesized chalcones were characterized by elemental analysis (CHN), FTIR, ¹H-NMR, ¹³C-NMR, DEPT 135, ¹H-¹H COSY, ¹H-¹³C HMOC and ¹H-¹³C HMBC spectroscopy. Selected chalcones, 23, 28, 34, and 41 were selected for crystallography analysis. The synthesized chalcones were evaluated for their pharmacological (anti-cancer) properties against breast cancer cell (4T1) and fibroblast cell (NIH-3T3) with cisplatin as positive control. Based on the data obtained, chalcones comprised of coumarin rings showed more potent cytotoxicity against breast cancer cell compared to chalcones comprised of common phenyl rings. Compound 17 exhibited the highest inhibitory activity against 4T1 cancer cell line among the synthesized chalcones and five times (IC₅₀ = 3 ± 0.00) more active compared to cisplatin (IC₅₀ = 16.5 ± 0.71). Chalcones 13, 14, 15, 16, 24 and 31 also showed high inhibitory activities against 4T1 cancer cell line with IC₅₀ < $20.5~\mu$ M. The results also indicated that fluorinated chalcones (IC₅₀ value ranged from $28-62~\mu$ M) may serve as better anti-breast cancer agents than the chalcones without fluorine (IC₅₀ > $92.5~\mu$ M).

CHAPTER 1: INTRODUCTION

1.1 Chalcone

Chalcones, a class of naturally occurring pigments which also known as benzelideneacetophenones are aromatic ketones that form the central core for a variety of important biological compounds (Rachmale *et al.*, 2011). They are abundant in edible plants and are considered to be precursors for flavonoids biosynthesis in plants. Chalcones can also be synthesized in laboratory (Patil *et al.*, 2009). Flavanoids consist mostly of antocynin, flavones and flanols, while chalcone, dihydroxychalcone and aurone are known as minor classes of flavanoids (Harborne, 1967).

Chemically, chalcones are open chain flavonoids in which the two aromatic rings are joined together by three carbons from the α,β -unsaturated carbonyl system (enone), which is a typical 1,4-conjugate addition (Michael addition) acceptor (Ko *et al.*, 2003). The general chemical structure of a chalcone is depicted in Figure 1.1.

Figure 1.1: General structure of chalcone.

The term 'chalcone' also refers to the structurally simple and important plant secondary metabolite, which serves as defense mechanism in plants to counteract reactive oxygen species (ROS) to prevent damage by micro-organisms, insects or herbivores (Vaya *et al.* 1997). Chalcone strongly inhibits the polymerization of tubulin by binding to the colchicines-binding site (Lawrence *et al.*, 2006).

A universal numbering system has been applied to distinguish the two aromatic rings of chalcones in which the two rings are labeled as A and B respectively. Figure 1.2 shows the chemical structure of 1-(4-hydroxyphenyl)-3-(2',4'-dihydroxyphenyl)-2-propen-1-one, with the corresponding atomic numbering (Bohm, 1998).

Figure 1.2: Chemical structure of 1-(4-hydroxyphenyl)-3-(2',4'-dihydroxyphenyl)-2-propen-1-one labeled with universal numbering system.

Chalcone is also acted as an important intermediate in biosynthesis of other classes of flavonoids in natural plants. Figure 1.3 shows chalcone acts as a main center core structure that undergoes different biosynthetic pathways to produce various classes of flavonoids such as flavanone (I), flavon (II), auron (III), flavonol (IV), antocianidin (V), dihydroflavonol (VI), dihydrochalcone (VII) and isoflavone (VIII) (Wong, 1976). The conversion of chalcone to other classes of flavonoids is catalyzed by certain plant enzymes. For an example, the metabolic pathway continues through a series of enzymatic modifications to yield I from chalcones, where research showed chalcone-flavanone isomerase was the enzyme that enhanced the biosynthesis (Swain, 1976).

Figure 1.3: Chalcone as an intermediate in biosynthetic pathways leading to different classes of flavonoids.

In the recent years, synthesis of chalcones and their derivatives have attracted considerable attention due to their significant biological activities, sufficient fluorescence in the visible light range, large Stokes shifts and high quantum yields (Zhang *et al.*, 2010). Chalcones have been reported to display an impressive array of biological activities such as antioxidant, antibacterial, anthelmintic, amoebicidal, antiulcer, antiviral, insecticidal, antiprotozoal, antileishmanial, antimalarial, antifungal, anti-inflammatory, anticancer, cytotoxic and immunosuppressive (Nowakowska, 2006; Trivedi *et al.*, 2008; Srinivasan *et al.*, 2009).

1.2 Problem Statement

There were several basic catalysts that have been applied in the synthesis of chalcone, such as sodium hydroxide, sodium methoxide, barium hydroxide, hydroapatite and lithium hydroxide. Even though these reactions provide advantages due to their Heterogenous nature. However, most of them require rather complicated neutralization process to obtain the synthesized compound. Recently, to improve the production-in synthesis of chalcones, piperidine has been tried. In this study, piperidine mediated chalcones formation without the neutralization requirement to obtain the solid yield. Hence, it is among the suitable candidate to be used as support in chalcone synthesis in respect to its unique properties.

1.3 Objectives of the research

In view of the important and uses of chalcones and their derivatives, this research project is aims to undertake an in-depth study on a few series of new chalcones.

The objectives of this project are:

- To synthesize novel chalcones from inexpensive commercially available aromatic compounds.
- To synthesize novel coumarin- and naphthalene-based chalcone derivatives with different alkyl chain length.
- 3. To elucidate all intermediary and targeted compounds using spectroscopic techniques (FT-IR, 1D-NMR, 2D-NMR), CHN elemental analysis and X-ray crystallography.
- 4. To evaluate the anti-cancer activity of the synthesized chalcones.

1.4 Scope of Study

The scope of this study is basically to synthesize and characterize the chalcones using Claisen-Schmidt condensation method and to determine the anti-cancer activity for the chalcone synthesized. There are some significant tasks to be carried out in order to achieve the objective of this study.

- I. In this study, the individual starting material, coumarin and naphthalene will react to synthesized chalcones.
- II. The characterization of the result chalcones using fourier transform infrared (FTIR), nuclear magnetic resonance (NMR), CHN microanalyses and single crystal X-ray diffraction (XRD).
- III. Pharmacological (anti-cancer) properties of the synthesized chalcones will be evaluated through inhibition rate against breast cancer cell (4T1) and fibroblast cell (NIH-3T3) with cisplatin as positive control.
- IV. Analyst the relationship of the alkyl group carbon chain lengths with the synthesized chalcones biological activity.

CHAPTER 2: LITERATURE REVIEW

2.1 Starting materials

For this research project, two naturally abundant compounds had been chosen as starting materials: coumarin and naphthalene. Both are valuable starting materials that represent a vast family of compounds which were naturally found in plants and are interesting bioactive targets for many organic medicinal chemists.

2.1.1 Coumarin

Coumarin; also known as 2H-1-benzopyran-2-one, is a fragrant organic chemical compound and naturally occurring polyphenolics distributed widely in plants and microorganisms. Vogel (1920) had isolated the first coumarin (Figure 2.1) from the fruit of *Dipteryx odorata Wild*. Coumarin can occur in two basic forms, either attached to sugar glucose or as a free molecule.

Figure 2.1. Structure of a coumarin.

Due to large number of natural products contain coumarin heterocyclic as nucleus, coumarins occupy an important position in natural and synthetic organic chemistry. They are widely used as additives in food, perfumes, photochemotherapeutic agents, insecticides and in optical brighteners (Fulchand *et al.*, 2008). In particular, coumarin derivatives have been found to exhibit a variety of biological activities such as antitumor (Santana *et al.*, 2000), antioxidant (Guiotto *et al.*, 1995), anti-inflammatory

(Ploypradith *et al.*, 2004), anticoagulant (Borges *et al.*, 2005), antibacterial (Maria *et al.*, 2013), anti-cancer (Riveiro *et al.*, 2008; Belluti *et al.*, 2010), anti-HIV (Zhao *et al.*, 1997; Mahajan *et al.*, 2009), and antiproliferative (Jung *et al.*, 2009) properties. Among these properties, cytotoxic effects were most extensively examined. Neo-tanshinlactone (Figure 2.2), a coumarin containing compound, has shown significant inhibition activity against two ER+ human breast cancer cell lines, which has been associated with low toxicity and has raised considerable interest because of its potential beneficial effects on human health. Nevertheless, others coumarin-derivatives such as Geiparvarin, Umbeliferone, Xanthyletin and Seselin (Figure 2.2) have provoked great interest for their possible therapeutic uses (Sashidhara *et al.*, 2010).

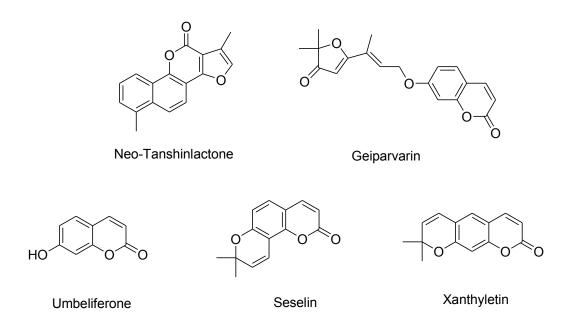


Figure 2.2. Chemical structures of some naturally occurring coumarins with potent anticancer activity.

Over the years, scientists had developed many strategic pathways for the synthesis of coumarins in laboratory. These include use of the Reformatsky, Wittig, Pechmann, Perkin, Claisen and Knoevenagel reactions. Knoevenagel reaction has been

applied in this project. The Knoevenagel reaction involves the condensation of benzaldehydes with activated methylene compounds and amine as a catalyst in organic solvents. Knoevenagel reaction method was more preferable compared to other methods because of its less severe reaction conditions (Knoevenagel *et al.*, 1984).

2.1.2 Naphthalene

Naphthalene is the simplest member of polycyclic aromatic hydrocarbons which consists of two fused benzene rings (Price & Jayjock, 2008). Naphthalene (Figure 2.3) appears as white crystalline solid with a characteristic odor and readily sublimes at room temperature. This compound is easily generated during burning of wood or cigarette, grilling of meats, petroleum refining and distillation of coal tar (ADSTR, 2005).

Figure 2.3. Structure of naphthalene.

Naphthalene has been largely used in manufacture as a chemical intermediate including moth repellents, plastics, wetting agent, lavatory scent discs, surfactant, dyes, resins, and soil fumigants (Brusick, 2008). Recent studies showed that naphthalene derivatives have attracted a great interest not only for their significant biological and photophysical properties (thermo- and photochromism, and non-linear optical behavior), but also as model compounds for assessing the nature of the hydrogen bonding (Martinez *et al.*, 2011).

Naphthalene and its derivatives have been reported to possess a wide spectrum of biological activity such as anti-inflammatory (Huang *et al.*, 2003; Jing *et al.*, 2013),

anti-cancer (Andrea *et al.*, 2012), anti-mycobacterial (Ram *et al.*, 2010; Upadhayaya *et al.*, 2010), anti-tubulin (Alvarez *et al.*, 2007), antimicrobial (Yapici *et al.*, 2005), anti-proliferative (Sheila *et al.*, 2013), anti-protozoal (Donald *et al.*, 2009), and bactericidal activity (Misra & Kushwaha, 1977). Study conducted by Tanaka *et al.* (2009) shows that anti-inflammatory potency of naphthalene-chalcone hybrid has been associated with the increase in HO-1 expression (Tanaka *et al.*, 2009). Additional to that, naphthalene has been shown as an ideal component of a fluorescent chemosensor due to its short fluorescence lifetime, low fluorescence quantum yield and ability to act as a donor as well as an acceptor (Sahana *et al.*, 2011).

Zhou and his co-workers (2009) had extracted a naphthaldehyde derivative characterized as 6-hydroxy-4-(4-hydroxy-3-methoxyphenyl)-3-hydroxymethyl-7-metho xy-3,4-dihydro-2-naphthaldehyde (Figure 2.4) from seeds of *Vitex negundo*, which proven to possess cytotoxic effect on breast, prostate, and ovarian cancer cells and induces apoptosis with cleavage in poly ADP ribose polymerase protein, up-regulation of Bax, and down-regulation of Bcl-2. Furthermore, Maioral *et al.* (2012) shown that synthetic chalcones, derived from 1-naphthaldehyde and 2-naphthaldehyde, possessed cytotoxic effect on human acute myeloid leukemia K562 cells and on human acute lymphoblastic leukemia Jurkat cells. From the experiment done by David (2004) also revealed that 2-hydroxy-1-naphthaldehyde derivatives showed higher activity and selectivity toward a range of neoplastic cells relative to normal cells.

Figure 2.4. Chemical structure of 6-hydroxy-4-(4-hydroxy-3-methoxyphenyl)-3-hydroxymethyl-7-methoxy-3,4-dihydro-2-naphthaldehyde extracted from seeds of Vitex Negundo.

2.2 Methods to synthesize chalcone

A chalcone can be prepared by Claisen-Schmidt condensation between a benzaldehyde and an acetophenone in the presence of base catalyst followed by dehydration to yield the corresponding product. It is also possible to synthesize chalcone under acidic conditions using perchloric acid and acetic acid (Siddiqui *et al.*, 2008). Several other methods have been developed as well to synthesize chalcone, such as Heck reaction (Bianco *et al.*, 2004), Hiyama coupling (Hatanaka & Hiyama, 1988), Negishi coupling (King *et al.*, 1977), Stille reaction (Santos *et al.*, 2007), Suzuki reaction (Suzuki, 1998) and so on.

2.2.1 Claisen-Schmidt reaction

Claisen-Schmidt reaction also known as aldol condensation has been the most convenient and widely used method for obtaining chalcones. It has been the most important enolate reactions of carbonyl compounds in which the acetophenone was reacted with benzaldehyde in the presence of alkali or acid as catalyst to form new carbon-carbon bonds. Condensation combined these two carbonyl compounds, often with the loss of small molecule such as water or an alcohol (Wade, 2006).

The condensation of the Claisen-Schmidt reaction can be carried out by using various basic catalysts such as sodium hydroxide (Alston & Albert, 2004), potassium hydroxide, sodium methoxide (Rojas *et al.*, 2002), barium hydroxide (Krohn *et al.*, 2002), hydroapatite and lithium hydroxide (Zahouily *et al.*, 2003). It is also possible to synthesize chalcone under acidic condition by using silica-sulphuric acid (Zhang *et al.*, 2003), dry hydrochloric acid, perchloric acid and acetic acid (Siddiqui *et al.*, 2008).

Solhy and his co-workers found out that hydroxyapatite was a very efficient heterogeneous catalyst for the preparation of chalcone derivatives via Claisen–Schmidt condensation using microwave irradiation (Figure 2.5). The catalyst was easily recovered and efficiently re-used. Solhy had combined hydroxyapatite with water (act as co-catalyst) to produce various substituted chalcones with excellent yield of 95% (Solhy *et al.*, 2010).

Figure 2.5: Synthesis of various substituted chalcones in the presence of water and hydroxyapatite as catalyst via Claisen–Schmidt condensation using microwave irradiation.

The mechanism of the Claisen-Schmidt reaction was proposed by Nayak and Rout in 1970. Gasull *et al.* (2000) has re-analyzed the mechanism of Nayak and proposed a new mechanism that has been used in most of the modern organic textbooks. As illustrated in Figure 2.6, there are several steps involved in the formation of chalcone:

- Step 1: A base removes an α proton to form an enolate ion.
- Step 2: Nucleophilic addition of the enolate ion to a carbonyl group.
- Step 3: A configurational equilibrium between *cis-s-cis* and *trans-s-trans* isomers.
- Step 4: Protonation of alkoxide by water molecule to form a neutral intermediate.
- Step 5: Intramolecular dehydration of the neutral intermediate to give the chalcone.

Figure 2.6: Proposed reaction mechanism for the chalcone formation under alkaline condensation in 65% EtOH-H₂O by Gasull *et al.* (2000).

2.2.2 Suzuki reaction

The Suzuki coupling reaction is the palladium-catalyzed cross coupling between organoboronic acid and aryl halides. Recent developments have extended the application of Suzuki reaction where the starting materials are not restricted to aryls, but include alkyls, alkenyls and alkynyls. Several reports have described the synthesis of polysubstituted chalcones based on the Suzuki reaction (Figure 2.7). The reaction has been performed using activated form of acid such as acyl chlorides, acid anhydrides or thio-esters coupling with phenylboronic acid (Eddarir et al., 2003).

Figure 2.7. Synthesis of chalcones by Suzuki coupling between cinnamonyl chloride and phenylboronic acid.

The advantages of this reaction are the substituents on the acryl chloride or on the boronic acid did not affect the reaction mechanism, the selection of coupling site was quit specified and unaffected by the presence of water. Other than that, the Suzuki coupling is applicable in laboratories as well as in industrial processes due to the inorganic by-product of the reaction is non-toxic and easily removed from the reaction mixture (Suzuki, 1998).

2.2.3 Heck reaction

The Heck reaction (also called the Mizoroki-Heck reaction) is a palladium catalyzed chemical reaction involves the coupling between aryl halide and aryl alkene in the presence of base (Heck & Nolley, 1972). This reaction was originated by Tsutomu Mizoroki (1971) which described the reaction between iodobenzene and styrene to form stilbene with potassium acetate base and palladium chloride catalyst (Figure 2.8).

Figure 2.8. The reaction scheme by Tsutomo Mizoroki (1971).

The reaction was further modified by Heck to eliminate two major difficulties found in Mizoroki reaction: difficulty in obtaining the required organomercury (lead or tin compounds) and the problem of working with thick slurries of salts, particularly if the reaction is carried out catalytically in palladium (Heck & Nolley, 1972). The improved reaction differs in lack of solvent, selection of catalyst and base used (Figure 2.9).

Figure 2.9. The reaction scheme by Richard F. Heck (1972).

Over the last few years, chalcones had been synthesized using the Heck coupling reaction. Palladium-catalyzed Heck coupling reaction between the aryl vinyl ketone with the aryl iodide will yield the corresponding chalcone (Figure 2.10) (Bianco *et al.*, 2004). This procedure affords the desired chalcone in yields that are

always higher than those using conventional methods such as Claisen-Schmidt condensation. It is possible to design various novel chalcone derivatives by monitoring the aryl halides or aryl vinyl ketones (Bianco *et al.*, 2003; Reichwald *et al.*, 2008).

Figure 2.10. Chalcone synthesis by Heck coupling reaction.

2.3 Bioactivities of chalcones

Cancer (also known as a malignant neoplasm) is a broad group of various diseases that involving uncontrolled, rapid and pathological proliferation of unregulated cell growth. Being the second leading cause of death in developing as well as advanced countries, it had increased the pressing need for new anticancer agents with high potency, less toxicity in non-cancerous cells, and unique targets of action (Chari *et al.*, 1992).

Chemotherapy had been the most common treatment for diseases especially by killing micro-organisms such as prokaryotes, eukaryotes and cancerous cells. The development of novel and efficient drugs by molecular manipulation of promising lead compounds is still a major line of approach. Following this aim hybrid molecules were designed from two natural occurring compounds: naphthalene and coumarin. Naphthalene and coumarin derivatives have well known pharmacological activities such as antimicrobial (Yapici *et al.*, 2005), anti-tubulin (Huang *et al.*, 2003), anti-mycobacterial (Upadhayaya *et al.*, 2010), antitumor, anti-inflammatory,

anti-HIV (Nohemi *et al.*, 2009), antithrombotic (Alvarez *et al.*, 2007), cardio protectors or enzymatic inhibitors (Park & Jang, 1995).

The hybrid molecules, chalcones, precursors of open chain flavonoids and isoflavonoids are present in edible plants, and have displayed a broad spectrum of pharmacological activity. Changes in their structure have offered a high degree of diversity that have proven useful for the development of new medicinal agents having improved potency and lesser toxicity (Rahman, 2011). The chemotherapeutic effects of various chalcones and their derivatives against a wide variety of microorganisms and diseases are given in Table 2.1.

Table 2.1: Biological applications of chalcones and their derivatives.

Compounds	Biological application	Reference
O OCH ₃	Inhibitors of malaria parasite (Chloroquine-sensitive <i>P. falciparum</i>)	Awasthi et al.
(HO) ₂ B N B(OH) ₂	Anti-cancer activity	Achanta et al.
НООНОН	Anti-inflammatory	Yadav et al.

OH O CH ₃	Anti-bacterial (Against Gram +ve bacteria, Staphylococcus aureus)	Hamdi <i>et al</i> .
O N=N OCH ₃	Anti-filarial (Setaria cervi using GST enzyme as a drug target)	Awasthi et al.
НООНСІ	Monoamine Oxidase (MAOs) inhibitory	Chimenti et al.
H ₃ CO ₃ S CH ₃	Cyclooxygenase (COX) inhibitory	Zarghi <i>et al</i> .
HO SCH ₃	Anti-oxidant	Vasil'ev et al.
OCH ₃	Anti-convulsant	Kaushik et al.

SCH ₃	Anti-fungal	Bag et al.
O N=N OCH ₃	Anti-filarial	Awasthi et al.

In 2000s, a broad series of various chalcones derivatives had been synthesized and evaluated for their pharmacological activities. Achanta *et al.* (2006) had reported the anticancer and the mechanism of action of eight boronic chalcones derivatives. From the test result, boronic chalcones shown potent growth inhibition activity in the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay and colony formation assay with IC₅₀ values of <1.5 μ M. Mechanistic studies showed that boronic chalcones induced significant cytotoxic effect in cancer cells by inhibitied the chymotrypsin like activity of the 20S proteasome *in vitro*.

Romagnoli *et al.* (2008) had synthesized a series of chalcone-like agents, in which the α,β unsaturated bond of the enone system is embedded within a thiophene ring were evaluated for antiproliferative activity and inhibition of tubulin assembly and colchicine binding to tubulin. The replacement of the double bond with a thiophene maintained antiproliferative activity (IC₅₀ < 2 μ M) and electron-releasing group (ERG) caused only minor changes in antiproliferative activity. The synthesized compounds were found to inhibit the growth of several cancer cell lines

at nanomolar to low micromolar concentrations and some compounds (IC₅₀, 0.8 μ M) having twice the potency of antitubulin agent CA-4 (IC₅₀, 1.4 μ M).

Echeverria *et al.* (2009) had reported the relationships between the structural characteristic of synthetic chalcones and their antitumoral activity toward hepatocellular carcinoma cells (HepG2). From the calculated reactivity indexes and the adiabatic electron affinities, it shown that the absence of methoxyl substituents has the major structure reactivity pattern along the series. Due to this, the chalcone 1-(2-hydroxyphenyl)-3-phenylprop-2-en-1-one was found to be possessed the most active structure-activity between the chalcone structure and the apoptosis in HepG2 cells.

Recently, another series of chalcones were synthesized and evaluated by Llango *et al.* (2010) for their *in vitro* cytotoxic activity by microculture Tetrazolium Test Assay method using two breast cancer cell lines MCF-7 and T47D. The synthesized chalcones shown significant cytotoxicity against both of the cell lines and values lied between 52-89 μ M. Due to the presence of nitro group in the compound, *N*-(4-hydroxy-3-(3-(4-nitrophenyl)acryloyl)phenyl)acetamide showed better activity than other compounds.

Szliszka *et al.* (2010) had synthesized five chalcones that hybrided with tumor necrosis factor-related apoptosis-inducing ligand (TRAIL). Results showed that all the five tested chalcones markedly augmented TRAIL-mediated apoptosis and cytotoxicity in prostate cancer cells and confirmed the significant role of chalcones in chemoprevention of prostate cancer. Their study suggested that chalcones help anticancer immune defense in which endogenous TRAIL takes part.

CHAPTER 3: EXPERIMENTAL

3.1 Chemicals

All the chemicals were used as received without further purification. Table

3.1 summarizes the chemicals used in this study.

Table 3.1: List of chemicals used in this study.

Chemicals	Source
1-Bromoethane	Merck
1-Bromobutane	Fluka
1-Bromohexane	Merck
1-Bromooctane	Merck
1-Bromodecane	Merck
1-Bromododecane	Merck
1-Bromotetradecane	Merck
2-Hydroxyacetophenone	Merck
2-Hydroxybenzaldehyde	Merck
2-Hydroxynaphthaldehyde	Fluka
3-Acetylcoumarin	Sigma-Aldrich
4-Fluoro-2-hydroxyacetophenone, 98%	Sigma-Aldrich
4-Hydroxycoumarin	Merck
Acetic acid (glacial)	Merck
Acetone	QRec
Deuterated chloroform, CDCl ₃ , with TMS (0.03 vol.%),	Merck
deuteration degree min. 99.8%	
Diethyl ether	QRec
Dimethylformamide (DMF)	QRec
Ethanol, 99.97%	QRec
Ethyl acetoacetate	Merck
Petroleum ether	QRec
Piperidine	Sigma-Aldrich
Phosphorylchloride	Merck
Potassium carbonate anhydrous	R & M Chemical
Sodium carbonate	Merck

3.2 Instruments

The instrumentation used for the qualitative characterization of synthesized starting materials and chalcone derivatives are listed below:

- Thin Layer Chromatography (TLC) analyses were carried out using aluminium-backed silica gel 60 F₂₅₄ plates. Visualization was done under UV light and by using iodine spotting.
- CHN microanalyses were carried out on a Perkin Elmer 2400 LS
 Series CHNS/O analyzer at the School of Chemical Sciences,
 University Sains Malaysia (USM).
- Fourier Transform Infrared (FT-IR) data were recorded using a Perkin Elmer 2000-FTIR spectrophotometer at the School of Chemical Sciences, USM.
- 1D and 2D Fourier Transform Nuclear Magnetic Resonance (FT-NMR) spectroscopy performed using a Bruker AC 500 MHz at the School of Chemical Sciences, USM.
- X-ray crystallographic analyses were done using either Bruker SMART Apex II or Apex II Duo CCD diffractometers at the School of Physical Sciences, USM.

3.3 Synthesis

The starting materials, aldehydes and ketones as well as the targeted chalcones were synthesized and purified at normal atmospheric pressure by conventional methods unless stated otherwise.

3.3.1 Preparation of 2-alkyloxy-1-naphthaldehydes, 2-8

2-Alkyloxy-1-naphthaldehydes were synthesized according to the synthetic scheme below (Figure 3.1).

$$\begin{array}{c} O \\ H \\ OH \end{array} \begin{array}{c} C_nH_{2n+1}Br, \ K_2CO_3 \\ \hline DMF, \ 60-70 \ ^{\circ}C \end{array} \begin{array}{c} O \\ H \\ OR \end{array}$$

$$\begin{array}{c} \textbf{2-8} \\ \\ R = C_2H_5, \ C_4H_9, \ C_6H_{13}, \ C_8H_{17}, \ C_{10}H_{21}, \ C_{12}H_{25}, \ C_{14}H_{29} \end{array}$$

Figure 3.1: Synthetic scheme for the formation of 2-alkyloxy-1-naphthaldehyde, **2-8**.

2-Hydroxy-1-naphthaldehyde (2.00 g, 11.62 mmol) was mixed with 1.5 equiv. of the corresponding bromoalkane in *N,N*-dimethylformamide (20.0 mL), which served as a solvent. Three equiv. of potassium carbonate anhydrous was then added to the solution and the mixture was refluxed for 12 hours. The mixture was cooled to room temperature and poured into large amount of crushed ice with stirring. The resulting precipitate was filtered and washed with cold distilled water. The precipitate was air-dried and recrystallized from ethanol in cool condition. The scientific names and the respective R group for 2-8 are tabulated in Table 3.2. The condensed spectroscopy data for 2-8 can be found in Appendix I.

Table 3.2: Scientific name and respective R group of 2-8.

Compound	R group	Scientific name
2	C_2H_5	2-ethoxy-1-naphthaldehyde
3	C ₄ H ₉	2-butoxy-1-naphthaldehyde
4	C_6H_{13}	2-hexoxy-1-naphthaldehyde
5	C_8H_{17}	2-octoxy-1-naphthaldehyde
6	$C_{10}H_{21}$	2-decoxy-1-naphthaldehyde
7	$C_{12}H_{25}$	2-dodecoxy-1-naphthaldehyde
8	$C_{14}H_{29}$	2-tetradecoxy-1-naphthaldehyde

3.3.2 Preparation of 3-acetyl-4-hydroxycoumarin, 9

3-Acetyl-4-hydroxycoumarin was synthesized according to the synthetic scheme below (Figure 3.2).

Figure 3.2: Synthetic scheme for the formation of 3-acetyl-4-hydroxycoumarin, 9.

4-Hydroxycoumarin (3.00 g, 18.60 mmol) was added to acetic acid (16.00 mL), which served as a reactant and solvent. Phosphorylchloride (5.60 mL) was then added to the solution and the mixture was refluxed for 12 hours (solution color changed from yellow to intense red). The mixture was cooled to room temperature and poured into large amount of crushed ice with stirring. The resulting precipitate was filtered and washed with cold distilled water. The precipitate was air-dried and recrystallized from ethanol.

3.3.3 Preparation of 3-acetylcoumarin, 10

3-Acetylcoumarin was synthesized according to the synthetic scheme below (Figure 3.3).

Figure 3.3: Synthetic scheme for the formation of 3-acetylcoumarin, 10.

Salicylaldehyde (2.70 mL, 25.00 mmol), ethyl acetoacetate (3.20 mL, 25.00 mmol) and ethanol (50.00 mL) were all added into a 250 mL round-bottom flask. A catalytic amount of piperidine (0.50 mL) was added dropwise and the reaction mixture was refluxed at 65-75 °C for 4 hours. The mixture was cooled to room temperature. The resulting precipitate was filtered and washed with small portion of cold ethanol. The precipitate was air-dried and recrystallized from ethanol/chloroform mixture.

3.3.4 Preparation of 3-acetyl-6,7-dimethyl-4-hydroxycoumarin, 11

3-Acetyl-6,7-dimethyl-4-hydroxycoumarin was synthesized according to the synthetic scheme below (Figure 3.4).

Figure 3.4: Synthetic scheme for the formation of 3-acetyl-6,7-dimethyl-4-hydroxycoumarin, 11.